

Video Article

Design and Fabrication of an Optical Fiber Made of Water

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Abstract

In this report, an optical fiber of which the core is made solely of water, while the cladding is air, is designed and manufactured. In contrast with solid-cladding devices, capillary oscillations are not restricted, allowing the fiber walls to move and vibrate. The fiber is constructed by a high direct current (DC) voltage of several thousand volts (kV) between two water reservoirs that creates a floating water thread, known as a water bridge. Through the choice of micropipettes, it is possible to control the maximal diameter and length of the fiber. Optical fiber couplers, at both sides of the bridge, activate it as an optical waveguide, allowing researchers to monitor the water fiber capillary body waves through transmission modulation and, therefore, deducing changes in surface tension.

Co-confining two important wave types, capillary and electromagnetic, opens a new path of research in the interactions between light and liquid-wall devices. Water-walled microdevices are a million times softer than their solid counterparts, accordingly improving the response to minute forces.

Video Link

The video component of this article can be found at https://www.jove.com/video/58174/

Introduction

Since the breakthrough of optical fibers in communication, awarded with a Nobel prize in 2009¹, a series of fiber-based applications grew alongside. Nowadays, fibers are a necessity in laser surgeries², as well as in coherent X-ray generation^{3,4}, guided-sound⁵ and supercontinuum⁶. Naturally, the research on fiber optics expanded from utilizing solids into exploiting liquids for optical wave guiding, where liquid-filled microchannels and laminar flow combine the transportation properties of a liquid with the advantages of optical interrogation^{7,8,9}. However, these devices clamp the liquid between solids and, therefore, forbid it to express its own wave character, known as capillary wave.

Capillary waves, similar to those seen when throwing a stone into a pond, are an important wave in nature. However, due to the obstacles of controlling a liquid without dampening its surface through channels or solids, they are hardly utilized for detection or application. In contrast, the device presented in this protocol has no solid boundaries; it is surrounded by and flows in air, allowing, therefore, capillary waves to develop, propagate, and interact with light.

To fabricate a water fiber, it is necessary to go back to a technique known as the floating water bridge, first reported in 1893¹⁰, where two beakers filled with distilled water and connected to a high-voltage source will form a fluidic, water thread-like connection between them¹¹. Water bridges can reach up to a length of 3 cm¹² or be as thin as 20 nm¹³. As for the physical origin, it has been shown that surface tensions, as well as dielectric forces, are both responsible for carrying the bridge's weight^{14,15,16}. To activate the water bridge as a water fiber, we couple light in with an adiabatically tapered silica fiber^{17,18} and out with a silica fiber lens¹⁹. Such a device can host acoustical, capillary, and optical waves, making it advantageous for multi-wave detectors and lab-on-chip^{20,21,22} applications.

Protocol

CAUTION: This experiment involves high voltage. It is the reader's responsibility to verify with the safety authorities that their experiment follows regulations before turning on the high voltage.

NOTE: Any kind of polar liquid can be utilized to produce liquid fibers, such as ethanol, methanol, acetone, or water. The polarity of the liquid dictates the stability and diameter of the created fiber^{23,24}. For best results, use deionized water with 18 M Ω resistance. Before choosing optical

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components, such as optical fibers and light sources, consult the literature to ensure a low absorption in the water/liquid fiber at the desired optical wavelength. The protocol can be paused at any given moment before filling the water reservoir (step 4.5).

1. Preparation of Water Reservoirs and Experimental Station

- Manufacture two poly(methyl methacrylate) (PMMA) reservoirs with magnetic connectors for the pipette and the high voltage, according to Figure 1.
 - 1. Cut two PMMA plates to 60 x 50 x 10 mm in size, drill cavities of 8 mm in depth and 7 mm in diameter on the backside of the plates. Glue connector magnets inside the cavities.
 - 2. For the capillary clamp, cut a stripe of PMMA to 45 x 10 x 2 mm and glue two magnets on the top side of it.
 - 3. For the electric connector, wrap magnets in a small piece of metallic foil and connect it electrically with crocodile clamps to the high-voltage (HV) source. The reservoirs hold approximately 100 300 µL of water. Place the wrapped magnets in fluidic contact with the water in the reservoir.
 - NOTE: Preferably, use magnetic connectors for clamps and high voltage. If possible, do not to use any kind of glue to attach the clamps or the connectors, as many types of glues dissolve under the influence of high voltage or in the presence of electric arcs and diminish the water fiber stability or optical quality.
- 2. Mount one PMMA reservoir on a 5-degree-of-freedom (DOF) micro-positioning stage.
- 3. Thoroughly clean all connectors and areas with isopropanol (spectral grade) followed by deionized water. Blow-dry with nitrogen. Cover the PMMA water reservoir and all clamps with polytetrafluoroethylene (PTFE) tape to avoid any leaking or dripping of water.
- 4. Position the set-up under an optical microscope for imaging. Use far-field objectives (5X, 0.14 NA, and 34 mm WD for long water fibers and 20X, 0.42 NA, and 20 mm WD lens for short water fibers) to avoid unwanted grounding between the HV area of the water fiber and the electrically conductive microscope set-up.
- 5. Set up two optical fiber clamps on linear transitional stages, one behind each water reservoir, according to **Figure 1**. Each fiber coupler should be able to move back and forward within its micropipette (discussed in the following section).

2. Choosing the Micropipettes and Voltage

- The inner diameter of the micropipette ensures a maximum radius of the fabricated water fiber. To create a 5-μm radius water fiber, use 150-μm-inner-diameter pipettes, paired with 125-μm-diameter optical fibers. For thicker (20 90 μm) and longer (800 1,000 μm) water fibers, use micropipettes with an inner diameter of 850 μm.
 - NOTE: As a rule of thumb, the water fiber maximum length is estimated by multiplying the maximal radius by 25. For details, refer to Table 1.
 - 1. Break the micropipette by hand over an edge to a length of 3 cm.
- 2. To create water fibers with a diameter of up to 110 µm, apply a voltage between the two water reservoirs between 1.5 kV and 3 kV. For water fibers reaching up to a millimeter in length, apply up to 8 kV. Compare with **Figure 1** for electrical wiring suggestions.

3. Preparation of the Optical Couplers

NOTE: For the best transmission result, use a single-mode tapered fiber to launch laser light into the water fiber and a highly multimode reflowed fiber lens as the output coupler (core > $100 \mu m$). However, for easy operation, use a low multimode fiber as the output coupler (for example, a 1550-nm single-mode fiber for a 780-nm wavelength).

1. Fabrication of a Tapered Fiber Coupler

NOTE: See Figure 2.

- 1. Strip the 780-nm single-mode fiber with a fiber stripper from its plastic cladding to expose an area of 10 15 mm of bare fiber. Clean the exposed area with delicate task wipes in combination with acetone. Pass the fiber through the desired micropipette before tapering it. Taper the fiber below the single-mode criteria with a slope smaller than 1/20.
- 2. Use a hydrogen flame for tapering the fiber with a flow rate of 140 mL/min, while simultaneously pulling the taper from both sides at 0.06 mm/s.
 - NOTE: The tapered portion is in total between 6 to 9 mm. If the fiber breaks before reaching the single-mode criteria, adjust the hydrogen flow to higher rates or place the fiber in a hotter area of the torch. If the area is longer, adjust the hydrogen flow to lower rates or place the fiber in a colder area of the torch.
- Turn off the flame and carefully increase the tension in the fiber until it breaks at its thinnest spot. Use this tapered fiber as the input coupler.
 - CAUTION: The tapered fiber is fragile.

2. Fabrication of a Fiber Lens Coupler

- 1. Strip the 1550-nm single-mode fiber tip with a fiber stripper and clean the exposed area with delicate task wipes in combination with acetone. Choose and prepare a pipette as described above and pass the fiber through it.
- 2. Heat the tip with an electric fusion splicer or CO₂ laser at 15-W power, focused through a 200-mm lens, until the glass fiber end becomes liquid and forms a slightly rounded shape, known as a fiber lens.

4. Assembling

1. If not done yet, insert the fiber couplers into the desired micropipettes.



- 2. Clamp the micropipette, using the premanufactured, magnetic PMMA clamp, with the fiber couplers onto the PMMA reservoirs. The non-tapered side of the micropipettes should reach into the water reservoir. Clamp each of the fiber couplers on a linear positioning stage.
- 3. Connect the tapered fiber coupler to a 780-nm, continuous wave, fiber-coupled 10-mW laser source and the fiber lens coupler to a power meter. Fill the reservoir with water and ensure that no air bubbles are stuck in the micropipette. If necessary, push or pull them with the optical fiber coupler (from step 3.1 or, accordingly, from step 3.2).
 - NOTE: At this stage, following the optical path, the stations are: the laser light source, the optical fiber, (and this fiber goes through) the fiber clamp on a linear stage, the water in the reservoir with electrical connection, the micropipette filled with water, the optical tapered fiber coupler, free space (later: water fiber), the fiber lens coupler (now the second fiber), the micropipette filled with water, the water reservoir with electric connection, the fiber clamp on a linear stage, and, lastly, the power meter.
- 4. Connect the ends of the mounted micropipettes by adjusting the 5-degre-of-freedom mount of the PMMA water reservoir to establish a fluidic contact between the micropipettes. Turn on the light source and the power meter. Adjust the fiber couplers to have a transmission with the help of the 5-DOF PMMA water reservoir mount.
 NOTE: Use appropriate laser safety equipment.
- 5. Connect the high voltage electrically with the water reservoir by placing the magnetic connectors wrapped in metallic foil over the magnetic counterparts in the PMMA water reservoir and attaching crocodile clamps to the metallic foil. Connect the crocodile clamps *via* electrical cables to the HV source (**Figure 2a**).

5. Running the Experiment

- Increase the voltage to the desired value. A starting point for a very short and narrow bridge is 1.5 kV. Stable bridges with 100 μm and more
 in length can be achieved with 2.5 3 kV.
- Slowly increase the distance between the micropipettes to the desired length according to the choice of the micropipettes (Figure 2b and 2c).
 Adjust couplers and pipettes with the 5-DOF stage and the 1-DOF stages to optimize the optical transmission.
- 3. Measure the coupling efficiency by taking a measurement on the power meter and taking the ratio of the coupled-in to the coupled-out laser power.
- 4. Disconnect the power meter and connect a photoreceiver to the output fiber coupler. Connect the photoreceiver to an oscilloscope. Record time trace measurements of the transmitted light, representing the capillary water fiber oscillations.
- Convert the time trace measurements via Fast Fourier Transformation to frequency domain. Take the central frequency over full width at half maximum to receive the capillary quality factor.
 NOTE: Create a spectrogram to check for frequency jitter.
- 6. Use the top view microscope set-up to characterize the geometrical structure of the water fiber. The fiber radius is obtained at the thinnest portion of the water fiber.

Representative Results

The coupling efficiency from a water fiber to a highly multimode fiber can be as high as $54\%^{25,26}$. The coupling efficiency to a single-mode fiber is up to $12\%^{25,26}$. Water fibers can be as thin as 1.6 µm in diameter and can have a length of 46 µm (**Figure 3**) 25,26 , or they can be up to 1.064 mm in length with a diameter of 41 µm (**Figure 3**) 25,26 . The transmission spectrogram reveals capillary oscillation of the water fiber, similar to that of a guitar string (**Figure 4**) 25,26 . The capillary quality factors were estimated to be as high as 14 for long fibers 25,26 . Considering the theory on water bridges, it is possible to estimate the ratio between the surface tension and the dielectric force 25,26 .

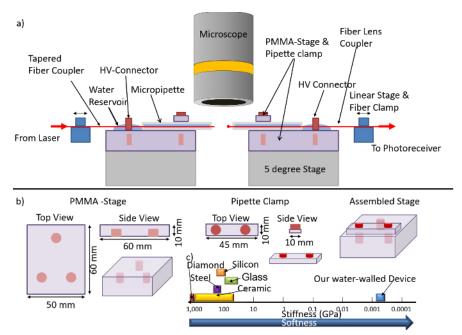


Figure 1: Schematics of the set-up. (a) This illustration shows the water fiber experimental set-up. (b) This sketch shows the water reservoir, the electrical connector, and the pipette clamp. (c) This panel shows the water-walled waveguide softness compared with common solids. This figure is reproduced in part from Douvidzon *et al.*^{25,26}. Please click here to view a larger version of this figure.

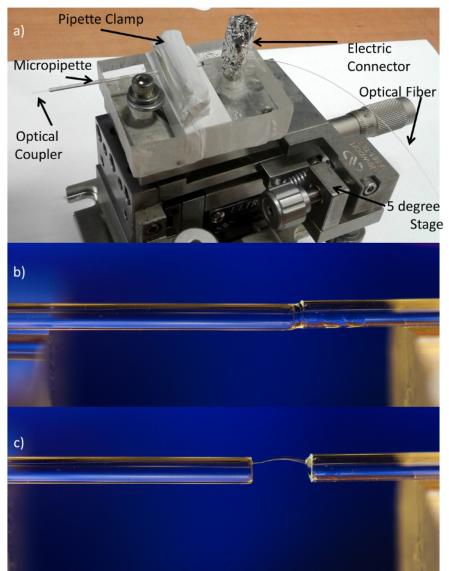


Figure 2: Set-up photos. (a) This panel shows the PMMA-water reservoir on a 5-DOF mount. with the PMMA-pipette clamp, the micropipette, the optical fiber, and the electric connector. (b) This panel shows that a fluidic contact between the micropipettes is created. (c) This panel shows that the distance between the micropipettes is increased to establish a water fiber. Please click here to view a larger version of this figure.

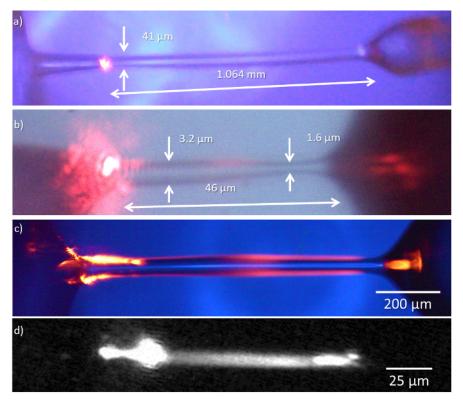


Figure 3: Water fiber characterization. (a) This panel shows a water fiber longer than 1 mm. The next two panels show (b) a micron-scale-thin water fiber, (c) the surface scattering due to capillary waves at the water fiber liquid-phase boundary. (d) This panel shows light propagation through the water fiber volume confirmed by a fluorescent dye measurement. This figure is reproduced from Douvidzon *et al.*^{25,26}. Please click here to view a larger version of this figure.

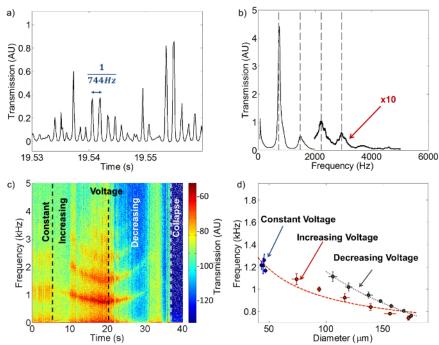


Figure 4: Experimentally measuring the water fiber "guitar-string" modes. (a) This panel shows a time trace measurement. (b) A fluctuation spectrum reveals a fundamental mode and, at integer multiplications, its three overtones (dash lines). (c) This panel shows a fluctuation spectrogram of a 0.94-mm-long fiber with changing voltage and, correspondingly, changing the fiber diameter, with voltage first constant, then increased, and finally, decreased. The color code describes the transmission. (d) This panel shows the fundamental frequency of the fiber as a function of the fiber diameter (circles) together with a theoretical prediction (dashed line). Horizontal and vertical error bars represent the uncertainty of eight consecutive, 250-ms-apart measurements of the central frequency and its corresponding fiber diameter. For all panels, the fiber length is 0.94 mm and the oscillation is optically interrogated with a photodetector. The diameter is measured *via* microscope. This figure is reproduced from Douvidzon *et al.*^{25,26}. Please click here to view a larger version of this figure.

	Water fiber			Pipette's internal diameter	
	Length [µm]	Radius [µm]	Potential [V]	Taper side [µm]	Lens side [µm]
Fig. 1b	830	51	6000	850	850
Fig. 2a	1064	20.5	6000	850	850
Fig. 2b	46	1.6 - 0.8	1500	150	850
Fig. 2c	820	32.5	5000	850	850
Fig. 2d	110	4.75	3000	150	150
Fig. 3	940	20 - 90	3000 - 8000	850	850
Fig. 4	24 - 73	2.7 - 3	2500	150	850

Table 1: Water fiber length and radius. This table shows the water fiber length and radius with respect to the electric potential and the pipette diameter. This table is reproduced from Douvidzon, *et al.*²⁵.

Discussion

To conclude, the major advantage and uniqueness of this technique is creating a fiber which hosts three different kinds of waves: capillary, acoustic, and optical. All three waves oscillate in different regimes, opening the possibility for multi-wave detectors. As an example, airborne nanoparticles affect the surface tension of liquids. Already at the current stage, it is possible to monitor changes in the surface tension through variations in the capillary eigenfrequency. Additionally, water-walled devices are a million times softer than their solid counterparts, improving the sensibility of sensors accordingly.

Based on experience with this set-up, we noticed a high dependency on the signal-to-noise ratio and the quality of the optical couplers. Therefore, it is recommended to pay close attention to the fabrication of the optical couplers. Consider an aquarium set-up to ensure a dust-free environment for the tapering station and the water fiber set-up. Also, the execution of the experiment involves a risk of breaking or damaging the tapered fiber coupler mechanically or through an electric arc. In that case, the optical transmission can drop and become noisy to such an extent that the capillary modes of the fiber are no longer visible in the spectrogram.

If capillary waves are not visible in the transmission measurements, remanufacture the couplers. Additionally, the water fiber and the optical fiber couplers do not attract each other. Adjusting the set-up for optimal transmission may require putting the water fiber a bit askew, to mechanically press the tapered fiber coupler inside the water fiber.

Another obstacle in this set-up to be aware of is the crucial electrical resistivity of the water. Even small amounts of ions in the liquid will cause the bridge to collapse. If the water fiber is shorter and less stable than expected, a contamination of the water might be the cause. Replace the water with $18~\mathrm{M}\Omega$ clean room water. Additionally, the high voltage attracts charged air particles in the surrounding of the water fiber, which dissolve and contribute to the instability. In this case, a closed chamber will help improve the water fiber longevity.

An outstanding aspect of this set-up is that any polar liquid can be utilized to create a liquid fiber, although deionized water is known for creating the longest, as well as, time-wise, the most stable water fibers. It is interesting to consider other liquids for different applications. Switching the water to a liquid or a mix of polar liquids with fitting viscosity, surface tension, or optical properties allows researchers to trim the fiber exactly to their demands.

Disclosures

The authors have nothing to disclose.

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References

- 1. Kao, C. K., Boyle, W. S., Smith, G. E. 'For Groundbreaking Achievements Concerning the Transmission of Light in Fibers for Optical Communication'. *The Nobel Prize in Physics*. https://www.nobelprize.org/nobel_prizes/physics/laureates/2009/press.html (2009).
- 2. Temelkuran, B., Hart, S. D., Benoit, G., Joannopoulos, J. D., Fink, Y. Wavelength-scalable hollow optical fibres with large photonic bandgaps for CO2 laser transmission. *Nature*. **420**, 650-653 (2002).
- 3. Rundquist, A. Phase-Matched Generation of Coherent Soft X-rays. Science. 280, 1412-1415 (1998).
- 4. Durfee, C. G. et al. Phase Matching of High-Order Harmonics in Hollow Waveguides. Physical Review Letters. 83, 2187-2190 (1999).
- 5. Dainese, P. et al. Stimulated Brillouin scattering from multi-GHz-guided acoustic phonons in nanostructured photonic crystal fibres. *Nature Physics.* **2**, 388-392 (2006).
- 6. Dudley, J. M. J., Genty, G., Coen, S. Supercontinuum generation in photonic crystal fiber. Reviews of Modern Physics. 78, 1135-1184 (2006).
- Wolfe, D. B. et al. Dynamic control of liquid-core/Liquid-cladding optical waveguides. Proceedings of the National Academy of Sciences of the United States of America. 101, 12434-12438 (2004).
- 8. Whitesides, G. M. The origins and the future of microfluidics. Nature. 442, 368-373 (2006).
- Ward, J. M., Yang, Y., Chormaic, S. N. Highly Sensitive Temperature Measurements With Liquid-Core Microbubble Resonators. IEEE Photonics Technology Letters. 25, 2350-2353 (2013).
- Armstrong. The Newcastle Literary and Philosophical Society. The Electrical Engineer. 154-155. http://ecfuchs.com/waterbridge_jw/ Armstron full article.pdf. January 19, 2016 (1893).
- 11. Fuchs, E. E. C. et al. The floating water bridge. Journal of Physics D: Applied Physics. 40, 6112-6114 (2007).
- 12. Fuchs, E. C. et al. The Armstrong experiment revisited. The European Physics Journal Special Topics. 223, 959-977 (2013).
- 13. Sirghi, L., Szoszkiewicz, R., Riedo, E. Volume of a nanoscale water bridge. Langmuir. 22, 1093-1098 (2006).
- 14. Woisetschläger, J., Gatterer, K., Fuchs, E. C. Experiments in a floating water bridge. Experiments in Fluids. 48, 121-131 (2009).
- 15. Widom, A., Swain, J., Silverberg, J., Sivasubramanian, S., Srivastava, Y. N. Theory of the Maxwell pressure tensor and the tension in a water bridge. *Physical Review E: Statistical, Nonlinear, and Soft Matter Physics.* **80**, 16301 (2009).
- 16. Aerov, A. A. Why the water bridge does not collapse. Physical Review E: Statistical, Nonlinear, and Soft Matter Physics. 84, 36314 (2011).
- 17. Knight, J. C., Cheung, G., Jacques, F., Birks, T. A. Phase-matched excitation of whispering-gallery-mode resonances by a fiber taper. *Optics Letters*. **22**, 1129 (1997).
- 18. Spillane, S. M., Kippenberg, T. J., Painter, O. J., Vahala, K. J. Ideality in a Fiber-Taper-Coupled Microresonator System for Application to Cavity Quantum Electrodynamics. *Physical Review Letters*. **91**, 43902 (2003).
- 19. Cohen, L. G., Schneider, M. V. Microlenses for coupling junction lasers to optical fibers. Applied Optics. 13, 89-94 (1974).
- 20. Vollmer, F. et al. Whispering-gallery-mode biosensing: label-free detection down to single molecules. Nature Methods. 5, 591-596 (2008).
- 21. Fainman, Y., Lee, L. P., Psaltis, D., Yang, C. Optofluidics: Fundamentals, Devices, and Applications. McGraw-Hill, Inc. (2010).
- 22. He, L., Ozdemir, S. K., Zhu, J., Kim, W., Yang, L. Detecting single viruses and nanoparticles using whispering gallery microlasers. *Nature Nanotechnology*. **6**, 428-432 (2011).
- 23. Woisetschläger, J. et al. Horizontal bridges in polar dielectric liquids. Experiments in Fluids. 52, 193-205 (2011).
- 24. Fuchs, E. C., Wexler, A. D., Agostinho, L. L. F., Ramek, M., Woisetschläger, J. Methanol, Ethanol and Propanol in EHD liquid bridging. *Journal of Physics: Conference Series.* **329**, 12003 (2011).
- 25. Douvidzon, M. L., Maayani, S., Martin, L. L., Carmon, T. Light and Capillary Waves Propagation in Water Fibers. Science Reports. 7, 16633 (2017).
- 26. Douvidzon, M., Maayani, S., Martin, L. L., Carmon, T. Water Fibers. Cornell University Library. https://arxiv.org/abs/1609.03362 (2016).