Video Article

Erratum: Reconstitution Of β-catenin Degradation In *Xenopus* Egg Extract

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Abstract

A correction was made to Reconstitution Of β-catenin Degradation In *Xenopus* Egg Extract. At the time of publication there were some instances where an incorrect volume notation was used. These instances were corrected from:

2.1.2. Add extract to 1/10 the volume of pelleted antibody or affinity beads (*e.g.*, 20 ml pelleted beads to 200 ml extract). In order to minimize dilution of the extract, withdraw as much liquid from the beads as possible before addition of the extract using gel loading tips with long, tapered tips.

2.2.5. Aliquot the appropriate volumes for degradation assay into pre-chilled microfuge tubes on ice. For radiolabeled β-catenin degradation assays, withdraw 2-5 ml extract for each time point.

3.2.3. At the designated time point, remove 1-5 ml of the sample and mix immediately with SDS sample buffer (5x volume) to stop the reaction. To make sure the degradation reaction is completely terminated, flick tube several times and vortex vigorously.

3.2.4. Perform SDS-PAGE/autoradiography. Run 1 ml equivalents (~50 mg of protein) of the extract for each time point/lane. Degradation of β-catenin in *Xenopus* egg extract should be evidenced by the time-dependent decrease in intensity of the radiolabeled β-catenin band **Figure 2**. Quantify results using ImageJ, ImageQuant, or other preferred imaging software if necessary.

4.2.2. Add *in vitro*-translated β-catenin-luciferase fusion (from 4.1) into prepared *Xenopus* reaction mix (from 2.2) on ice and mix well as in 3.2.1. NOTE: The activity of the β-catenin luciferase that is added to the extract is typically between 20 - 50,000 relative luminescence units (RLU)/ml of extract (based on measurements obtained from 4.1.2). Starting signal should be approximately 100,000 RLU (2-5 ml of the *in vitro*-translated β-catenin-luciferase fusion).

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Disclosures

No conflicts of interest declared.