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Title: PIPEMAT-RS: Development and Validation of a Standardized MATLAB Pipeline for Resting-State EEG Preprocessing

Authors and Affiliations:

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Author Questionnaire

- **1. Microscopy**: Does your protocol require the use of a dissecting or stereomicroscope for performing a complex dissection, microinjection technique, or something similar? **No**
- **2. Software:** Does the part of your protocol being filmed include step-by-step descriptions of software usage? **Yes**
- 3. Filming location: Will the filming need to take place in multiple locations? NO

Current Protocol Length

Number of Steps: 12 Number of Shots: 30



Introduction

Videographer: Obtain headshots for all authors available at the filming location.

Videographer's NOTE: It was a bit difficult to find a location for the testimonies, since it was a public hospital. We filmed in the most suitable place I could find.

As for the photos of the authors, only Lucas was present at the hospital, so I couldn't take any photos of anyone else.

REQUIRED:

- 1.1. <u>Lucas Murrins Marques:</u> We developed a standardized EEG preprocessing pipeline to improve signal quality, reduce noise, and support consistent neurophysiological research across diverse clinical and experimental datasets.
 - 1.1.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B-roll: 3.2.1, 3.2.2.*

What are the most recent developments in your field of research?

- 1.2. <u>Lucas Murrins Marques:</u> Recent pipelines like RELAX and Automagic offer automated EEG cleaning but often lack flexibility or clear documentation for researchers and students who need transparency and consistency of results.
 - 1.2.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera.

What technologies are currently used to advance research in your field?

- 1.3. <u>Lucas Murrins Marques:</u> We use EEG, MATLAB, EEGLAB, and ICA-based artifact rejection combined with automated classifiers like ICLabel to enhance data quality and streamline preprocessing workflows.
 - 1.3.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B-roll: 4.2.1, 4.2.2., 4.2.3.*

What significant findings have you established in your field?



- 1.4. <u>Lucas Murrins Marques:</u> It has been found that PIPEMAT-RS improves EEG signal quality, increases SNR, and supports robust biomarkers in stroke, fibromyalgia, and chronic pain studies.
 - 1.4.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B-roll: LAB MEDIA: Figure 2B, 2C.*

What research gap are you addressing with your protocol?

- 1.5. <u>Lucas Murrins Marques:</u> This protocol addresses the lack of an editable, well-documented, and standardized pipeline for resting-state EEG that is both consistent and accessible to researchers worldwide.
 - 1.5.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B-roll: 3.6.1, 3.6.2, 3.6.3.*

Videographer: Obtain headshots for all authors available at the filming location.



Testimonial Questions:

Videographer's NOTE: It was a very short script, so there are few files. I took the opportunity to take a wide shot and a close shot for all the testimonies.

It was a bit difficult to find a location for the testimonies, since it was a public hospital. We filmed in the most suitable place I could find.

How do you think publishing with JoVE will enhance the visibility and impact of your research?

- 1.6. <u>Lucas Murrins Marques: Instructor Professor in Mental Health, Santa Casa de São Paulo School of Medical Sciences:</u> Publishing with JoVE will greatly expand the reach of our pipeline by making it visually accessible and easy to understand, especially for international labs and early-career researchers seeking clear, step-by-step EEG preprocessing guidance.
 - 1.6.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera.

Can you share a specific success story or benefit you've experienced—or expect to experience—after using or publishing with JoVE?

- 1.7. <u>Lucas Murrins Marques:</u> <u>Instructor Professor in Mental Health, Santa Casa de São Paulo School of Medical Sciences</u>: We expect JoVE to streamline onboarding in our lab by reducing training time. The video format will also facilitate the adoption of PIPEMAT-RS in partner institutions and support collaborations in multicenter EEG studies.
 - 1.7.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera.



Ethics Title Card

This research has been approved by the Ethics Committee of the Clinics Hospital, University of São Paulo Medical School



Protocol

2. File Format Conversion Procedure

Demonstrator: Lucas Murrins Marques

Videographer: Please record the screen for all the SCREEN shots as a backup.

NOTE: The author wants to use still images only and to make it dynamic add emphasis, zoom in, zoom out as needed.

- 2.1. To begin, open MATLAB (Mat-lab) and navigate to the directory containing the raw electroencephalogram data files [1]. Define the directory path to the raw data files [2] and create a list containing the subjects [3-TXT].
 - 2.1.1. WIDE: Talent seated at a workstation opening MATLAB and browsing to the specified directory.
 - 2.1.2. SCREEN: Screen_212.tif Video Editor: Emphasize or highlight the selected part in the image.
 - 2.1.3. SCREEN: Screen_212.tif **TXT: Skip this step if the dataset is already in .mat format** *Video Editor: Emphasize or highlight the selected part in the image including the text written in green above this (% List of raw EEG....).*
- 2.2. Load the raw data file using the **load** function for .mat (dot-Mat) files [1] and save each file in .mat format, ensuring data integrity and retaining all relevant metadata [2].
 - 2.2.1. SCREEN: Screen_221.tif Video Editor: Only show the MATLAB window.
 - 2.2.2. SCREEN: Screen_222.tif

3. EEG Montage, Downsampling and Filtering, and Artifact Rejection

- 3.1. Open MATLAB and navigate to the directory containing the MATLAB files [1].
 - 3.1.1. SCREEN: Screen 3.1.1.tif
- 3.2. Use the **pop_loadset** (*Pop-Loadset*) function from the **EEGLAB** (*E-E-G-Lab*) toolbox to load each subject's EEG (*E-E-G*) dataset [1]. Ensure the appropriate electrode location file corresponding to the EEG cap configuration is available in the working directory [2].



- 3.2.1. SCREEN: Screen 3.2.1.tif Video Editor: Only show the MATLAB window and emphasize or highlight the selected part in the image.
- 3.2.2. SCREEN: Screen 3.2.2.tif Video Editor: Emphasize or highlight the selected part in the image.
- 3.3. Use the pop chanedit (pop-underline-chanedit) function to apply the electrode locations [1]. Visually inspect the channel layout using the EEGLAB's GUI (G-U-I) to confirm the correct positions of all electrodes [2]. Save the updated EEG dataset using the **pop** saveset (pop-underline-saveset) function [3].

3.3.1. SCREEN: Screen_3.3.1.tif

3.3.2. SCREEN: Screen 3.3.2.tif

3.3.3. SCREEN: Screen 3.3.3.tif

- 3.4. If required, reduce the sampling rate of the EEG data to 250 hertz using the pop resample (Pop-re-sample) function [1]. Apply a band-pass filter from 1 hertz to 50 hertz to channels 1 to 128 using the pop_basicfilter (Pop Basic Filter) function with a Butterworth design and filter order of 2. Use zero-phase forward and reverse filtering to prevent phase distortion [2].
 - 3.4.1. SCREEN: Screen 3.4.1.tif Video Editor: Emphasize or highlight the selected part in the image
 - 3.4.2. SCREEN: Screen 3.4.2.tif Video Editor: Only show the MATLAB window and emphasize or highlight the selected part in the image.
- 3.5. Apply a notch filter at 50 or 60 hertz to channels 1 through 64 using pop basicfilter with the **PMnotch** (*P-M-Notch*) design and a filter order of 180 [1]. For the artifact rejection, use the clean rawdata (Clean Raw Data) function with the given parameters to automatically detect and remove flatline channels, noisy segments, and low-frequency drifts [2,3].

3.5.1. SCREEN: Screen 3.5.1.tif

3.5.2. SCREEN: Screen 3.5.2.tif Video Editor: Only show the MATLAB window.

AND,

3.5.3. TEXT on PLAIN BACKGROUND:

FlatlineCriterion (5): Removes channels flat for more than 5 s.



ChannelCriterion ([-1]): Disables channel correlation-based rejection.

LineNoiseCriterion (0.7): Removes channels with excessive line noise above this threshold.

Highpass (-1): Disables additional high-pass filtering during cleaning.

BurstCriterion (8): Removes data bursts that exceed 8 standard deviations.

WindowCriterion (0.85): Requires 85% of data in a window to be clean to retain it.

Video Editor: If possible, show 3.5.3 as an inset or something similar during 3.5.2.

3.6. Visually inspect the EEG data using **eegplot** (*E-E-G-Plot*) from **EEGLAB** to identify any residual artifacts missed by the automated method [1]. Manually mark and remove the remaining artifacts to ensure high data quality [2]. Use **pop_reref** (*pop-underline-reref*) to rereference the EEG signals to the average of all electrodes, ensuring the original reference electrode is retained [3].

3.6.1. SCREEN: Screen 3.6.1.tif

3.6.2. SCREEN: Screen 3.6.2.tif Video Editor: Highlight the green part in the plot.

3.6.3. SCREEN: Screen_3.6.3.tif

4. Independent Component Analysis and ICLabel Classification Procedure

4.1. Load the file with the suffix _loc_filt_cleanraw_reref.set (Loc-underline-filt-underline-cleanraw-underline-reref-dot-set) using pop_loadset [1]. Run Independent Component Analysis or ICA (I-C-A) using the pop_runica (pop-runica) function with the runica algorithm to decompose the EEG data into independent components [2-TXT]. Save the new dataset with the suffix _loc_filt_cleanraw_reref_ICA using pop_saveset to indicate ICA completion [3].

4.1.1. SCREEN: Screen 4.1.1.tif

4.1.2. SCREEN: Screen_4.1.2.tif **TXT: The 'runica' algorithm maximizes the statistical independence of the components** *Video Editor: Only show the MATLAB window and emphasize or highlight the selected part in the image.*

4.1.3. SCREEN: Screen 4.1.3.tif

4.2. Import the ICA dataset using **pop_loadset [1]**. Run the **pop_iclabel** (*Pop-I-C-Label*) function with the **default** model to classify independent components into brain and



artifact categories [2]. Identify all components with brain activity probability above 0.7 [3].

- 4.2.1. SCREEN: Screen_4.2.1.tif4.2.2. SCREEN: Screen_4.2.2.tif4.2.3. SCREEN: Screen_4.2.3.tif
- 4.3. Use the pop_subcomp (pop-subcomp) function to remove all components with ICLabel (I-C-Label) brain probability below 0.7 [1]. Retain only components with a brain activity probability above 0.7 to preserve genuine signals while ensuring the effective removal of artifacts [2]. Save the cleaned dataset with the suffix _loc_filt_cleanraw_reref_ICA_Cleaned (Loc-underline-filt-underline-cleanraw-underline-reref-underline-I-C-A-underline-Cleaned) using pop_saveset [3].
 - 4.3.1. SCREEN: Screen_4.3.1.tif Video Editor: Only show the MATLAB window and emphasize or highlight the selected part in the image.
 - 4.3.2. SCREEN: Screen_4.3.2.tif 4.3.3. SCREEN: Screen_4.3.3.tif
- **4.4.** Finally, import the dataset with artifact-free ICA components using the **pop_loadset** function **[1]** and save the fully preprocessed and normalized dataset with the suffix **_loc_filt_cleanraw_reref_ICA_Normalized** using **pop_saveset [2]**.
 - 4.4.1. SCREEN: Screen_4.4.1.tif Video Editor: Only show the MATLAB window and emphasize or highlight the selected part in the image.
 - 4.4.2. SCREEN: Screen 4.4.2.tif



Results

5. Results

- **5.1.** The initial state of the EEG dataset prior to any preprocessing, including both [1] the raw signal traces [2] and their corresponding spectral characteristics [3], is illustrated in this figure [4].
 - 5.1.1. LAB MEDIA: Figure 2A.
 - 5.1.2. LAB MEDIA: Figure 2A. Video Editor: Highlight the left image (Scroll Plot).
 - 5.1.3. LAB MEDIA: Figure 2A. Video Editor: Highlight the right image (Spectral Plot).
 - 5.1.4. LAB MEDIA: Figure 2A.
- **5.2.** The raw EEG data exhibited high levels of noise and irregular signal traces across multiple channels, compromising clarity and interpretability [1].
 - 5.2.1. LAB MEDIA: Figure 2A.
- **5.3.** Manual cleaning of the EEG data reduced signal artifacts [1] and improved waveform uniformity [2], but some residual noise and inconsistencies remained visible [3].
 - 5.3.1. LAB MEDIA: Figure 2A, 2B. *Video Editor: Highlight the left image in 2B (Scroll Plot).*
 - 5.3.2. LAB MEDIA: Figure 2A, 2B. *Video Editor: Highlight the right image in 2B (Spectral Plot).*
 - 5.3.3. LAB MEDIA: Figure 2B.
- **5.4.** Data processed with PIPEMAT-RS (*Pipe-Mat-R-S*) displayed the cleanest EEG waveforms with consistent signal structure and minimal visible artifacts across channels [1].
 - 5.4.1. LAB MEDIA: Figure 2B, 2C. Video Editor: Highlight 2C.

Pronunciation Guides:

? montage

Pronunciation link:

https://www.merriam-webster.com/dictionary/montage

IPA: /'man.taz/

Phonetic Spelling: mon-tahzh



downsampling

Pronunciation link:

https://www.collinsdictionary.com/us/dictionary/english/downsampling

IPA: /ˈdaʊnˌsæmpəlɪŋ/

Phonetic Spelling: down-SAMP-ling

2 artifact

Pronunciation link:

https://www.merriam-webster.com/dictionary/artifact

IPA: /ˈartəˌfækt/

Phonetic Spelling: AR-tuh-fakt

notch

Pronunciation link:

https://www.merriam-webster.com/dictionary/notch

IPA: /natʃ/

Phonetic Spelling: notch

resample

Pronunciation link:

https://www.merriam-webster.com/dictionary/resample

IPA: /riˈsæmpəl/

Phonetic Spelling: ree-SAMP-uhl

IndependentPronunciation link:

https://www.merriam-webster.com/dictionary/independent

IPA: /ˌɪndəˈpɛndənt/

Phonetic Spelling: in-duh-PEN-dent