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Title: A Model of Acute Lung Injury Following Visceral Ischemia-Reperfusion by Supra-Coeliac Aortic Cross Clamping in Rats

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Author Questionnaire

1. We have marked your project as author-provided footage, meaning you film the video yourself and provide JoVE with the footage to edit. JoVE will not send the videographer. Please confirm that this is correct.

√ Correct

2. Microscopy: Does your protocol require the use of a dissecting or stereomicroscope for performing a complex dissection, microinjection technique, or something similar? **YES**, all done

SCOPE: 2.6.1, 2.9.1-2.9.3

- **3. Software:** Does the part of your protocol being filmed include step-by-step descriptions of software usage? **NO**
- **4. Proposed filming date:** To help JoVE process and publish your video in a timely manner, please indicate the <u>proposed date that your group will film</u> here: **08/31/2025 AUTHORS: Please film the introduction section**

When you are ready to submit your video files, please contact our Content Manager, <u>Utkarsh Khare</u>.

Current Protocol Length

Number of Steps: 17 Number of Shots: 33



Introduction

- 1.1. <u>Mickael Palmier:</u> We present a reproducible rat surgical model of acute lung injury after supra-celiac aortic cross-clamping, enabling detailed study of ischemia—reperfusion mechanisms in pulmonary injury [1].
 - 1.1.1. INTERVIEW: Named talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B. roll: 2.11*

What advantage does your protocol offer compared to other techniques?

- 1.2. <u>Mickael Palmier:</u> Our protocol is feasible, reproducible, low-cost, and easy to learn, enabling rapid implementation, biomarker exploration, and testing of diverse pharmacological interventions [1].
 - 1.2.1. INTERVIEW: Named talent says the statement above in an interview-style shot, looking slightly off-camera.

What research questions will your laboratory focus on in the future?

- 1.3. <u>Mickael Palmier:</u> Our surgical model of acute lung injury will enable us to investigate various pharmacological strategies aimed at reducing the severity of pulmonary damage following aortic cross-clamping [1].
 - 1.3.1. INTERVIEW: Named talent says the statement above in an interview-style shot, looking slightly off-camera. *Suggested B. roll: 2.9*



Ethics Title Card

This research has been approved by the Regional University Ethics Committee at Rouen University Hospital



Protocol

2. Microsurgical Induction of Aortic Ischemia-Reperfusion Injury and Analgesic Management in Rats

Demonstrator: Myriam Cherel

- 2.1. To begin, place an anesthetized rat in dorsal recumbency and immobilize the front and hind limbs with adhesive tape while maintaining physiological joint motion ranges [1-TXT]. Transfer the rat onto a warming pad set to 37 degrees Celsius and monitor rectal temperature [2].
 - 2.1.1. LAB MEDIA: Protocol.3.1.mov 00:00-00:30 TXT: Analgesia: Buprenorphine (0.05 mg/kg, SQ) 1 h before the procedure; Apply Lidocaine hydrochloride gel 1% to the skin; Anesthesia:; 5% isoflurane for induction; 2-2.5% isoflurane for maintenance
 - 2.1.2. LAB MEDIA: Protocol.3.1.mov. 00:36-00:49
- 2.2. Apply ophthalmic eye ointment to the eyes to prevent dryness [1]. After shaving the abdominal fur, thoroughly disinfect the area using povidone-iodine solution and alcohol [2].

2.2.1. LAB MEDIA: Protocol.3.3.1.mov. 00:13-00:23

2.2.2. LAB MEDIA: Protocol.3.3.2.mov 00:00-00:14

- 2.3. To perform the surgery, make a skin incision at the center of the abdomen to expose the linea alba [1-TXT]. Incise the abdominal muscle centered on the linea alba to prevent bleeding from surrounding muscle tissues [2].
 - 2.3.1. LAB MEDIA: Protocol_4.2.2.mov 00:00-00:20

TXT: Perform surgery under a microsurgical loupe, if possible

- 2.3.2. LAB MEDIA: Protocol_4.2.2.mov 00:56-01:20
- 2.4. Maintain the abdominal wall on both sides using self-retaining retractors [1]. Examine the cavity with cotton-tipped buds for any anomalies [2], then gently mobilize the viscera, stomach, spleen, and liver to the right side of the abdomen to expose the supraceliac aorta [3].



2.4.1. LAB MEDIA: Protocol 4.2.2.mov 02:15-02:28

2.4.2. LAB MEDIA: Protocol 4.2.2.mov 02:36-02:50

2.4.3. LAB MEDIA: Protocol 4.3.2.mov 00:24-00:39, 00:53-01:04

2.5. Keep all viscera inside the abdominal cavity using a compress folded lengthwise and soaked in warm saline solution at 37 degrees Celsius [1]. Then cover all exposed viscera with a compress soaked in warm saline solution at 37 degrees Celsius to prevent hypothermia and dehydration [2].

2.5.1. LAB MEDIA: Protocol 4.3.2.mov 01:11-01:26

2.5.2. LAB MEDIA: Protocol 4.3.2.mov 00:40-00:52

2.6. Identify and encircle the supra-celiac aorta with a 5-0 (*Five-Oh*) monofilament suture using dissecting and ligature forceps [1-TXT].

2.6.1. SCOPE: Protocol 4.4.3.mov00:00-00:20

TXT: Verify the origins of the celiac trunk and the underlying mesenteric artery

- 2.7. Identify the subrenal inferior vena cava through the transparent retroperitoneum [1-TXT].
 - 2.7.1. LAB MEDIA: Protocol 4.4.5.mov 00:00-00:07

TXT: Release the anterior surface of the subrenal IVC to direct IV administration of pharmacological agents

- 2.8. For the intravenous injection, use a 30-gauge needle and apply a small piece of hemostatic compress at the injection site to achieve hemostasis [1-TXT].
 - 2.8.1. LAB MEDIA: Protocol 4.10.1.mov 03:06-03:14.

TXT: Administer injection before clamping for pharmacological treatment or after for post-conditioning

2.9. Now clamp the celiac aorta with an atraumatic microvascular clamp for 40 minutes [1]. Verify the clamp's effectiveness by confirming the disappearance of aortic pulsations [2] and discoloration of the viscera and limbs multiple times during clamping [3].

2.9.1. SCOPE: Protocol 4.6.1.5.mov 00:04-00:20

2.9.2. SCOPE: Protocol_4.6.1.5.mov 01:50-02:03

2.9.3. SCOPE: Protocol_4.6.1.5.mov 00:59-01:16.



2.10. Close the abdomen partially using three simple 5-0 braided absorbable sutures to reduce water and heat loss [1]. Then cover the incision with a sterile compress soaked in warm saline for the duration of ischemia [2].

2.10.1. LAB MEDIA: Protocol_4.6.2.mov 00:27-00:57,02:10-02:20

2.10.2. LAB MEDIA: Protocol 4.6.2.mov 02:27-02:35

2.11. At the end of the ischemic period, reopen the abdomen and expose the aorta [1]. Remove the vascular clamp and verify resumption of aortic pulsations and recoloration of the viscera and limbs. This marks the beginning of the reperfusion period [2].

2.11.1. LAB MEDIA: Protocol_4.7.1mov 00:15,00:38-00:50

2.11.2. LAB MEDIA: Protocol_4.7.1.mov 00:55-01:33.

2.12. Return the viscera to their original position [1]. Then close the abdominal wall with a 5-0 braided absorbable suture on the linea alba [2]. Use a 5-0 monofilament absorbable suture on the skin [3].

2.12.1. LAB MEDIA: Protocol 4.8.1.1.mov 00:00-00:12

2.12.2. LAB MEDIA: Protocol 4.8.1.1.mov 00:18-00:36, 02:18-02:30

2.12.3. LAB MEDIA: Protocol_4.8.1.2.mov 00:40-00:50, 02:15-02:22

2.13. Place the rat in the left lateral decubitus position to prevent compression of the inferior vena cava during hypovolemia [1]. Then transfer it into an individual cage under a heat lamp until it awakens, which occurs approximately 10 minutes later [2].

2.13.1. LAB MEDIA: Protocol 4.9.0.mov 00:00-00:06

2.13.2. LAB MEDIA: Protocol 4.9.1.mov 00:00-00:08

3. Blood and Tissue Collection, Molecular, Histological, and Gravimetric Analyses Following Aortic Ischemia-Reperfusion

3.1. For blood sampling, directly puncture the inferior vena cava below the renal veins using a 5-milliliter syringe equipped with a 23-gauge needle [1].

3.1.1. LAB MEDIA: Protocol 5.1.1.mov 00:00-00:11

3.2. After the animal has been euthanized, harvest the left kidney [1]. Then harvest the left



hepatic lobe [2], and the distal small intestine 1 centimeter before the cecum through the open abdomen [3]. Quickly perform a median sternotomy to expose and identify the heart and lungs [4].

3.2.1.	LAB MEDIA: Protocol_5.2.1mov	00:12-00:40
3.2.2.	LAB MEDIA: Protocol_5.2.1mov	05:44-05:57
3.2.3.	LAB MEDIA: Protocol_5.2.1mov	01:16-01:30,
3.2.4.	LAB MEDIA: Protocol 5.2.1mov	02:40-02:50,03:34-03:45.

- 3.3. Free the left lung from its ligament and remove it entirely for histological analysis [1]. Then free the right multilobed lung from its ligaments and isolate the anterior lobe for nucleic acid and protein analysis [2-TXT].
 - 3.3.1. LAB MEDIA: Protocol 5.2.1..mov 03:48-04:06, 04:15-04:23.
 - 3.3.2. LAB MEDIA: Protocol_5.2.1..mov 05:10-05:33. **TXT: Store the** remaining portion in a cup with warm physiological serum for gravimetry
- 3.4. Extract the heart as a whole block, including sections of the aorta, pulmonary artery, and both vena cavae [1-TXT].
 - 3.4.1. LAB MEDIA: Protocol 5.2.1..mov 04:54-05:08.

TXT: Sample a piece of the left ventricle for nucleic acid and protein analysis



Results

4. Results

- 4.1. Blood gas analysis following supracoeliac aortic cross clamping showed reduced pH [1], decreased partial oxygen pressure [2], increased partial carbon dioxide pressure [3], and lower bicarbonate levels compared to normal ranges [4].
 - 4.1.1. LAB MEDIA: Table 2. Video editor: Highlight the row labeled "pH" in the column "After Supracoeliac Aortic cross clamping"
 - 4.1.2. LAB MEDIA: Table 2. Video editor: Highlight the row labeled "PaO₂ (mmHg)" in the column "After Supracoeliac Aortic cross clamping"
 - 4.1.3. LAB MEDIA: Table 2. Video editor: Highlight the row labeled "PaCO₂ (mmHg)" in the column "After Supracoeliac Aortic cross clamping"
 - 4.1.4. LAB MEDIA: Table 2. Video editor: Highlight the row labeled "HCO₃⁻ (mmol/L)" in the column "After Supracoeliac Aortic cross clamping"
- 4.2. The histopathologic lung injury score increased markedly in the supracoeliac aortic cross clamping group [1]. Polymorphonuclear neutrophils per square millimeter were significantly elevated after supracoeliac aortic cross clamping [2].
 - 4.2.1. LAB MEDIA: Table 3 Video editor: Highlight the "Histopathologic lung injury score" row and column "Supracoeliac Aortic cross clamping"
 - 4.2.2. LAB MEDIA: Table 3 Video editor: Highlight the row labeled "PNN" for column "
 Supracoeliac Aortic cross clamping"
- 4.3. Intravascular leukocyte counts were significantly higher in the aortic cross clamping group compared to the sham group [1].
 - 4.3.1. LAB MEDIA: Table 3 Video editor: Highlight the row labeled "Intravascular cells" for column "Supracoeliac Aortic cross clamping"
- 4.4. Pulmonary inflammation markers were also significantly elevated after supracoeliac aortic cross clamping [1]. Additionally, pulmonary edema, as measured by gravimetric analysis, was found to be increased after aortic clamping [2].
 - 4.4.1. LAB MEDIA: Table 4. Video editor: Highlight the rows labeled "TNF-alpha" and "IL-1 β" for column "Supracoeliac Aortic cross clamping"
 - 4.4.2. LAB MEDIA: Figure 3 Video Editor: Please highlight the red squares



Pronunciation Guide:

Recumbency

Pronunciation link:

https://www.merriam-webster.com/dictionary/recumbency

IPA: /rɪˈkʌmbənsi/

Phonetic Spelling: ri-kum-buhn-see

Physiological

Pronunciation link:

https://www.merriam-webster.com/dictionary/physiological

IPA: /ˌfɪziəˈlɑːdʒɪkəl/

Phonetic Spelling: fiz-ee-uh-lah-juh-kl

Ophthalmic

Pronunciation link:

https://www.merriam-webster.com/dictionary/ophthalmic

IPA: /af'θælmɪk/

Phonetic Spelling: ahf-thal-mik

Povidone-iodine

Pronunciation link:

https://www.merriam-webster.com/dictionary/povidone-iodine

IPA: /ˈpoʊ.vɪˌdoʊn ˈaɪ.əˌdaɪn/

Phonetic Spelling: poh-vi-dohn eye-uh-dine

Linea alba

Pronunciation link:

https://www.howtopronounce.com/linea-alba

IPA: /ˈlɪniə ˈælbə/

Phonetic Spelling: lin-ee-uh al-buh

Viscera

Pronunciation link:

https://www.merriam-webster.com/dictionary/viscera

IPA: /ˈvɪsərə/

Phonetic Spelling: vis-er-uh

Supra-celiac (as in supra-celiac aorta)

Pronunciation link:

https://www.howtopronounce.com/supraceliac

IPA: /ˌsuːprəˈsiːliæk/

Phonetic Spelling: soo-pruh-see-lee-ak



Mesenteric

Pronunciation link:

https://www.merriam-webster.com/dictionary/mesenteric

IPA: /ˌmɛzənˈtɛrɪk/ or /ˌmɛsənˈtɛrɪk/

Phonetic Spelling: mez-en-ter-ik or mess-en-ter-ik

Retroperitoneum

Pronunciation link:

https://www.howtopronounce.com/retroperitoneum

IPA: /ˌrɛtroʊˌpɛrətnˈiəm/

Phonetic Spelling: reh-troh-peh-rih-tee-nee-uhm

Intravenous

Pronunciation link:

https://www.merriam-webster.com/dictionary/intravenous

IPA: /ˌɪntrəˈviːnəs/

Phonetic Spelling: in-truh-vee-nuhs

Hemostasis

Pronunciation link:

https://www.merriam-webster.com/dictionary/hemostasis

IPA: / hiːmoʊˈsteɪsɪs/

Phonetic Spelling: hee-moh-stay-sis

Atraumatic

Pronunciation link:

https://www.merriam-webster.com/dictionary/atraumatic

IPA: / eɪtrɔːˈmætɪk/

Phonetic Spelling: ay-traw-mat-ik

Ischemia

Pronunciation link:

https://www.merriam-webster.com/dictionary/ischemia

IPA: /ɪˈskiːmiə/

Phonetic Spelling: ih-skee-mee-uh

Reperfusion

Pronunciation link:

https://www.merriam-webster.com/dictionary/reperfusion

IPA: /ˌriːpərˈfjuːʒən/

Phonetic Spelling: ree-per-fyoo-zhun



Decubitus (as in lateral decubitus position)

Pronunciation link:

https://www.merriam-webster.com/dictionary/decubitus

IPA: /dɪˈkjuːbɪtəs/

Phonetic Spelling: dih-kyoo-bih-tuhs

Cecum

Pronunciation link:

https://www.merriam-webster.com/dictionary/cecum

IPA: /ˈsiːkəm/

Phonetic Spelling: see-kuhm

Sternotomy

Pronunciation link:

https://www.howtopronounce.com/sternotomy

IPA: /stərˈnɑːtəmi/

Phonetic Spelling: ster-nah-tuh-mee

Histological

Pronunciation link:

https://www.merriam-webster.com/dictionary/histological

IPA: / hɪstəˈlaːdʒɪkəl/

Phonetic Spelling: his-tuh-lah-jih-kl

Gravimetric

Pronunciation link:

https://www.merriam-webster.com/dictionary/gravimetric

IPA: / grævə metrik/

Phonetic Spelling: grav-uh-meh-trik

Supracoeliac (alternate spelling in Results)

Pronunciation link:

https://www.howtopronounce.com/supracoeliac

IPA: /ˌsuːprəˈsiːliæk/

Phonetic Spelling: soo-pruh-see-lee-ak

Histopathologic

Pronunciation link:

https://www.merriam-webster.com/dictionary/histopathologic

IPA: / histog pæθəˈlaːdʒik/

Phonetic Spelling: his-toh-path-uh-lah-jik



Polymorphonuclear

Pronunciation link:

https://www.merriam-webster.com/dictionary/polymorphonuclear

IPA: / paːlɪˌmɔːrfəˈnuːkliər/

Phonetic Spelling: pah-lee-mor-foh-noo-klee-er

Neutrophils

Pronunciation link:

https://www.merriam-webster.com/dictionary/neutrophil

IPA: /ˈnuːtrəˌfɪl/

Phonetic Spelling: noo-truh-fil

Pulmonary

Pronunciation link:

https://www.merriam-webster.com/dictionary/pulmonary

IPA: /ˈpʊlməˌnɛri/ or /ˈpʌlməˌnɛri/

Phonetic Spelling: pull-muh-neh-ree or puhl-muh-neh-ree

Edema

Pronunciation link:

https://www.merriam-webster.com/dictionary/edema

IPA: /ɪˈdiːmə/

Phonetic Spelling: ih-dee-muh