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**Title: Air-Inflation of Murine Lungs with Vascular Perfusion-Fixation**

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## Author Questionnaire

- 1. Microscopy:** Does your protocol demonstrate the use of a dissecting or stereomicroscope for performing a complex dissection, microinjection technique, or similar? **N**
- 2. Software:** Does the part of your protocol being filmed demonstrate software usage? **N**
- 3. Interview statements:** Considering the Covid-19-imposed mask-wearing and social distancing recommendations, which interview statement filming option is the most appropriate for your group? **Please select one.**



Statements are read by JoVE VO talent

- 3. Filming location:** Will the filming need to take place in multiple locations (greater than walking distance)? **No**

### Protocol Length

Number of Shots: **47**

*Videographer: When filming the protocol, please avoid filming the faces of any of the demonstrators*

# Introduction

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## 1. Introductory Interview Statements

**NOTE to JoVE VO Talent: Please record all introduction and conclusion statements**

### REQUIRED:

- 1.1. This is an inexpensive method that combines the advantages of air inflation and vascular perfusion-fixation for preserving lungs for structural and functional analyses. [1].
  - 1.1.1. *4.7.2 for 'vascular perfusion-fixation'*
- 1.2. The main advantage of this technique is the preservation of both cell morphology and location within the airspaces of the lung. [1].
  - 1.2.1. *4.2.2.*

### OPTIONAL:

- 1.3. Care should be taken while placing the Luer stub adapter into the trachea and the perfusion needle into the right ventricle to ensure adequate inflation and fixation.
  - 1.3.1. *Suggested B-roll: 3.7.1 for 'luer stub adapter'*

### Ethics Title Card

- 1.4. Procedures involving animal subjects have been approved by the Institutional Animal Care and Use Committee (IACUC) at National Jewish Health.

# Protocol

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## 2. Air-Inflation Apparatus Preparation

*Videographer: When filming the protocol, please avoid filming the faces of any of the demonstrators*

- 2.1. To set up the air-inflation apparatus, place a syringe for the water column into a ring holder [1] and measure a vertical height of 25 centimeters from the animal platform to 25 centimeters on the water column [2].
  - 2.1.1. WIDE: Talent placing syringe into ring holder
  - 2.1.2. Talent measuring/marking height
- 2.2. Attach the end of the water column tube to the stopcock on the air chamber [1] and attach a tube from the female Luer of the air chamber to the stopcock on the animal processing container [2].
  - 2.2.1. Talent attaching water column tube to stopcock
  - 2.2.2. Talent attaching female Luer tube to stopcock
- 2.3. Confirm that the cap to the air chamber [1] and the stopcock on the outside of the animal processing container are closed [2] and that the stopcock on the tubing leading from the water column to the air-inflation chamber is open [3].
  - 2.3.1. Cap closure being checked
  - 2.3.2. Stopcock closure being checked
  - 2.3.3. Stopcock being opened/shot of open stopcock
- 2.4. Then fill the syringe with water to the 25-centimeter mark [1]. Water will flow through the syringe and tubing into the air chamber [2]. Once the pressure has equalized, the water will stop flowing [3-TXT].
  - 2.4.1. Talent filling syringe

2.4.2. Water leaving syringe through tubing

2.4.3. Water stopping flowing **TEXT: See text for water leakage correction details**

### **3. Animal Preparation**

3.1. To prepare the lungs for inflation, make one lateral incision in the abdominal wall below the rib cage of a euthanized mouse [1] and a second, lateral, abdominal wall incision above the hips [2].

3.1.1. WIDE: Talent making incision *Videographer: More Talent than mouse in shot*

3.1.2. Shot of first incision, then second incision being made

3.2. Cut along the midline from the inferior incision toward the superior incision [1] and use blunt scissors to carefully make an incision into the lateral side of the diaphragm. The lungs should collapse as soon as the diaphragm is punctured [2-TXT].

3.2.1. Incision being made between incisions

3.2.2. Incision being made in diaphragm/lungs collapsing **TEXT: Caution: Do not puncture lungs**

3.3. Cut transversely along the diaphragm to open the thoracic cavity [1] and cut superiorly along the sternum from the xiphoid process to the jugular notch and laterally above the rib cage to fully expose the heart and lungs [2]. *Videographer: This step is important!*

3.3.1. Diaphragm being cut

3.3.2. Incision(s) being made/heart and lungs being exposed

3.4. Pin down the sides of the ribcage [1] and make a midline incision in the neck above the trachea [2].

3.4.1. Pin(s) being placed

3.4.2. Incision being made

- 3.5. Remove the skin, muscle, thyroid gland, and connective tissue surrounding the trachea [1] and use curved forceps to slide two pieces of suture under the posterior trachea [2].

- 3.5.1. Tissue being removed

- 3.5.2. Suture/thread being placed

- 3.6. Use one piece of suture to hold the inflation Luer-stub adapter in place [1] and use an 18-gauge x 1-inch needle to make a small hole in the trachea [2]. *Videographer: This step is important!*

- 3.6.1. Suture being placed

- 3.6.2. Hole being made

- 3.7. Insert a 20-gauge Luer-stub adapter into the hole [1] and tie the suture around the trachea immediately distal to where the Luer-stub adapter enters the hole [2]. *Videographer: This step is important!*

- 3.7.1. Adapter being inserted

- 3.7.2. Suture being tied

- 3.8. Then transfer the animal to the animal processing container [1] and attach the Luer-stub adapter to the female Luer on the inside of the animal processing container [2].

- 3.8.1. Talent placing animal into container *Videographer: More Talent than mouse in shot*

- 3.8.2. Adapter being attached to Luer

#### **4. Lung Air-Inflation, Perfusion, and Fixation**

- 4.1. To inflate the lungs, place the 25-gauge x 5/8-inch needle attached to the perfusion apparatus tubing into the right ventricle of the heart [1] and cut the abdominal aorta to allow blood to drain from the heart and to promote the flow of perfusate through the lungs [2]. *Videographer: This step is important!*

- 4.1.1. WIDE: Talent placing needle *Videographer: More Talent than mouse in shot*
- 4.1.2. Shot of needle in place, then aorta being cut
- 4.2. Open the stopcock on the outside of the animal processing container [1] and inflate the lungs at 25 centimeters of water for 5 minutes [2], while that the water level in the syringe does not decrease too quickly [3]. *Videographer: This step is important!*
  - 4.2.1. Stopcock being opened
  - 4.2.2. Lungs being inflated
  - 4.2.3. Water level decreasing
- 4.3. During the last minute of the lung inflation, turn on the peristaltic pump to a flow rate of 10 milliliters/minute [1]. Heparin solution should flow from the bottle through the tubing into the animal [2].
  - 4.3.1. Talent turning on pump *Videographer: This step is important!*
  - 4.3.2. Solution flowing into animal
- 4.4. After inflating the lungs for 5 minutes, turn off the pump [1] and switch the perfusate from heparin to fixative [2].
  - 4.4.1. Talent turning off pump
  - 4.4.2. Talent selecting fixative
- 4.5. Lower the water column syringe to 20 centimeters. It is normal for air bubbles to move within the water column as the pressure changes [1].
  - 4.5.1. Talent lowering water column
- 4.6. Check the water level in the syringe. It should be at the 25-centimeter mark [1-TXT].
  - 4.6.1. Shot of water level in syringe **TEXT: Add more H<sub>2</sub>O to syringe as necessary**

- 4.7. Then wait 1 minute to allow the lungs to deflate from 25 to 20 centimeters of water pressure [1] before restarting the pump at a 6.5 milliliter/minute flow rate for 10-15 minutes of vascular fixative perfusion [2].

- 4.7.1. Lungs deflating

- 4.7.2. Fixative being perfused into lungs

## **5. Lung Extraction**

- 5.1. For extraction of the inflated and fixed lung tissue, tightly tie the second piece of thread around the trachea distal to the Luer-stub adapter [1] and remove the Luer-stub adapter from the trachea [2].

- 5.1.1. WIDE: Talent tying suture *Videographer: More Talent than mouse in shot*

- 5.1.2. Shot of tied suture, then adapter being removed

- 5.2. Remove the needle from the heart [1] and use blunt scissors to cut the connective tissue posterior to the mediastinum to free the lungs and heart from the thoracic cavity, taking care to avoid puncturing the lungs [2].

- 5.2.1. Needle being removed

- 5.2.2. Tissue being dissected

- 5.3. Carefully remove the heart from the lungs [1] and place the lungs into 20-25 milliliters of fixative in a 50-milliliter conical tube [2].

- 5.3.1. Heart being removed

- 5.3.2. Talent placing lungs into tube

- 5.4. Pull the suture holding the trachea through the opening of the conical tube and secure the suture by the threads of the cap [1].

- 5.4.1. Suture being pulled/secured



- 5.5. Then invert the tube to ensure that the buoyant, air-inflated lungs remain fully submerged in fixative [1] and process the lungs for histologic studies according to standard protocols [2].

- 5.5.1. Tube being inverted

- 5.5.2. LAB MEDIA: Figure 4

## Results

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### 6. Results: Representative Lung Inflation, Extraction, and Histological Evaluation

- 6.1. When the diaphragm is entered during dissection [1], the integrity of the pleural space will be abolished and the lungs should collapse [2].

6.1.1. LAB MEDIA: Figure 2B

6.1.2. LAB MEDIA: Figure 2B *Video Editor: please emphasize collapsed lung (white tissue)*

- 6.2. Upon the application of 25 centimeters of water pressure [1], air will enter the lungs via the trachea and inflation should be easily observed [2].

6.2.1. LAB MEDIA: Figure 2C

6.2.2. LAB MEDIA: Figure 2C *Video Editor: please emphasize inflated lung (white tissue)*

- 6.3. Once the lungs have fully expanded [1], the inflation pressure can be decreased to 20 centimeters to keep the lungs inflated without over-distension [2].

6.3.1. LAB MEDIA: Figure 2D

6.3.2. LAB MEDIA: Figure 2D *Video Editor: please emphasize white tissue*

- 6.4. The lungs should remain inflated after tracheal ligation [1] and removal from the thorax [2].

6.4.1. LAB MEDIA: Figures 3A and 3B *Video Editor: please emphasize white tissue in Figure 3A*

6.4.2. LAB MEDIA: Figures 3A and 3B *Video Editor: please emphasize white tissue in Figure 3B*

- 6.5. Deflation can occur if the lungs are punctured during the animal preparation or lung extraction [1].

6.5.1. LAB MEDIA: Figure 3C *Video Editor: please emphasize white tissue*

- 6.6. Histological analysis of inflated lung sections [1] reveals that very few immune cells are present in the airway lumens of tissues fixed using traditional liquid-based inflation [2].

6.6.1. LAB MEDIA: Figure 4

6.6.2. LAB MEDIA: Figure 4 *Video Editor: please emphasize empty space in right Figure 4A image*

6.7. In contrast, inflammatory cells are preserved within the airspaces of tissues fixed via vascular perfusion with air-inflation [1].

6.7.1. LAB MEDIA: Figure 4 *Video Editor: please emphasize cells in empty space in right Figure 4B image/arrows in right Figure 4B image*

# Conclusion

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## 7. Conclusion Interview Statements

NOTE to JoVE VO Talent: Please record all introduction and conclusion statements

- 7.1. Following air-inflation with perfusion fixation, lung tissue may be embedded in formalin for tissue sectioning and subsequent analysis via histology techniques including staining, immunohistochemistry and immunofluorescent imaging. [1].

- 7.1.1. *5.5.1 for 'embedded in formalin'*