

Author Questionnaire

- 1. Microscopy:** Does your protocol require the use of a dissecting or stereomicroscope for performing a complex dissection, microinjection technique, or something similar? **NO**
- 2. Software:** Does the part of your protocol being filmed include step-by-step descriptions of software usage? **Yes, all done**
- 3. Filming location:** Will the filming need to take place in multiple locations? **No**

Current Protocol Length

Number of Steps: 16

Number of Shots: 42 (41 SC)

Introduction

Videographer: Obtain headshots for all authors available at the filming location.

- 1.1. **Gang Liu:** Our research evaluates three lymph node staging systems in colorectal signet ring cell carcinoma using machine learning and competing risk models to optimize prognostic accuracy and survival prediction.

1.1.1. INTERVIEW: Named talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B-roll: 2.3.1*

What technologies are currently used to advance research in your field?

- 1.2. **Jinyan Jia:** Bioinformatics methods, including machine learning, competing risk models and Kaplan-Meier survival estimation, are used to enhance survival prediction and lymph node classification accuracy.

1.2.1. INTERVIEW: Named talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B-roll: 4.1.1*

What research questions will your laboratory focus on in the future?

- 1.3. **Zixuan Yu:** Extending follow-up periods, validating in diverse populations, refining prognostic nomograms, and exploring molecular traits of colorectal signet ring cell carcinoma to enhance clinical decision-making tools.

1.3.1. INTERVIEW: Named talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B-roll: 4.2.1*

Videographer: Obtain headshots for all authors available at the filming location.

Protocol

2. Data Acquisition for Modelling

Demonstrator: Jinyan Jia

- 2.1. To begin, download and install SEER (*S-E-E-R*) [1], then obtain the statistics 8.4.3 software from the SEER database website [2]. Log into the software and click on **Case List Session** followed by **Data** [3] and select the **Incidence SEER Research Plus Data, 17 Registries, Nov 2022 Sub (2000-2020)** (*two thousand to twenty twenty*) database [4].
 - 2.1.1. WIDE: Talent taking a seat at the computer table.
 - 2.1.2. SCREEN: 2.1.2 00:05-00:18.
 - 2.1.3. SCREEN: 2.1.3.
 - 2.1.4. SCREEN: 2.1.4.
- 2.2. Now, click on **Selection** followed by **Edit** and choose **Race, Sex, Year of diagnosis** equal to '2004' through '2015' [1]. Then, select **Site recode ICD-O-3 WHO 2008** (*site-recode-I-C-D-Oh-3-W-H-Oh-two thousand and eight*) [2].
 - 2.2.1. SCREEN: 2.2.1.
 - 2.2.2. SCREEN: 2.2.2.
- 2.3. Click on **Table**, and in the available variables interface, select all the diagnosis details required [1].
 - 2.3.1. SCREEN: 2.3.1
- 2.4. Then, click on **Output**, name the data, [1] and click on **Execute** to output and save the data [2].
 - 2.4.1. SCREEN: 2.4.1.
 - 2.4.2. SCREEN: 2.4.2.
- 2.5. Next, open the X-tile software, click on **File** and choose **Open** [1]. Select the data file to import it into the software [2]. Once the data is loaded, map the variable **Censor** corresponding to survival status, the **Survival time** and **Marker1** as the variable to be analyzed, ensuring the data matches correctly [3].
 - 2.5.1. SCREEN: 2.5.1. *Video editor: If possible blur the background and keep only the X-TILE software interface in focus*
 - 2.5.2. SCREEN: 2.5.2.
 - 2.5.3. SCREEN: 2.5.3.
- 2.6. Now, click on **Do** followed by **Kaplan-Meier** and **Marker1** to perform the Kaplan-Meier survival analysis [1] and generate the survival curve [2].

- 2.6.1. SCREEN: 2.6.1
- 2.6.2. SCREEN: 2.6.2. 00:03-00:06

NOTE: Please display step 3.1 here before 2.7

- 2.7. Then, randomly assign a total of 2,409 eligible patient data with SRCC to a training cohort number 1,686 and a validation cohort number 723 in a 7 to 3 ratio [1]. Use the provided code for random splitting [2].

- 2.7.1. SCREEN: 2.7.1..
- 2.7.2. SCREEN: 2.7.2.

3. Machine Learning Models Development and Verification

Download and install the required versions of R-Studio and R software [1]. Click on **New File** and select **R Script** to create a new R programming interface [2]. Then, enter the relevant code in the code editor and click on **Run** to execute the code [3]. **NOTE:**

Please move step 3.1 before 2.7

- 3.1.1. SCREEN: 3.1.1. **TXT: RStudio version: 2024.04.2+764; R software version: 4.4.1**
- 3.1.2. SCREEN: 3.1.2.
- 3.1.3. SCREEN: 3.1.3.

- 3.2. Use the provided code to screen the variables included in the machine learning models by Cox regression analysis [1]. Additionally, explore the impact of LODDS (*L-O-D-D-S*), LNR (*L-N-R*), and pN (*P-N*) staging on cancer-specific survival in SRCC patients [2-TXT].

- 3.2.1. SCREEN: 3.2.1.
- 3.2.2. SCREEN: 3.2.2.

TXT: The traindata.csv is data obtained from the SEER database

- 3.3. Use the code to compare the prognostic prediction abilities of three lymph node systems LODDS, LNR, and pN staging [1] across the training, validation, and external validation cohorts [2].

- 3.3.1. SCREEN: 3.3.1.
- 3.3.2. SCREEN: 3.3.2. 00:10-00:16

- 3.4. Then use the code to build an XGBoost (*X-G-Boost*) model [1] and generate bar graphs representing the relative importance of variables [2]. Generate receiver operating characteristic curves [3] and calibration curves to assess the performance of the three lymph node systems [4].

- 3.4.1. SCREEN: 3.4.1. 00:07-00:14
- 3.4.2. SCREEN: 3.4.2.
- 3.4.3. SCREEN: 3.4.3.
- 3.4.4. SCREEN: 3.4.4.

- 3.5. Next, employ the code to build a random forest model and generate bar graphs of the relative importance of variables [1]. Similarly, generate receiver operating characteristic curves [2] and calibration curves to evaluate and compare the three lymph node systems [3].
 - 3.5.1. SCREEN: 3.5.1.
 - 3.5.2. SCREEN: 3.5.2.
 - 3.5.3. SCREEN: 3.5.3.
- 3.6. With the appropriate code, build a neural network model [1] and produce bar graphs of the relative importance of variables [2]. Generate receiver operating characteristic and calibration curves to compare the predictive performance of the three lymph node systems [3].
 - 3.6.1. SCREEN: 3.6.1.
 - 3.6.2. SCREEN: 3.6.2.
 - 3.6.3. SCREEN: 3.6.3.
- 3.7. Then, perform univariate analysis [1] and plot the cumulative incidence function curve using the data.csv file [2]. Replace “Site” with other factors to perform univariate analysis for each factor [3].
 - 3.7.1. SCREEN: 3.7.1. 00:09-00:12
 - 3.7.2. SCREEN: 3.7.2.
 - 3.7.3. SCREEN: 3.7.3. 00:04-00:12
- 3.8. For multivariate analysis, apply the code [1] and visualize with data1.csv (data 1 C-S-V) [2].
 - 3.8.1. SCREEN: 3.8.1. 00:09-00:13
 - 3.8.2. SCREEN: 3.8.2. **TXT: Click on Export, then click Save as PDF**
- 3.9. Finally, plot the nomogram, receiver operating characteristic curve, and calibration curve [1]. Train the model using data from the training cohort and use validation and external validation cohort data to validate the model [2].
 - 3.9.1. SCREEN: 3.9.1. 00:06-00:19
 - 3.9.2. SCREEN: 3.9.2.

Results

4. Representative Results

- 4.1. Based on multivariate Cox regression analysis, LNR, LODDS, and pN staging were all significantly associated with cancer-specific survival in SRCC patients [1].
 - 4.1.1. LAB MEDIA: Figure 2. *Video editor: Highlight A, B, and C sequentially.*
- 4.2. LNR showed the highest importance in the RF and XGBoost models [1], while LODDS had the greatest predictive ability in the NN model, suggesting LODDS as the most reliable LN system overall [2].
 - 4.2.1. LAB MEDIA: Figure 3. *Video editor: Focus on the panels A, B and highlight LNR bar*
 - 4.2.2. LAB MEDIA: Figure 3. *Video editor: Focus on the panels C and highlight "LODDS" bar in C.*
- 4.3. The XGBoost, RF, and NN models achieved high predictive accuracy with AUC values ranging from 0.777 to 0.851 [1] and calibration curves that aligned closely with the 45-degree line, confirming model reliability [2].
 - 4.3.1. LAB MEDIA: Figure 4A–C.
 - 4.3.2. LAB MEDIA: Figure 4A–F
- 4.4. Competing risk model analysis identified T staging, N staging, M staging, LODDS classification, and primary tumor location as independent prognostic factors [1].
 - 4.4.1. LAB MEDIA: Figure 5. *Video editor: Sequentially Highlight the graphs.*
- 4.5. The competing risk nomogram demonstrated accurate 1-, 3-, and 5-year cancer-specific survival predictions [1], supported by well-aligned calibration and ROC curves with AUCs above 0.75 [2].
 - 4.5.1. LAB MEDIA: Figure 6A *Video editor: Focus on the lines starting with "1-year, 3-year and 5-year"*
 - 4.5.2. LAB MEDIA: Figure 6E, F, G

Pronunciation guide

1. SEER

Pronunciation link:

<https://www.merriam-webster.com/dictionary/seer>

IPA: /'sɪr/

Phonetic Spelling: seer

2. Incidence

Pronunciation link:

<https://www.merriam-webster.com/dictionary/incidence>

IPA: /'ɪnsɪdəns/

Phonetic Spelling: in-suh-dns

3. Registry

Pronunciation link:

<https://www.merriam-webster.com/dictionary/registry>

IPA: /'rɛdʒɪstri/

Phonetic Spelling: reh-juh-stree

4. ICD-O-3

Pronunciation link:

No confirmed link found (ICD is commonly read as initials: I-C-D; "O" as "oh"; 3 as "three")

IPA: /ˌaɪ.sɪ.di.oo.'θriː/

Phonetic Spelling: eye-cee-dee-oh-three

5. WHO (as World Health Organization abbreviation)

Pronunciation link:

<https://www.merriam-webster.com/dictionary/WHO>

IPA: /'duː/ or /'huː/ (commonly pronounced as initials "W-H-O" or said as "who")

Phonetic Spelling: double-yoo-aych-oh

6. X-tile

Pronunciation link:

No confirmed link found

IPA: /'eks.taɪl/

Phonetic Spelling: eks-tile

7. Censor

Pronunciation link:

<https://www.merriam-webster.com/dictionary/censor>

IPA: /'sensə/

Phonetic Spelling: sen-ser

8. Kaplan-Meier

Pronunciation link:

<https://www.howtopronounce.com/kaplan-meier>

IPA: /'kæplən 'maɪə/

Phonetic Spelling: kap-luhn my-er

9. SRCC (Signet Ring Cell Carcinoma - abbreviated)

Pronunciation link:

No confirmed link found (pronounced as initials: S-R-C-C)

IPA: /,ɛs.ɑːr.siː.siː/

Phonetic Spelling: ess-ar-see-see

10. R-Studio

Pronunciation link:

No confirmed link found (commonly spoken as “R Studio”)

IPA: /ɑr 'stuːdiou/

Phonetic Spelling: ar stoo-dee-oh

11. Cox regression

Pronunciation link (for 'Cox'):

<https://www.merriam-webster.com/dictionary/Cox>

IPA: /kaks/

Phonetic Spelling: koks

Pronunciation link (for 'regression'):

<https://www.merriam-webster.com/dictionary/regression>

IPA: /rɪˈɡrɛʃən/

Phonetic Spelling: ri-gre-shun

12. LODDS

Pronunciation link:

No confirmed link found (pronounced as initials: L-O-D-D-S)

IPA: /ˌɛl.oʊ.diː.diː.ɛs/

Phonetic Spelling: el-oh-dee-dee-ess

13. LNR

Pronunciation link:

No confirmed link found (pronounced as initials: L-N-R)

IPA: /ˌɛl.ɛn.ər/

Phonetic Spelling: el-en-ar

14. pN staging

Pronunciation link:

No confirmed link found (pronounced as initials: P-N)

IPA: /ˌpiːˈɛn/

Phonetic Spelling: pee-en

15. XGBoost

Pronunciation link:

<https://www.howtopronounce.com/xgboost>

IPA: /'eks.dʒi:.bu:st/

Phonetic Spelling: eks-jee-boost

16. Receiver Operating Characteristic

Pronunciation link (for 'receiver'):

<https://www.merriam-webster.com/dictionary/receiver>

IPA: /rɪ'si:və/

Phonetic Spelling: rih-see-ver

Pronunciation link (for 'characteristic'):

<https://www.merriam-webster.com/dictionary/characteristic>

IPA: /,kærəktə'rɪstɪk/

Phonetic Spelling: keh-ruh-k-tuh-ris-tik

17. Neural Network

Pronunciation link (for 'neural'):

<https://www.merriam-webster.com/dictionary/neural>

IPA: /'nʊrəl/ or /'njʊrəl/

Phonetic Spelling: nyoo-ruhl or noo-ruhl

18. Nomogram

Pronunciation link:

<https://www.merriam-webster.com/dictionary/nomogram>

IPA: /'nɑ:məˌgræm/

Phonetic Spelling: noh-muh-gram