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The diagram illustrates a five-step experimental workflow:

- Submerged BME2 organoid culture:** A petri dish containing a dome-shaped organoid culture.
- Dissociation & counting:** The organoid is dissociated into a suspension in a tube, which is then counted using a hemacytometer.
- Seed organoid plate (6 o'clock position of dome):** The dissociated organoids are seeded into a 96-well plate.
- Treat with escalating doses:** The seeded organoids are treated with escalating doses of a drug (represented by blue and white capsules) using a multi-channel pipette.
- Luminescence-based viability assay:** The treated organoids are analyzed using a luminescence-based viability assay on a plate reader.

Time intervals are indicated: **~ 7 days** between seeding and treatment, and **5 days** between treatment and the final viability assay.

A diagram of a 96-well plate with 12 columns and 8 rows. The columns are numbered 1 to 12 at the top, and the rows are labeled A to H on the left. The wells contain colored circles representing a color gradient. The color starts as blue in column 1 and transitions through light blue, white, and yellow to orange in column 12. The transition is most pronounced in the middle rows (D, E, F, G) and least pronounced in the top (A, B) and bottom (H) rows.