

Video Article

DNA Transfection of Mammalian Skeletal Muscles using *In Vivo* Electroporation

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Abstract

A growing interest in cell biology is to express transgenically modified forms of essential proteins (e.g. fluorescently tagged constructs and/or mutant variants) in order to investigate their endogenous distribution and functional relevance. An interesting approach that has been implemented to fulfill this objective in fully differentiated cells is the *in vivo* transfection of plasmids by various methods into specific tissues such as liver¹, skeletal muscle^{2,3}, and even the brain⁴. We present here a detailed description of the steps that must be followed in order to efficiently transfect genetic material into fibers of the *flexor digitorum brevis* (FDB) and *interosseus* (IO) muscles of adult mice using an *in vivo* electroporation approach. The experimental parameters have been optimized so as to maximize the number of muscle fibers transfected while minimizing tissue damages that may impair the quality and quantity of the proteins expressed in individual fibers. We have verified that the implementation of the methodology described in this paper results in a high yield of soluble proteins, i.e. EGFP and ECFP³, calpain, FKBP12, β 2a-DHPR, etc.; structural proteins, i.e. minidystrophin and α -actinin; and membrane proteins, i.e. α 1s-DHPR, RyR1, cardiac Na/Ca²⁺ exchanger, NaV1.4 Na channel, SERCA1, etc., when applied to FDB, IO and other muscles of mice and rats. The efficient expression of some of these proteins has been verified with biochemical³ and functional evidence⁵. However, by far the most common confirmatory approach used by us are standard fluorescent microscopy and 2-photon laser scanning microscopy (TPLSM), which permit to identify not only the overall expression, but also the detailed intracellular localization, of fluorescently tagged protein constructs. The method could be equally used to transfect plasmids encoding for the expression of proteins of physiological relevance (as shown here), or for interference RNA (siRNA) aiming to suppress the expression of normally expressed proteins (not tested by us yet). It should be noted that the transfection of FDB and IO muscle fibers is particularly relevant for the investigation of mammalian muscle physiology since fibers enzymatically dissociated from these muscles are currently one of the most suitable models to investigate basic mechanisms of excitability and excitation-contraction coupling under current or voltage clamp conditions^{2,6-8}.

Video Link

The video component of this article can be found at <http://www.jove.com/video/1520/>

Protocol

Experimental Procedures for *in vivo* electroporation of FDB and IO muscles

1. Before starting the *in vivo* electroporation protocols, mammalian expression plasmids must be amplified to yield concentration in the range of 2-5 μ g plasmid/ μ l of TE. *Note: We routinely use commercial amplification kits and follow the manufacturer's procedures. Commercial expression plasmids carrying the CMV promoter work very well in skeletal muscle in vivo transfections.*
2. Aliquot the necessary volume (10-20 μ l) of the plasmid solution and save it in two 0.5 ml Eppendorf tubes (one for each foot of the mouse).
3. Prepare a solution containing 2 mg/ml hyaluronidase in sterile Tyrode.
4. Using an anesthetizing box, deeply anesthetize a mouse using 4% isoflurane in O₂ with an approved gas anesthetic machine. Place the animal on a heating pad (37 °C) and maintain the anesthesia using a rodent face mask. Monitor the anesthetic depth by toe pinch reflex.
5. Under observation with a dissection microscope, inject 10 μ l of the hyaluronidase solution under the footpads of one foot of the mouse using a 1" long 33 gauge sterile needle. Penetrate the skin at a point close to the heel of the foot and advance the needle subcutaneously towards the base of the toes for ~1/4".
6. Repeat the procedure with the other foot if so desired.
7. Disconnect the anesthesia and place the mouse in a cage. Allow it to fully recover from anesthesia.
8. After one hour, anesthetize the animal for a second time and place it on the heating pad. Following the same procedure described for the hyaluronidase solution, inject a total of 20-50 μ g of the plasmid DNA (depending on the size of the plasmid construct). The total injection volume should be less than 20 μ l/foot. *Note: when 15-20 μ l is necessary, it is advisable to close the skin at the needle entry point with tissue-glue.*
9. Disconnect the anesthesia and place the mouse in a cage. Allow it to fully recover from anesthesia and wait for 10-15 min.
10. Anesthetize the animal for the third time and place it on the heating pad.

11. Select one foot of the animal. Place one gold-plated acupuncture needle under the skin at heel, and a second one at the base of the toes. Electrodes are oriented parallel to each other and perpendicular to the long axis of the foot.
12. Connect the head of the needles (electrodes) to the electrical stimulator using micro-clip connectors. Electroporate the muscles by applying 20 pulses, 20 ms in duration/each, at 1Hz. Depending on the spacing of the electrodes, the pulses' voltage amplitude is adjusted (by monitoring with an oscilloscope) to yield an electric field of ~100 V/cm. *Note: No contractions in response to the stimuli should be observed if the level of anesthesia is adequate.*
13. If so desired, repeat the above procedures in the contralateral foot of the animal.
14. Return the animal to its cage and once fully recovered from anesthesia maintain it under observation. *Note: if the procedure went normally, the animal should regain full mobility within 30 minutes and afterwards is ready to be sent back to the animal room at the vivarium. The injections of hyaluronidase and DNA in the footpads do not have noticeable adverse effects on the animals. Once recovered from anesthesia, mice are able to amble normally around the cage. As an additional precaution, add to the drinking water of the animal Carprofen at 0.0027 mg/ml for 2 days as an analgesic.*
15. Protein expression can be assayed 2-8 days after transfection. However, sustained expression of many proteins has been observed for months.

NOTE: All animal procedures were approved by the UCLA Chancellor's Animal Research Committee as mandated by the Animal Welfare Act and the PHS Policy on Humane Care and Use of Laboratory Animals.

Representative Results:

The correct implementation of the *in vivo* electroporation procedures described above should result in the effective transfection of plasmids in FDB and IO muscles. However, the efficiency of expression of transgenic protein variants will depend on the plasmid, the size and complexity of the protein, the functional properties of the protein, and a number of other variables out of our control. As illustrated in Figure 1 for an FDB muscle electroporated in the presence of a commercial plasmid (*pmR-mCherry*) encoding for *mCherry* protein, most of the muscle fibers are transfected with our protocol as illustrated by the fact the most of them display red fluorescence. This does not rule out the possibility that individual fibers exhibit different degrees of protein expression, or that few fibers are not transfected at all³. It should be noted that the efficient expression of large amounts of fluorescent proteins such as *mCherry*, EGFP, ECFP, and EYFP, does not impair the muscle fibers' excitability and excitation-contraction coupling properties. In fact, they are indistinguishable from those in sham transfected muscles (results not shown).

Practical approaches used to verify the localized expression of fluorescently-tagged proteins in skeletal muscle fibers.

The intracellular localization of the *di novo* expressed transgenic proteins is routinely evaluated by simultaneous acquiring, using the TPLSM, fluorescent images of the respective tags and images of well-identified markers of cellular structures. The most typical of these latter are: a) second harmonic generation (SHG) images, which arise from the myosin anisotropy of the sarcomeric A bands (centered at the M-lines)^{9,10} and, b) di-8-ANEPPS fluorescence images, which can be obtained by labeling the surface and transverse tubular (T-tubule) system membranes of the muscle fibers with this impermeant potentiometric dye¹¹. In fluorescence images of muscle fibers stained with di-8-ANEPPS, the T-tubules appear as narrow bands of fluorescence oriented approximately orthogonal to the long axis of the fiber. These bands are unequally spaced from each other: they are separated by a long distance which spans across the M-lines, and a short one that spans across the Z-line¹¹.

Expression and localization of α -actinin-EGFP in skeletal muscle fibers.

An example of the expression of a tagged variant of the structural muscle protein α -actinin is shown in Figure 2. This protein is known to be a major component of the Z-line and, as such, is routinely used as a marker of this structure¹². We transfected FDB and IO muscles with the plasmid *pEGFPN1- α -actinin1* encoding for human (non-muscle) α -actinin tagged at the C terminal with EGFP. Six days after transfection, we found that, as assessed by EGFP fluorescence distribution, α -actinin is mostly expressed at narrow bands equally spaced along the fiber axis. A single band per sarcomere is seen (Figures 2A & 2D). The colocalization of these bands with the Z-lines is demonstrated by comparing the distribution of EGFP fluorescence with the SHG (Figure 2B) and di-8-ANEPPS (Figure 2E) images. As shown by the overlay image (Figure 2C), α -actinin-EGFP bands alternate with the SHG bands, indicating that they are located midway between two consecutive M-bands, coinciding with the location of Z-lines. In transfected muscle fibers stained with di-8-ANEPPS, α -actinin-EGFP bands are seen centered between every pair of T-tubules (Figure 2F) which are known to flank the Z-lines (i.e. separated by a shorter distance), thus indicating that transgenic α -actinin is targeted to the Z-line.

Expression of DHPR α 1s tagged at the N-terminus with EGFP

The efficiency of muscle transfection with *pEGFPC1.1-DHPR α 1s* is verified in TPLSM images (e.g. Figure 3A) showing that most fibers express EGFP-DHPR α 1s (a transmembrane protein). The most prominent feature of transfected fibers is the double-banded pattern of EGFP fluorescence (Figures 3A & 3B) as would be expected if this protein was targeted to the T-tubules. It can also be observed in Figure 3 that while different fibers display various levels of fluorescence intensity, the banded fluorescence pattern of an individual fiber seems to be maintained homogeneous along the fiber. At higher magnification (Figure 3B), it can be clearly seen that the uneven spacing among bands is similar to that observed in fibers stained di-8-ANEPPS (e.g. Figure 2E). The overlay images (Figure 3B & 3E) illustrate that the SHG bands are located within the larger spacing between EGFP-DHPR α 1s bands, corroborating that this protein is at the T-tubules. Additional FRET measurements with the non-fluorescent lipophilic anion DPA- (data not shown) further demonstrate that the EGFP moiety is within few nanometers of the inner leaflet of the T-tubules' membranes.

Localization and functional evaluation of the expression of EYFP-CIC1

Fibers transfected with *pEYFP-CIC1*, which encodes an EYFP-tagged construct (at the N-terminal) of the skeletal muscle chloride channel (CIC1), display EYFP fluorescence bands (Figure 4A) with a pattern similar to that observed for EGFP-DHPR α 1s' expression (Figure 3A) and corresponding to the T-tubule arrangement as illustrated with di-8-ANEPPS staining (Figure 2E). As expected, the superimposition of EYFP-CIC1

(Figure 4A) and SHG images (Figure 4B) as shown in the overlay (Figure 4C), illustrate that the SHG bands are centered at the large spacing of between EYFP fluorescence bands.

In order to assess whether the expression of EYFP-CIC1 results in a significant increase in the resting conductance of the muscle fibers, as expected from the overexpression of functional chloride channels, we enzymatically dissociated muscle fibers from the transfected FDB muscle and studied their electrophysiological properties using a two-microelectrodes experimental setup described previously^{6,11,13}. Figure 5 shows results from two fibers: one expressing large amounts of EYFP-CIC1 as assessed from global fluorescence intensity measurements in a standard fluorescence microscope (not shown), and the other is a non-transfected control. The voltage record in Figure 5A (obtained from the muscle fiber expressing EYFP-CIC1) demonstrates that due to the excessive resting conductance, the fiber is almost non-excitable. Current stimulus pulses of up to 400nA (0.5ms) needed to be used in order to elicit a very small active response (no overshoot, Figure 5A). After replacement of the external Tyrode solution to one containing 500 μ M of the chloride channel blocker 9-ACA, 200 nA current pulses were sufficient to elicit an action potential (Figure 5B), although much slower and broader than those recorded in the control fiber (Figure 5C). As expected, addition of 9-ACA had minimal effects on this control fiber (Figure 5D).

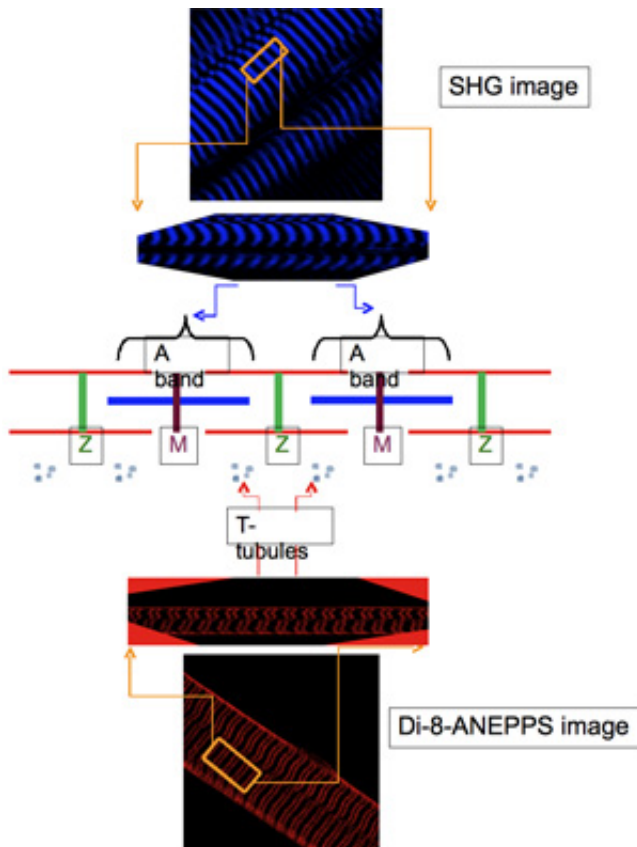


Figure 1. Transfection efficiency of the in-vivo electroporation method. Brightfield (A) and fluorescence (B) images of an FDB muscle transfected with *pmR-mCherry*. Days after electroporation protocol: 12 days. Please [click here](#) for a larger version of figure 1.

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Figure 2

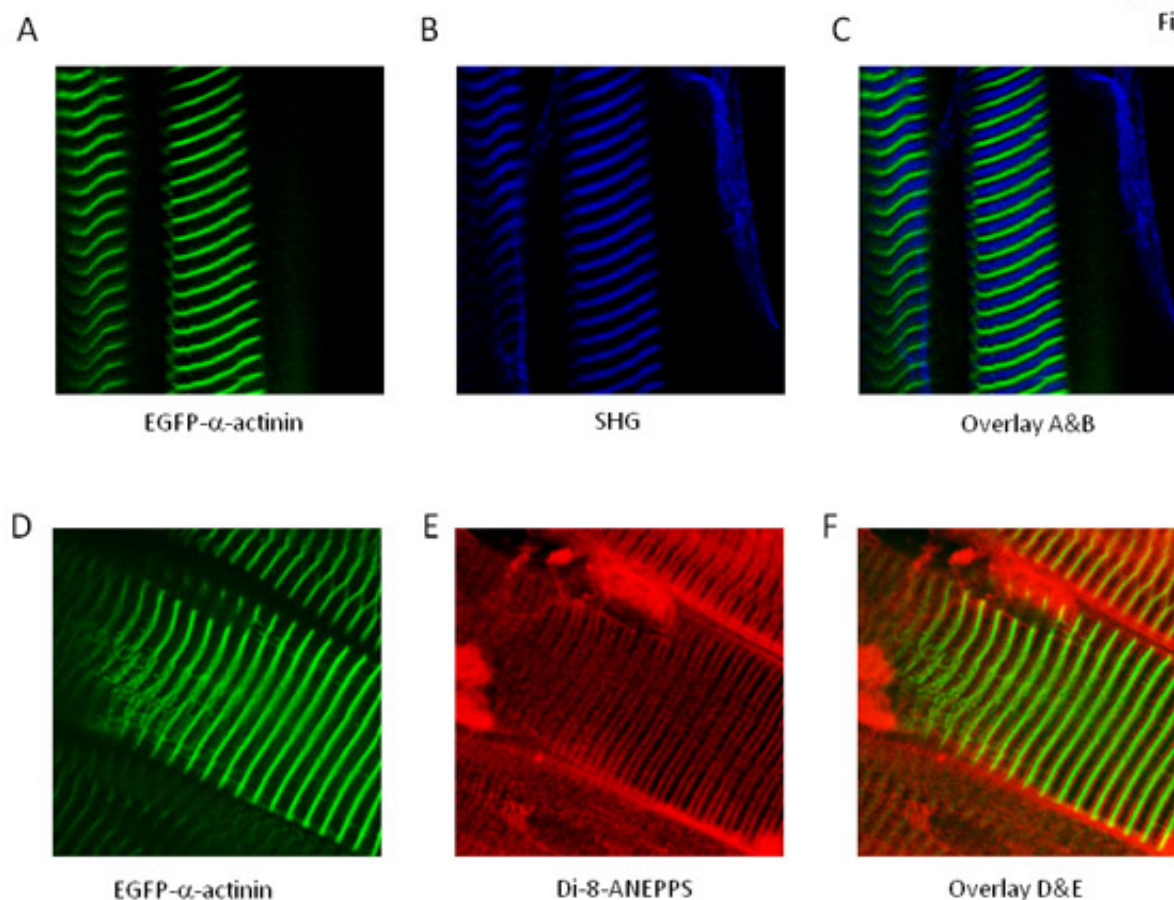


Figure 2. Expression and targeting of α -actinin-EGFP in FDB fibers. Panels A and B are EGFP fluorescence and SHG images, respectively, of a fiber expressing α -actinin-EGFP. Panel C is a superposition of the images in A and B. Panels D and E are fluorescence images of another fiber expressing α -actinin-EGFP and stained with di-8-ANEPPS, respectively. Panel F is the superposition of images in D and E. Days after electroporation protocol: 6 days.

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Figure 3

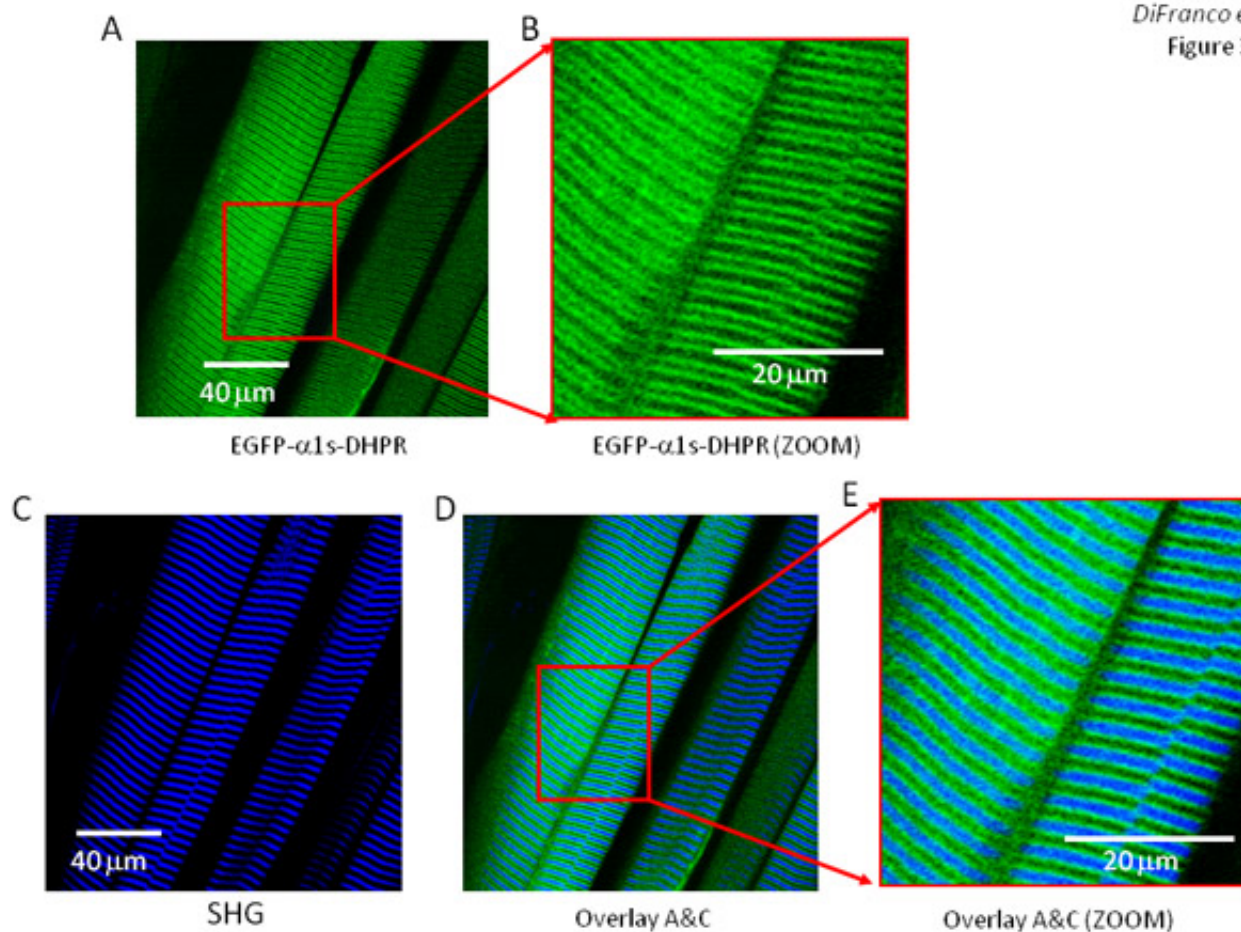


Figure 3. Expression and targeting of EGFP-DHPR $\alpha 1s$ in FDB fibers. Panel A, EGFP fluorescence image of a group of fibers expressing EGFP-DHPR $\alpha 1s$. Panel B is an enlargement of the square in panel A in order to better show the banded pattern of the protein expression. Panel C is the SHG image corresponding to the image in panel A. Panel D is the overlay of the images A and C. Panel E is an enlargement of the area indicated in panel D. Days after electroporation protocol: 20 days.

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Figure 4

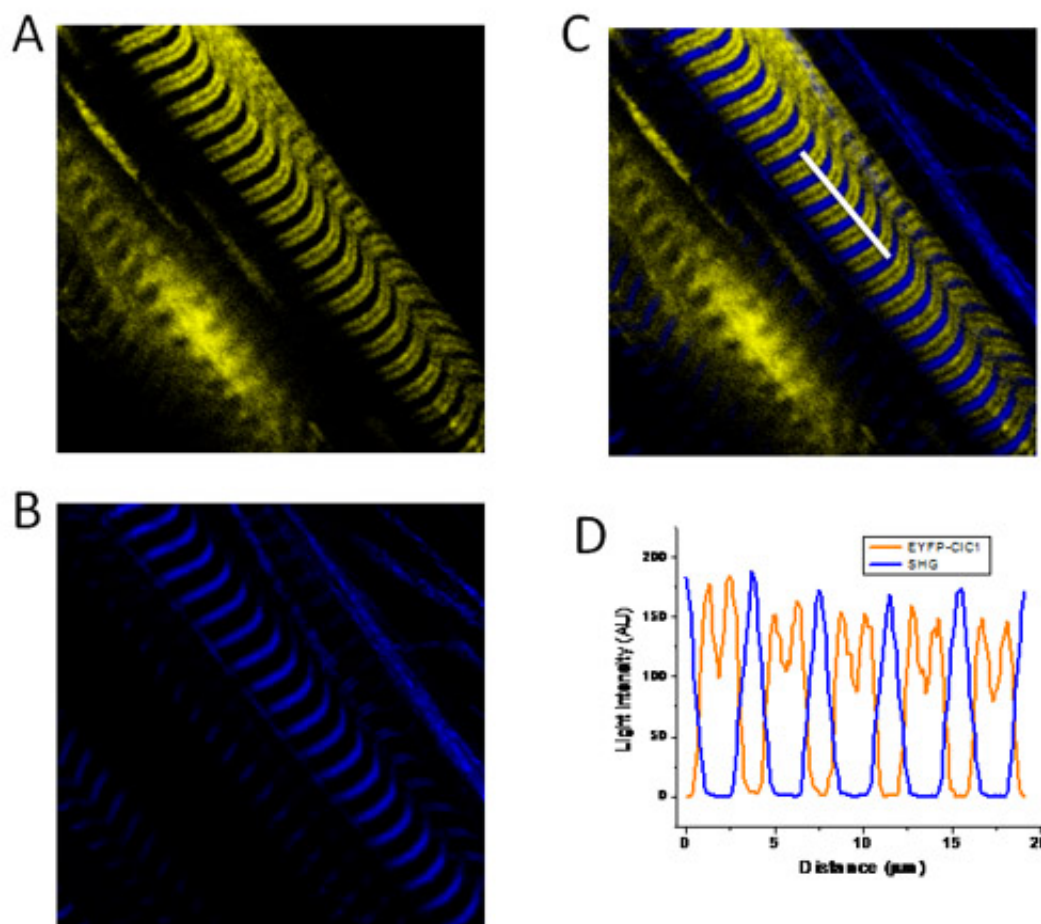


Figure 4. Expression and targeting of EYFP-CIC1 in FDB fibers. Panels A and B are EYFP fluorescence and SHG images, respectively, of fibers expressing EYFP-CIC1. Panel C is the superposition of the images in panels A and B. Panel D is an intensity profile measured along the white line highlighted in the image in panel C. Days after electroporation protocol: 7 days.

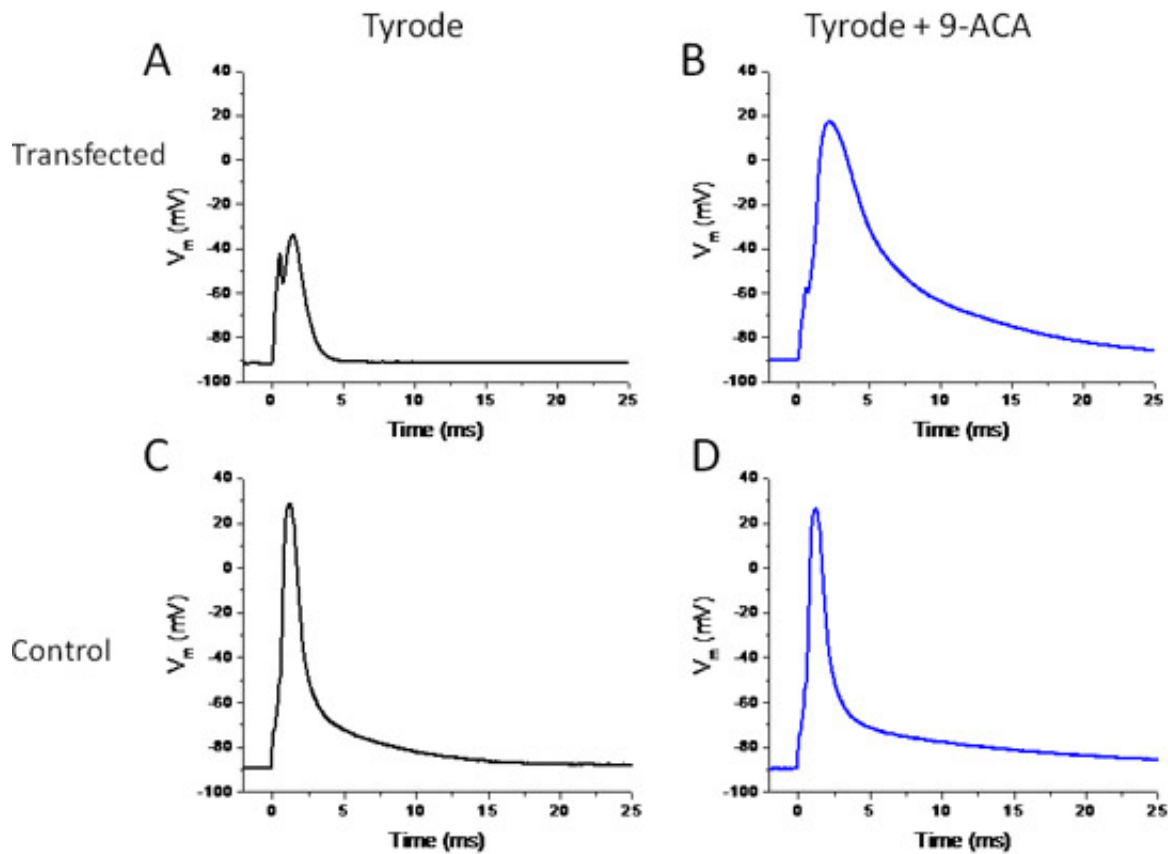


Figure 5. Electrophysiological assessment fibers expressing transgenic EYFP-CIC1. Panels A and B are voltage records from a fiber expressing EYFP-CIC1 in response to current pulses before and after treatment with 9-ACA, respectively. Panels C and D are voltage records (action potentials) elicited in response to current pulses in a non-transfected fiber before and after treatment with 9-ACA, respectively.

Discussion

We describe here the detailed steps that should be followed in order to attain effective transfections of DNA plasmids into skeletal muscle fibers by *in vivo* electroporation. The main advantages of our approach are the simplicity of implementation, and its minimal invasiveness which results in negligible health hazard to the animals. In reality, the above described electroporation protocols do not entail much more than two subcutaneous injections per foot, followed by an electrical stimulation protocol, all of which are well-tolerated by the animals under anesthesia. In our institution these manipulations are altogether determined to be non-surgical procedures since they do not involve opening of the skin. Besides, the mildness of the electroporation procedures is a critical factor for the overall success in attaining an efficient plasmid transfection and the subsequent protein expression by the muscle fibers since any significant cell damage would impair their ability to efficiently engage in active protein synthesis.

Minor adjustments to the procedures described here can be (and have been) implemented for larger muscles of the mouse foot (e.g. *extensor digitorum longus*, EDL; *tibialis anterior*, TA; and, soleus muscles). Furthermore, by just scaling up the total amount of DNA, we have implemented the exact procedures described here in order to transfect the FDB and IO muscles of adult rats with great success.

Although transfection protocols consistently result in a variable level of expression of transgenic proteins by muscle fibers, this can be taken advantage of in physiological experiments; non-expressing fibers can be used as controls, and fibers with different level of expression can be used to correlate the level of expression with the level of function change. Also, CMV promoters have been found to be suitable in yielding high levels of protein expression, other muscle-specific promoters may improve the efficacy of protein expression. It should be noted that when it is not possible (or convenient) to express fluorescently-tagged protein constructs in muscle fibers, we have succeeded in expressing untagged proteins and monitor the level of expression with *bicistronic* plasmids that simultaneously encode a fluorescent protein.

Disclosures

The authors have nothing to disclose.

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