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## **Title: Orthotopic Left Lung Transplantation in Rats**

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## **Author Questionnaire**

- 1. Microscopy:** Does your protocol require the use of a dissecting or stereomicroscope for performing a complex dissection, microinjection technique, or something similar? **Yes, all done**
- 2. Software:** Does the part of your protocol being filmed include step-by-step descriptions of software usage? **NO**
- 3. Filming location:** Will the filming need to take place in multiple locations? **NO**

### **Current Protocol Length**

Number of Steps: 27

Number of Shots: 53

# Introduction

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*Videographer: Obtain headshots for all authors available at the filming location.*

- 1.1. **Hailin Liao:** My research focuses on developing a standardized, reproducible rat lung transplantation model to investigate post-transplant complications, particularly chronic rejection, and to better understand mechanisms driving long-term graft failure.

1.1.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B.roll: 4.7*

What advantage does your protocol offer compared to other techniques?

- 1.2. **Hailin Liao:** Our protocol offers a shorter learning curve, single-operator capability, and higher reproducibility compared to existing techniques.

1.2.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera.

How will your findings advance research in your field?

- 1.3. **Yi Lu:** Our standardized rat lung transplantation model provides a reliable platform for studying complications after lung transplantation and can accelerate translational research to improve graft survival.

1.3.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B.roll:4.8.2*

What research questions will your laboratory focus on in the future?

- 1.4. **Yi Lu:** In the future, our laboratory will focus on the study of chronic rejection after lung transplantation, in order to improve the long-term survival rate of lung transplant recipients.

1.4.1. INTERVIEW: Named Talent says the statement above in an interview-style shot, looking slightly off-camera.

*Videographer: Obtain headshots for all authors available at the filming location.*

**Ethics Title Card**

This research has been approved by the Animal Care and Use Committee at Guangzhou  
Lai'an Technology Co., Ltd

# Protocol

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## 2. Donor Lung Harvest for Syngeneic Lung Transplant in Rats

**Demonstrator:** Hailin Liao

- 2.1. To begin, obtain 14-gauge and 16-gauge intravenous catheters and blades [1]. Trim the catheters into a cuff with a tail with a blade [2]. Then divide each cuff into a body and a tail, each approximately 2 to 3 millimeters in length [3].
  - 2.1.1. WIDE: Talent holding 14 and 16 G intravenous catheters and blades.
  - 2.1.2. SCOPE: 2.1.2.Recommended selection 42s-53s and 1min07s-1min52s.mp4 00:42-00:54.
  - 2.1.3. SCOPE: 2.1.2.Recommended selection 42s-53s and 1min07s-1min52s.mp4 01:07-01:22.
- 2.2. Using a scalpel, create superficial scratches on the cuff body to increase friction for suture fixation [1].
  - 2.2.1. SCOPE: 2.2.1.Recommended selection 1min40s-1min44s.mp4 01:40-01:49
- 2.3. To harvest the donor lung, retract the tongue of an anesthetized rat outward and upward using forceps [1-TXT]. Position a surgical lamp anterior to the neck to illuminate the glottal opening clearly [2].
  - 2.3.1. Talent using forceps to extend the tongue of an anesthetized rat. **TXT: Anesthesia: Pentobarbital injection (i.p) (80 mg/kg)**
  - 2.3.2. Talent adjusting a surgical lamp in front of the rat's neck.
- 2.4. Now insert a 14-gauge intravenous cannula into the airway through the glottis [1]. Set the ventilator to pressure-controlled mode [2]. Then input the weight parameters, adjust the pressure to 15 centimeters of water, and connect the ventilator [3].
  - 2.4.1. Talent inserting the cannula carefully into the glottal opening.
  - 2.4.2. Talent inputs weight, adjusts pressure and connects the ventilator.
- 2.5. Observe the rise and fall of the chest to ensure synchronization with the ventilator frequency [1].

- 2.5.1. Shot of chest movement aligned with ventilator rhythm.
- 2.6. Next, fix the limbs and head using tape or restraints [1]. Disinfect the chest and abdomen with alcohol [2].
  - 2.6.1. Talent securing limbs and head.
  - 2.6.2. Talent disinfecting the chest and abdomen with alcohol. .
- 2.7. Then lift the abdominal skin using tweezers [1] and cut the skin from the abdomen to the front of the neck with a pair of scissors [2].
  - 2.7.1. Talent lifting abdominal skin with tweezers.
  - 2.7.2. Shot of an incision being made from abdomen up to the neck.
- 2.8. Make a midline incision along the abdominal wall [1]. Inject heparin through the exposed peritoneal vein, then allow systemic circulation for 3 minutes to ensure complete heparinization[2-TXT].
  - 2.8.1. Talent performing the midline incision.
  - 2.8.2. Talent injecting heparin with a syringe into the vein.**TXT: Heparin: 1000 IU/kg**
- 2.9. Cut the diaphragm and open the chest cavity from the midline of the sternum [1]. Then fix the chest wall on both sides with hemostats [2]. Remove the thymus to expose the chest cavity organs [3].
  - 2.9.1. Talent cutting diaphragm and mid-sternum.
  - 2.9.2. Shot of the chest wall being fixed with hemostats.
  - 2.9.3. Talent removing thymus with forceps.
- 2.10. Sequentially cut the superior vena cava, inferior vena cava, and the left and right auriculae [1]. Now inject cold saline into the root of the pulmonary artery using a 20-milliliter syringe over 1 minute for low-pressure perfusion until the donor lung turns completely white [2].
  - 2.10.1. Talent cutting major vessels.
  - 2.10.2. Talent slowly injecting cold saline at the root of the pulmonary artery until lung turns white.
- 2.11. Cut tissues connecting the donor lung and extract the donor heart and lung *en bloc* [1]. Soak the donor heart-lung block in precooled saline [2].
  - 2.11.1. Talent cutting connecting tissues and removing the organ block.
  - 2.11.2. Talent placing the heart-lung block in a saline-filled container.

### **3. Donor Lung Modification for Transplant**

- 3.1. Position the heart-lung block on ice under a microscope [1]. Grip the trachea with a hemostat and secure it in plasticine [2].

3.1.1. Talent placing organ on ice under microscope.

3.1.2. ~~SCOPE: The trachea is being gripped with a hemostat.~~

**AUTHOR'S NOTE: Shot deleted**

3.1.3. Shot of the trachea being secured in plasticine.

- 3.2. ~~Cover the lungs with wet sterile lens paper [1]. Separate the lungs to expose the pulmonary hilum fully [2].~~

3.2.1. Talent placing wet paper over lungs.

3.2.2. ~~SCOPE: The lungs are being separated until hilum is exposed.~~

**AUTHOR'S NOTE: Shot deleted**

- 3.3. Now use forceps to separate the pulmonary artery, bronchus, and pulmonary vein under the microscope [1]. Ligate the bronchus close to the lung using 6-0 surgical sutures [2]. Use spring scissors to cut the pulmonary artery, bronchus, and pulmonary vein [3].

3.3.1. SCOPE: 3.3.1.Recommended selection 12s-27s and 33s-48s and 1min52s-2min40s.mp4 00:12-00:27, 00:40-00:44, 02:34-02:40

3.3.2. SCOPE: 3.3.2.mp4 00:01-00:18.

*Added shot: SCREEN: 3.3.3 add SCOPE.mp4 00:01-00:09, 00:14-00:18, 00:26-00:35*

- 3.4. Extract the pulmonary artery, bronchus, and pulmonary vein from respective cuffs made from 16-gauge, 14-gauge, and 16-gauge indentation catheters [1]. Fix the tube walls to the cuff with 8-0 surgical sutures [2].

3.4.1. SCOPE: 3.4.1.mp4 00:01-00:12,.

3.4.2. SCOPE: 3.4.2.mp4 00:40-00:50 .

- 3.5. Preserve the donor lungs in saline and place them back on ice until implantation [1].

3.5.1. Talent placing donor lungs in saline on an iced platform.

### **4. Donor Lung Implantation**

- 4.1. Depilate the left thoracodorsal region of the recipient animal [1]. Position the animal in the right lateral decubitus posture on a thermostatic operating table [2-TXT].

- 4.1.1. Talent removing fur from the recipient's thoracodorsal region.
- 4.1.2. Talent positioning recipient in the right lateral decubitus posture. **TXT: Disinfect surgical field with 75% alcohol**
- 4.2. Now make an incision through the skin and chest wall at the point of the apical impulse in the fourth intercostal space to access the thoracic cavity [1]. Use an eyelid retractor to open the thoracic cavity [2]. Then gently push the left lung aside using a moist cotton swab to expose and sharply transect the inferior pulmonary ligament [3].
  - 4.2.1. SCOPE: 4.2.1 (2) .Recommended selection 8s-1min25s.mp4 00:05-00:14, 01:03-01:25.
  - 4.2.2. SCOPE: 4.2.2.mp4 00:00-00:16.
  - 4.2.3. SCOPE: 4.2.3.mp4 00:01-00:26.
- 4.3. Next, grasp the left lung with forceps and retract it outside the thoracic cavity [1]. Secure the hilum with a hemostat then stabilize it with modeling clay [2].
  - 4.3.1. Talent holding the left lung with forceps and pulling the lung gently outside.
  - 4.3.2. Talent fixing hilum with hemostat and applying modeling clay for stability.
- 4.4. Now dissect the pulmonary hilum to separate the pulmonary artery, pulmonary vein, and bronchus [1].
  - 4.4.1. SCOPE: 4.4.1.Recommended selection 15s-58s and 1min20s-1min33s.mp4 00:15-00:19, 01:20-01:33, .
- 4.5. Clamp the proximal ends of the pulmonary artery, pulmonary vein, and bronchus using vascular clamps [1]. Pre-tie the recipient's pulmonary artery, bronchus, and pulmonary vein with 8-0 surgical sutures for rapid anastomosis [2].
  - 4.5.1. SCOPE: 4.5.1.mp4 00:00-00:14.
  - 4.5.2. SCOPE: 4.5.2.mp4 00:13-00:36
- 4.6. Trim a platform on the recipient's clamped left lung to facilitate the placement of the donor lung [1]. Make small incisions at the distal ends of the pulmonary artery, pulmonary vein, and bronchus [2].
  - 4.6.1. SCOPE: 4.6.1.mp4 00:00-00:18.
  - 4.6.2. SCOPE: 4.6.2.mp4 00:00-00:10, 00:15-00:23, 00:31-00:35.



4.7. Rinse the pulmonary artery and pulmonary vein with heparinized saline to prevent clot formation [1]. Then take the donor lung from ice and position it on the prepared platform of the recipient's left lung [2].

4.7.1. SCOPE: 4.7.1.mp4 00:03-00:26

4.7.2. Shot of the donor lung being removed from ice and being placed on the recipient's left lung.

4.8. Use forceps to lift one side of the cut opening , Then sequentially implant and ligate the cuffs of the pulmonary artery, pulmonary vein, and bronchus into the recipient [1].

4.8.1. SCOPE: 4.8.1 (1).mp4 00:16-00:45 .

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4.9. Open the micro hemostatic clip and observe the donor lung turning from white to red [1]. Inspect the anastomotic sites for any signs of bleeding [2].

4.9.1. SCOPE: 4.9.1.mp4 00:02-00:14 .

4.9.2. SCOPE: 4.9.2.mp4 00:00-00:12 .

4.10. Now remove the recipient's left lung and return the donor lung into the thoracic cavity [1-TXT].

4.10.1. SCOPE: 4.10.1(1).mp4 00:03-00:16

. **TXT: Increase ventilator pressure to aid expansion**

4.11. Wipe the chest cavity dry with sterile cotton [1]. Then close the thoracic cavity layer by layer [2]. Wait for spontaneous breathing recovery before removing the ventilator [3-TXT]. After one hour, the recipients were free to eat [4].

4.11.1. SCOPE: 4.11.1.mp4 00:00-00:08.

4.11.2. SCOPE: 4.11.2(3).mp4 00:56-01:10

4.11.3. Talent monitoring breathing and disconnecting ventilator. **TXT: Remove tube with recipient to gradually recover spontaneous breathing**

*Added shot: Recipient eating food.*

## Results

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### 5. Results

- 5.1. Six months after transplantation, CT(C-T) imaging showed that the ventilation of the transplanted lungs was comparable to the sham group, indicating preserved lung function [1]. The macroscopic appearance of the lungs did not differ visibly between the sham and transplanted groups, showing no signs of deterioration or abnormality [2].
  - 5.1.1. LAB MEDIA: Figure 5. *Video editor: Please highlight areas pointed at by the red arrows in the transplantation image of CT*
  - 5.1.2. LAB MEDIA: Figure 5. *Video editor: Highlight both lung images in the “Macroscopic view” column*
- 5.2. Hematoxylin and Eosin staining of the lung tissues revealed no significant pathological changes in the transplanted lungs compared to the sham group [1], with preserved alveolar structures and absence of inflammatory infiltration [2].
  - 5.2.1. LAB MEDIA: Figure 5. *Video editor: Highlight the entire stained tissue sections in the “HE staining” column for both groups.*
  - 5.2.2. LAB MEDIA: Figure 5. *Video editor: Zoom in on the magnified inset images*

Pronunciation Guide:

🔍 **Orthotopic**

Pronunciation link:

<https://www.howtopronounce.com/orthotopic>

IPA: /ˌɔːrθəˈtɑːpɪk/

Phonetic Spelling: or-thuh-tah-pik

🔍 **Anastomosis**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/anastomosis>

IPA: /əˌnæs.təˈmoʊ.sɪs/

Phonetic Spelling: uh-nass-tuh-moh-sis

🔍 **Auriculae**

Pronunciation link:

<https://www.howtopronounce.com/auriculae>

IPA: /ɔːˈrɪk.jʊ.li/

Phonetic Spelling: aw-rik-yuh-lee

🔍 **Hilum**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/hilum>

IPA: /ˈhaɪ.ləm/

Phonetic Spelling: hy-luhm

🔍 **Peristalsis**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/peristalsis>

IPA: /ˌper.əˈstɔɪ.lɪsɪs/

Phonetic Spelling: peh-ruh-stawl-sis

🔍 **Cuff** (as in catheter cuff)

Pronunciation link:

<https://www.merriam-webster.com/dictionary/cuff>

IPA: /kʌf/

Phonetic Spelling: kuhf

🔍 **Suture**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/suture>

IPA: /ˈsuː.tʃə/

Phonetic Spelling: soo-chur

🔍 **Trachea**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/trachea>

IPA: /ˈtreɪ.ki.ə/

Phonetic Spelling: tray-kee-uh

🔍 **Thymus**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/thymus>

IPA: /'θaɪ.məs/

Phonetic Spelling: thigh-muhs

🔍 **Hemostats**

Pronunciation link:

<https://www.howtopronounce.com/hemostats>

IPA: /'hi:.mə.stæts/

Phonetic Spelling: hee-muh-stats

🔍 **Decubitus** (as in “right lateral decubitus”)

Pronunciation link:

<https://www.howtopronounce.com/decubitus>

IPA: /dɪ'kju:.bɪ.təs/

Phonetic Spelling: dih-kyoo-buh-tuhs

🔍 **Eosin** (from “Hematoxylin and Eosin staining”)

Pronunciation link:

<https://www.merriam-webster.com/dictionary/eosin>

IPA: /'i:.ə.sɪn/

Phonetic Spelling: ee-uh-sin

🔍 **Alveolar**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/alveolar>

IPA: /æl'vi:.ə.lə/

Phonetic Spelling: al-vee-uh-ler

🔍 **Ligation**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/ligation>

IPA: /laɪ'geɪ.jən/

Phonetic Spelling: ly-gay-shuhn

🔍 **Cannula**

Pronunciation link:

<https://www.merriam-webster.com/dictionary/cannula>

IPA: /'kæn.jə.lə/

Phonetic Spelling: kan-yuh-luh