

Video Article

# Controlled-release of Chlorine Dioxide in a Perforated Packaging System to Extend the Storage Life and Improve the Safety of Grape Tomatoes

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## Abstract

A controlled-release chlorine dioxide (ClO<sub>2</sub>) pouch was developed by sealing a slurry form of ClO<sub>2</sub> into semipermeable polymer film; the release properties of the pouch were monitored in containers with or without fruit. The pouch was affixed to the inside of a perforated clamshell containing grape tomatoes, and the effect on microbial population, firmness, and weight loss was evaluated during a 14 day storage period at 20 °C. Within 3 days, the ClO<sub>2</sub> concentration in the clamshells reached 3.5 ppm and remained constant until day 10. Thereafter, it decreased to 2 ppm by day 14. The ClO<sub>2</sub> pouch exhibited strong antimicrobial activity, reducing *Escherichia coli* populations by 3.08 log CFU/g and *Alternaria alternata* populations by 2.85 log CFU/g after 14 days of storage. The ClO<sub>2</sub> treatment also reduced softening and weight loss and extended the overall shelf life of the tomatoes. Our results suggest that ClO<sub>2</sub> treatment is useful for extending the shelf life and improving the microbial safety of tomatoes during storage without impairing their quality.

## Video Link

The video component of this article can be found at <https://www.jove.com/video/55400/>

## Introduction

A diet rich in fresh fruits and vegetables may help to reduce the risk of many diseases, including coronary heart disease and specific types of cancers<sup>1</sup>. However, there are a number of foodborne microbial pathogens, such as *Escherichia coli*, *Salmonella enterica*, and *Listeria monocytogenes*, associated with the consumption of fresh fruits and vegetables that can cause illness or even death among consumers who eat contaminated produce<sup>2</sup>. For example, *E. coli* O157:H7 outbreaks have been associated with grapes, tomatoes, and strawberries<sup>3,4</sup>, and hepatitis A outbreaks have been associated with fresh blueberries<sup>5</sup>. In addition, microbial contamination can cause substantial product loss through postharvest decay<sup>6</sup>. *Alternaria alternata* is an important plant pathogenic fungus that is known to cause leaf spots and other diseases in over 380 host species of plants<sup>7</sup>. It has been shown to be the cause of an *Alternaria* black spot<sup>8</sup>, a stem canker disease and a leaf blight of tomatoes<sup>9</sup>. Therefore, a safe and effective postharvest decontamination treatment is needed to both control foodborne pathogens and to prevent postharvest decay in fresh produce.

Low- and non-residue technologies are new trends for alternative sanitizers. A variety of postharvest fungicides have been used to reduce spoilage organisms and to prevent foodborne illness. Ozone, a strong antimicrobial agent, has been shown to preserve the quality and freshness of strawberries and blueberries<sup>10,11</sup>. However, ozone may cause oxidation of fruit surface tissue and can result in discoloration and the deterioration of flavor quality<sup>12</sup>. Chlorine has been used to sanitize fresh produce, such as blueberries and apples<sup>13</sup>. While effective, chlorine can react with nitrogen-containing compounds or ammonia, resulting in carcinogenic byproducts<sup>14</sup>, especially when used for the sanitization of fresh fruit<sup>15</sup>.

Chlorine dioxide (ClO<sub>2</sub>), an alternative sanitizer, was approved by both China and the US for the postharvest treatment of fruits and vegetables<sup>16</sup>. ClO<sub>2</sub> is a water-soluble oxidizing agent with an oxidation capacity 2.5 times greater than that of free chlorine<sup>17</sup>. ClO<sub>2</sub> is highly effective at low concentrations and with a short contact time<sup>18</sup>. ClO<sub>2</sub> has low toxicity and minimal corrosiveness at the concentrations used for disinfection, and it is recognized as one of the most effective bactericidal and fungicidal agents for use in a variety of settings<sup>19,20,21</sup>.

Numerous research results have shown that ClO<sub>2</sub> can control foodborne pathogens and postharvest decay<sup>16</sup>. For example, ClO<sub>2</sub> gas has been used to inactivate *L. monocytogenes*, *Salmonella*, and *E. coli* O157:H7 and to prevent blueberry and strawberry spoilage<sup>22,23</sup>. ClO<sub>2</sub> gas reduces the risk of microbial contamination while maintaining the attributes of fresh fruit, and it was effective at controlling the postharvest decay of

strawberries<sup>24</sup>. However, it is unstable at high concentrations and non-transportable, historically requiring costly generators on site or inefficient two-part powder mixing.

However, a new ClO<sub>2</sub> product with a ready-made, controlled-release formulation (*i.e.*, it does not require a generator or the premixing of ingredients) has been shown to be highly effective at controlling food spoilage organisms and pathogens in preliminary experiments<sup>25</sup>. It is a safe, cost-effective, non-corrosive, easily transportable, and controlled-release form of ClO<sub>2</sub>, with no adverse effects on the environment. Previous experiments have demonstrated that this slow-release ClO<sub>2</sub> powder wrapped in filtration material and placed in clamshell packaging significantly reduced the decay of fresh blueberries and strawberries, decreased berry water loss, and maintained fruit firmness during postharvest storage<sup>25,26</sup>. Recently, a controlled-release ClO<sub>2</sub> packet was developed by sealing a slurry form of ClO<sub>2</sub> in a semipermeable polymer film. The objectives of this work were to: 1) monitor ClO<sub>2</sub> gas release properties in both a closed container and in perforated clamshells, 2) investigate the effect of a controlled-release ClO<sub>2</sub> pouch enclosed in a container on foodborne pathogens and the decay of grape tomatoes, and 3) evaluate the effects of the controlled-release ClO<sub>2</sub> on the storage quality of grape tomatoes.

## Protocol

### 1. Measurement of Gaseous ClO<sub>2</sub> in the Headspace of a Closed Chamber

1. Obtain the materials: ClO<sub>2</sub> pouch (0.5 g of ClO<sub>2</sub> slurry (9.5% a.i.) in a polymer film selected for its release rate (total surface area of 6 cm<sup>2</sup>); the exact components are proprietary), a glass chamber (19.14 L), and a lid with switchable gas inlet and outlet.
2. Attach the ClO<sub>2</sub> pouch to the lid using double-sided tape.
3. Close the chamber by sealing the lid with petroleum jelly.
4. Connect the inlet and outlet of a ClO<sub>2</sub> gas detector to the chamber.  
NOTE: This is a gas circulation system, and no gas loss occurred when taking measurements.
5. Switch on the inlet and outlet gas flow and measure the ClO<sub>2</sub> concentration in the chamber after incubating for 0, 1, 2, 3, 4, 24, 26, 28, and 48 h.
6. Monitor the temperature and relative humidity (RH) in the chamber with temperature and RH data loggers.

### 2. Fruit Preparation and Storage

1. Obtain 15 kg of fresh grape tomatoes (*Solanum lycopersicum* var. *cerasiforme*) from a local retailer. Ensure that the fruits are healthy and have no visual flaws.
2. **Preparation of inoculum**
  1. Use strains of *E. coli* (wild type) and *A. alternata* from citrus fruit surfaces<sup>27</sup> for inoculation.
  2. Culture *E. coli* on *E. coli* agar (ECA) at 35 °C for 1 day<sup>27</sup> and then re-culture the organisms on a new plate for 1 day. Confirm the organisms by sampling the ECA plates with a bac-loop, streaking the bacteria on Levine eosin methylene blue (EMB) agar, and incubating for 24 h at 35 °C; cultures that turn reflective, metallic green are positive for *E. coli*.
  3. Culture *A. alternata* on potato dextrose agar (PDA) at 25 °C until spores appear.
  4. Scrape the *E. coli* cells from the agar plate into 50 mL of sterile distilled water until the estimated concentration reaches 9 log CFU/mL using a comparison with McFarland equivalence turbidity standards. Add 1,950 mL of sterile water containing 0.1% Tween-20 to make 2 L total of the final inoculum.
  5. Verify the cell concentration by dilution plating on EC agar plates. Scrape the *A. alternata* spores from the culture medium and suspend them into 2 L of sterile distilled water containing 0.1% Tween-20.  
NOTE: The final *E. coli* population was 7.5 log CFU/g, and the *A. alternata* population was 5.5 log CFU/g.
3. **Place 7 kg of the tomatoes into a 10 L stainless steel pan that is completely covered by an autoclavable bag. Place the bag and pan in a safety hood. Apply the inoculum solution (2 L) to the fruits using a trigger sprayer applied from the top while gently stirring the fruits with a gloved hand.**
  1. After 5 min, place the tomatoes in a single layer on sterilized sheets and allow them to air dry for 2 h. Put about 200 g of fruit each into twenty-four 1 lb (~1.14 L) perforated clamshells.
4. Carefully fold the contaminated foils and place them into the steel pan. Remove the gloves and put them into the pan. Wrap the autoclavable bag and autoclave all contaminated supplies at 121 °C for 25 min.
5. Attach ClO<sub>2</sub> pouches to the lids of 12 clamshells. Use the other 12 clamshells as controls. Weigh each whole clamshell. Store the fruit at 20 °C for 14 days.
6. Take samples on days 3, 7, 10, and 14. Sample three clamshells, representing 3 replicates, per treatment per day.

### 3. Monitoring of ClO<sub>2</sub> Concentration in the Clamshells

1. Insert the inlet and outlet tubing of the ClO<sub>2</sub> gas detector into the center of the clamshells, with a 2 cm distance between the two ends, and take the ClO<sub>2</sub> measurement on days 3, 7, 10, and 14.

### 4. Determination of Microbial Population and Fruit Quality Attributes

1. **Agitate 5 fruits (about 60 g) from each replicate at 100 rpm for 1 h in a sterilized sampling bag along with 99 mL of sterile potassium phosphate buffer (0.01 M, pH 7.2) on an orbital shaker.**
  1. Plate serial dilutions (1-, 10-, and 100-fold) of the buffer wash, 50 µL each, on ECA (for *E. coli*) and PDA (for *A. alternata*) using a spiral plater.

2. Incubate the ECA plates at 35 °C for 24 h and the PDA plates at 25 °C for 3 days. Read the microbial colony count using an optical plate reader. Sanitize all equipment which contacted the contaminated fruit after use.
2. Measure fruit firmness with a fruit firmness tester using the manufacturer's protocol. Calibrate the tester before each use. Measure 20 fruit for each replicate and express the results as the pressure force, Newton (N), required to compress the fruit by 1 mm (converted to  $\text{N} \cdot \text{m}^{-1}$ ).
3. Weigh the whole clamshell with the fruit at the beginning of and during storage and calculate the weight loss in comparison to the initial weight.

## 5. Statistical Analysis

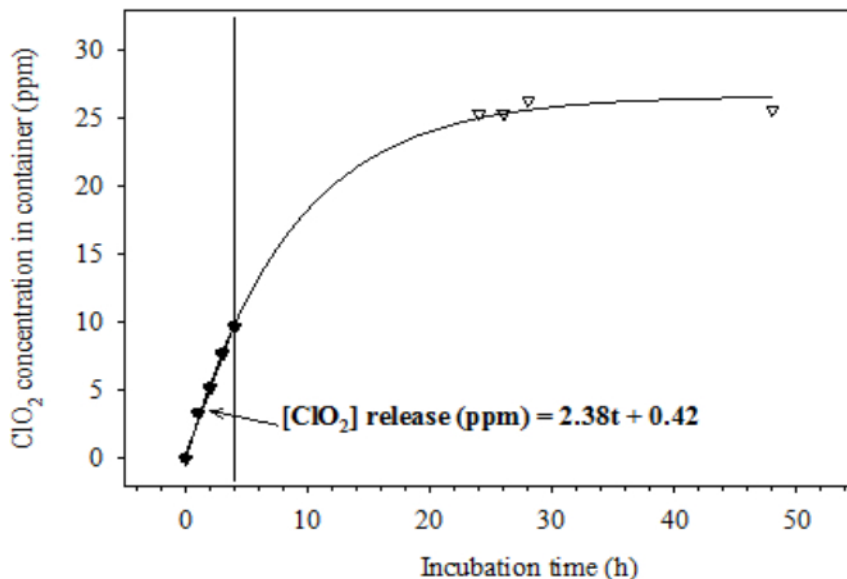
1. Replicate all experiments in triplicate. Analyze the data using analysis of variance (ANOVA). Determine the mean separation by Duncan's multiple range test; the significance is defined at  $p < 0.05$ .

## Representative Results

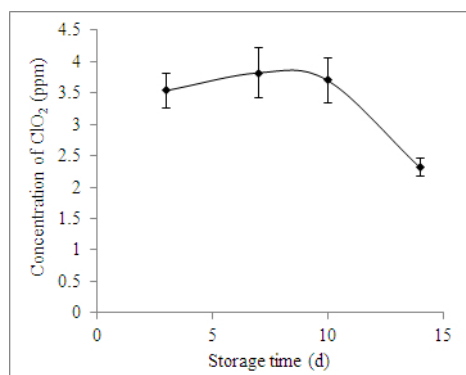
The release of  $\text{ClO}_2$  exhibited a linear pattern over the first few hours. The concentration increased about 2.38 ppm/h over the first 4 h. The release speed slowed after 24 h of incubation, and the  $\text{ClO}_2$  concentration reached 25.4 ppm. However, the concentration tended to be stable after 24 h of incubation (**Figure 1**).

The headspace  $\text{ClO}_2$  concentration in the clamshell with grape tomatoes was about 4 ppm between day 3 and day 10, it decreased after 10 days of storage, and it was about 2 ppm on day 14 (**Figure 2**). The initial populations of *E. coli* and *A. alternata* in the fruit after inoculation were 4.3 and 3.4 log CFU/g, respectively (**Figure 3**). Treatment with  $\text{ClO}_2$  pouches reduced the populations of *E. coli* and *A. alternata* by 3.08 and 2.85 log CFU/g, respectively, after 14 days of storage (**Figure 3**).

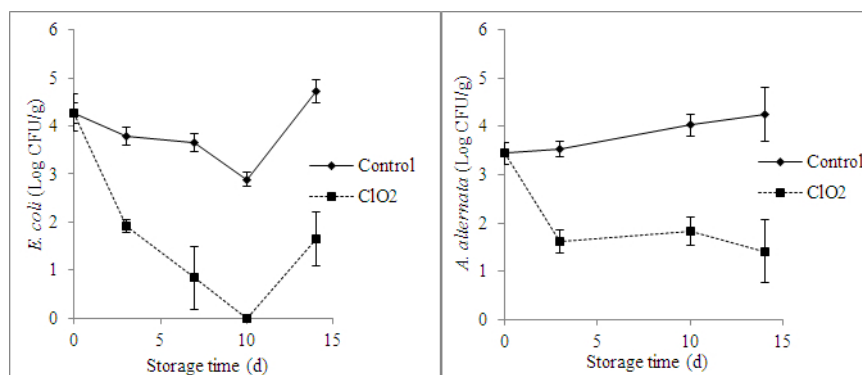
The effects of  $\text{ClO}_2$  treatment on fruit firmness and weight loss are presented in **Figures 4** and **5**.  $\text{ClO}_2$  prevented a loss of firmness and weight in the fruit, and these effects grew with extended storage time (**Figures 4** and **5**).



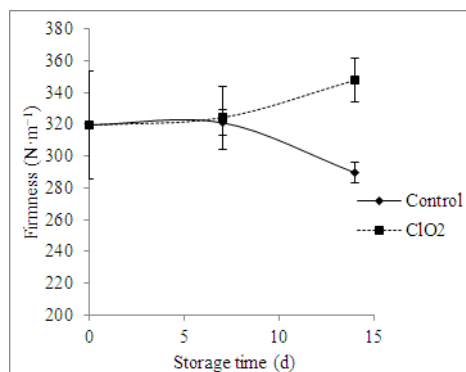
**Figure 1:**  $\text{ClO}_2$  release profile of a 0.5-g  $\text{ClO}_2$  pouch in a sealed, empty 19.14-L glass container at 20 °C and relative humidity 91%.



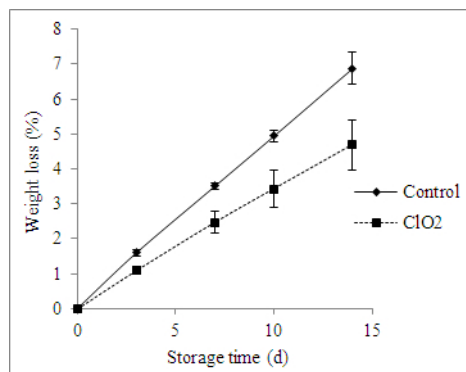
**Figure 2:** Concentration of ClO<sub>2</sub> in 1 lb perforated clamshell packaging with 200 g of grape tomatoes at 20 °C. The values are the mean ± SD.



**Figure 3:** Effect of ClO<sub>2</sub> treatment on *E. coli* and *A. alternata* populations on the surfaces of inoculated grape tomatoes stored for 14 days at 20 °C. The values are the mean ± SD.



**Figure 4:** Effect of ClO<sub>2</sub> treatment on the firmness of grape tomatoes stored for 14 days at 20 °C. The values are the mean ± SD.



**Figure 5:** Effect of ClO<sub>2</sub> treatment on the weight loss of grape tomatoes stored for 14 days at 20 °C. The values are the mean ± SD.

## Discussion

Chlorine dioxide is an ideal biocide to prevent food decay. However, it is unstable at high concentrations and non-transportable, requiring costly generators or inefficient two-part powder mixing. This study examined the application of a stable, ready-to-use form of chlorine dioxide to reduce food spoilage and the incidence of foodborne illness. In contrast to other chlorine dioxide application technologies currently in use, the commercial ClO<sub>2</sub> used here is cost effective, has a long shelf life, and does not require large generators or premixing. However, due to the strong oxidative properties of ClO<sub>2</sub>, the gas release properties of ClO<sub>2</sub> are difficult to measure and therefore are rarely reported. In a previous study, a titration method was used to measure the release rate<sup>28</sup>. However, this method is less accurate and more complicated. Some research evaluated the concentration of ClO<sub>2</sub> by absorbing it in water and then measured it using gas chromatography with mass spectrometric (GC-MS) detection<sup>29</sup>. However, this GC-MS instrument is complicated and expensive<sup>30</sup>. In our research, a ClO<sub>2</sub> gas detector was used to measure the concentration of ClO<sub>2</sub>. This detector has multiple sensors that provide more accurate results in a shorter time.

In our protocol, for the preparation of the inoculum, the use of a deep-sided, 10 L steel pan as a basin for the application of inoculum, itself placed within an autoclavable bag, as well as sterile foil on which to dry the fruit, allows for quick clean-up and helps to avoid human exposure to possibly pathogenic organisms through incidental contact. Spraying the fruit within the confines of the autoclavable bag reduced the dispersion of microbial aerosols. Drying the fruit on foil allowed for complete removal and the subsequent sterilization of all surfaces with which the contaminated fruit had come into contact.

Chlorine dioxide exhibited strong antimicrobial activity against *E. coli* and *A. alternata* in grape tomatoes (**Figure 3**). ClO<sub>2</sub> solution has been used to wash fruits and vegetables. Treatment with ClO<sub>2</sub> gas at 4.1 mg/L (1,484 ppm) for 20 min at 23 °C significantly reduced the population of *Salmonella*, *E. coli* O157:H7, and *L. monocytogenes* on fresh-cut lettuce, cabbage, and carrots, without causing adverse effects on sensory properties<sup>31</sup>. Higher than 3-log reductions of *E. coli* O157:H7 were achieved after 4 mg/L (1,448 ppm) ClO<sub>2</sub> gas treatments for 10 min at 21 °C and 90% RH on apple surfaces<sup>32</sup>. The effects of ClO<sub>2</sub> treatment on fruit firmness and weight loss are presented in **Figures 4 and 5**. The firmness of ClO<sub>2</sub>-treated tomatoes increased compared to the control fruit (**Figure 4**). ClO<sub>2</sub>-treated fruit demonstrated inhibited enzyme activity, including in peroxidase and polyphenol oxidase, which was attributed to an important role in the softening process<sup>33</sup>, or inhibited respiration rates and ethylene production<sup>34,35</sup>. A linear relationship between softening and weight loss was demonstrated in blueberries<sup>36</sup>. It was suggested that ClO<sub>2</sub> could reduce fruit metabolism in addition to preventing weight loss and retaining firmness<sup>37</sup>. It was concluded that the ClO<sub>2</sub> pouch was a promising, non-thermal, pathogen-reduction technique for fresh fruits and vegetables. It maintained firmness and reduced the weight loss of grape tomatoes.

One limiting characteristic of this method of sanitation is that although this ClO<sub>2</sub> technology may reduce the tomato surface inoculum of *A. alternata*, reducing the risk of new postharvest infections by this fungus, it will not be able to control established, latent infections of *A. alternata*<sup>38</sup>. Established infections are typically produced in the field before harvest and are the main cause of postharvest tomato black spots, which cause significant economic losses to the industry. Another limiting characteristic is the rapid reaction of ClO<sub>2</sub>, which prevents the product from effectively combating microorganisms that are deeply embedded within a water-rich environment or dense organic material<sup>39</sup>. Typically, the sanitizing potential of the product at low concentrations quickly loses effectiveness before being able to sufficiently penetrate to the interior of large fruits. The solution to this problem, a higher concentration of the product, carries with it problems of its own, including phytopathic effects and plant tissue bleaching. Therefore, for each unique application of commodity versus pathogen, it is necessary to find a sanitizer concentration that balances anti-microbial effectiveness with acceptable commodity damage.

In summary, ClO<sub>2</sub> can be used as a sanitizer to control foodborne pathogens, yeasts, and molds on fruit. The findings in this study suggest that ClO<sub>2</sub> at low concentrations for longer time durations in active packaging is useful for improving the microbial safety and reducing decay during storage without impairing the physical properties of the fruit. Future applications of this protocol include testing the effectiveness of slow-release ClO<sub>2</sub> pouches as an addition to existing commercial packaging against the foodborne pathogens and spoilage organisms of any number of fresh food products, including fruits, vegetables, meats, and breads.

## Disclosures

The authors declare that they have no competing financial interests.

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