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Self-Assembly of Hybrid Lipid Membranes Doped with Hydrophobic Organic Molecules at the Water/Air Interface --Manuscript Draft--

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1 TITLE:

Self-Assembly of Hybrid Lipid Membranes Doped with Hydrophobic Organic Molecules at the
 Water/Air Interface

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KEYWORDS:

20 Lipid bilayers, organic molecule, thin film, water/air interface, self-assembly, hybrid lipid

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SUMMARY:

We report a protocol for producing a hybrid lipid membrane at the water/air interface by doping the lipid bilayer with copper (II) 2,9,16,23-tetra-tert-butyl-29H,31H-phthalocyanine (CuPc) molecules. The resulting hybrid lipid membrane has a lipid/CuPc/lipid sandwich structure. This protocol can also be applied to the formation of other functional nanomaterials.

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ABSTRACT:

Because of their unique properties, including an ultrathin thickness (3-4 nm), ultrahigh resistivity, fluidity and self-assembly ability, lipid bilayers can be readily functionalized and have been used in various applications such as bio-sensors and bio-devices. In this study, we introduced a planar organic molecule: copper (II) 2,9,16,23-tetra-tert-butyl-29H,31H-phthalocyanine (CuPc) to dope lipid membranes. The CuPc/lipid hybrid membrane forms at the water/air interface by self-assembly. In this membrane, the hydrophobic CuPc molecules are located between the hydrophobic tails of lipid molecules, forming a lipid/CuPc/lipid sandwich structure. Interestingly, an air-stable hybrid lipid bilayer can be readily formed by transferring the hybrid membrane onto a Si substrate. We report a straightforward method for incorporating nanomaterials into a lipid bilayer system, which represents a new methodology for the fabrication of biosensors and biodevices.

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INTRODUCTION:

- As essential frameworks of cell membranes, the interior of cells are separated from the exterior
- by a lipid bilayer system. This system consists of amphiphilic phospholipids, which are composed

of hydrophilic phosphoric ester "heads" and hydrophobic fatty acids "tails". Due to remarkable fluidity and self-assembly ability of lipid bilayers in aqueous environment^{1,2}, artificial lipid bilayers can be formed using simple methods^{3,4}. Various types of membrane proteins, such as ion channels, membrane receptors and enzymes, have been incorporated into the artificial lipid bilayer to mimic and study the functions of cell membranes^{5,6}. More recently, lipid bilayers have been doped with nanomaterials (e.g., metal nanoparticles, graphene, and carbon nanotubes) to form functional hybrid membranes⁷⁻¹³. A widely used method for forming such hybrid membranes involves the formation of doped lipid vesicles, which contain hydrophobic materials such as modified Au-nanoparticles⁷ or carbon nanotubes¹¹, and the resulting vesicles are then fused into planar supported lipid bilayers. However, this approach is complex and time-consuming, which limits the potential uses of such hybrid membranes.

In this work, lipid membranes were doped with organic molecules to produce hybrid lipid membranes that formed at the water/air interface by self-assembly. This protocol involves three steps: preparation of the mixed solution, formation of a hybrid membrane at the water/air interface, and the transfer of the membrane onto a Si substrate. Compared with other previously reported methods, the method described here is simpler and does not require sophisticated instrumentation. Using this method, air-stable hybrid lipid membranes with a larger area can be formed in a shorter time. The nanomaterial used in this study is a semiconducting organic molecule, copper (II) 2,9,16,23-tetra-tert-butyl-29H,31H-phthalocyanine (CuPc), which is widely used in a number of applications, including solar cells, photodetectors, gas sensors and catalysis^{14,15}. CuPc, a small organic molecule with a planar structure, has a high affinity for the "tails" of phospholipids duo to its hydrophobic characteristics. Other groups have reported that CuPc molecules can self-assemble on single-crystal surfaces with the formation of highly ordered structures^{16,17}. Therefore, it is highly possible that the CuPc molecules could be incorporated into the lipid bilayers through self-assembly.

We provide a detailed description of the procedures used to form membranes and provide some suggestions for smoothly implementing this procedure. In addition, we present some presentative results of the hybrid lipid membranes, and discuss potential applications of this method.

PROTOCOL:

1. Preparation of a hybrid solution

1.1. Wash four 4 mL disposable glass vials and screw caps (surface treatment: Teflon coating) in an ultrasonic bath for 10 min in distilled water (purified with a filtration system), followed by ethanol and chloroform, respectively. Dry the glass vials and caps in a stream of nitrogen gas.

1.2. In an anaerobic glove box, prepare a CuPc stock solution (10 mg/mL) in a washed glass vial by dissolving powdered CuPc in chloroform.

1.3. Filter the CuPc solution through a 0.2 μm polytetrafluoroethylene (PTFE) membrane.

1.4. Store the filtered solution in a washed glass vial filled with nitrogen, and seal the vial with parafilm. 1.5. Take the purchased 1,2-diphytanoyl-sn-glycero-3-phosphocholine (DPhPC) chloroform solution (25 mg/mL) out of the -80 °C refrigerator and allow it to warm to room temperature. 1.6. Stir the DPhPC solution using a vortex mixer at 2300 rpm for 10 s. 1.7. Rinse a glass micro-syringe with chloroform for 5 times. 1.8. Transfer 200 µL of the DPhPC chloroform solution into a pre-washed glass vial using the washed syringe. Evaporate the solvent in the vial with a gentle stream of nitrogen. 1.9. Rinse a glass micro-syringe 5 times with chloroform. 1.10. Add 202.6 µL of chloroform to the glass vial with DPhPC using the cleaned syringe. 1.11. Add 47.4 μL of filtered 10 mg/mL CuPc solution into the DPhPC solution. The molar ratio of DPhPC to CuPc should be 10:1. 1.12. Rinse the glass micro-syringe 5 times with chloroform. 1.13. Add 250 µL of hexane to the solution using the syringe. The final concentration of the solution should be 10 mg/mL. 1.14. Mix the prepared solution using a vortex mixer at 2300 rpm for 10 s. 1.15. Filter the CuPc solution through a 0.2 μm polytetrafluoroethylene (PTFE) membrane. 1.16. Seal the glass vial with parafilm. Put it in to a grip sealed bag filled with nitrogen and place the grip sealed bag in a freezer at -20 °C. NOTE: After step 1.13, the DPhPC and CuPc were dissolved in a mixed solvent, which was composed of chloroform and hexane (volume ratio of 1:1). In addition, the molar ratio of DPhPC to CuPc is not limited to 10:1. With a constant concentration of lipids (10 mg/mL), different molar ratios can be used. According to previous experimental results, a range from 10:1 to 3:1 is preferred for forming a high-quality hybrid lipid membrane. 2. Formation of a hybrid membrane at the water/air interface 2.1. Cut Si substrates (3 cm x 3 cm) from a Si wafer.

2.2. Clean the 3 cm x 3 cm Si substrates in an ultrasonic bath for 10 min in purified water, followed

by ethanol and then chloroform. Treat the Si substrate with an O₂ plasma for 5 min to remove adsorbed organic materials from the surface and to improve hydrophilicity.

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2.3. Wash a Teflon beaker with an inner diameter of 7.5 cm with flowing purified water for 3 min.

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2.4. Put the cleaned Si substrate in the washed Teflon beaker, which is tilted at an angle of 30° to the horizontal.

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2.5. Pour a sufficient amount of purified water into the Teflon beaker until the entire Si substrate
 is submerged.

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2.6. Take prepared hybrid solution out of freezer and allow it to warm to room temperature.

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2.7. Stir the hybrid solution using a vortex mixer at 2300 rpm for 15 s.

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2.8. Rinse a glass micro-syringe (50 μL) 5 times with chloroform.

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2.9. Drop 3-5 μL of the hybrid solution onto the water surface using the syringe to form a floating
 hybrid lipid membrane.

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NOTE: When the solution is dropped, it is important to hold the droplet close to (less than 1 cm) the water surface. It should also be noted that a single-layer hybrid membrane is not visible to the naked eye, but the multi-layer hybrid membrane appears as thin, blue-colored film. In order to transfer the multi-layer hybrid membrane to the Si substrate, it is important to drop the mixed solution as close as possible to the Si substrate.

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3. Transferring the membrane onto a Si substrate

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3.1. After the evaporation of organic solvent (it will take less than 2 seconds), lower the water level at a rate of 3 mm/min by pumping out the water through a rubber tube which is driven by a peristaltic pump, to transfer the floating hybrid membrane onto the Si substrate.

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3.2. After the transfer process is complete (about 5 min), place the Si substrate on a cleanroom wiper, and allow all of residual water to evaporate.

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NOTE: Lowering the water level at such a low rate serves to minimize the turbulence of water and to protect the membrane.

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REPRESENTATIVE RESULTS:

- 172 The as-formed membrane has a uniform light blue color due to the presence of CuPc molecules.
- 173 The area of the colored membrane is normally several square centimeters. In Figure 1A and
- 174 Figure 1B, we show a microscopic image and an atomic force microscope (AFM) image (including
- a height profile) of the hybrid lipid membrane on a Si substrate. In the AFM image, the membrane
- in the upper left is thick, with a thickness of 79.4 nm and that at the lower right is thin, with a

thickness of 4.9 nm. The thin membrane shows a surface roughness of Ra = 0.4 nm, which is close to that of the cleaned Si substrate. In addition, based on multiple measurements, membrane thicknesses show a discrete distribution with 5-nm increments¹⁸. Since the reported thickness of a DPhPC lipid bilayer membrane is about 4 nm¹⁹, it can be concluded that the 5 nm thin membrane is a CuPc doped lipid bilayer membrane and that the thick membranes are composed of stacks of doped lipid bilayers.

Energy dispersive X-ray (EDX) analysis was used to further investigate the composition of the hybrid membrane on the Si substrate. As calculated from the data shown in **Figure 2**, the atomic ratios of representative elements such as Cu, P, N and C, are 0.33%, 0.97%, 4.06% and 68.56%. Considering that a molar ratio of 3 to1 (DPhPC to CuPc) was used in preparing the hybrid membrane, the theoretical molar ratio of Cu:P:N:C should be 1:3:11:192, which is close to the measured element ratio in the hybrid membrane, indicating that the ratio between the lipids and the CuPc molecules is maintained after the film formation and transfer processes.

FIGURE AND TABLE LEGENDS:

Figure 1: As-formed hybrid membrane doped with CuPc. (A) A confocal microscopy image of the hybrid membrane. (B) AFM image of the hybrid membrane. The membrane shown in (B) includes a hybrid multilayer membrane with a thickness of 79.4 nm and a contiguous monolayer hybrid membrane with a thickness of 4.9 nm.

Figure 2: EDX pattern of the hybrid membrane on a Si substrate. Corresponding elements to the features are shown in the figure.

DISCUSSION:

In the precursor solution of the hybrid membrane, a mixed organic solvent (chloroform and hexane) rather than pure chloroform is used to dissolve lipids and CuPc. If pure chloroform is used, the density of the precursor solution would be higher than water. Therefore, it is highly likely that the solution would sink to the bottom of water rather than spread on water surface. Adding hexane, a low density solvent, to the precursor solution, ensures that the solution will float on the water surface and form a uniform hybrid membrane after the evaporation of the solvent. It should also be noted that immediately after the solution comes into contact with the water surface, a slight surface wave was always observed, possibly due to the spreading of the membrane and the organic solvent, which would alter the surface tension of water. Since the hybrid membrane is ultrathin and fragile, a small disturbance would damage the intactness of membrane with visible cracks being formed. Therefore, in order to prevent further damage, it is very important to minimize the airflow around the membrane and to avoid any vibrations of water after the formation of hybrid membranes. If possible, the Teflon beaker should be placed on a vibration isolation table.

The well-established Langmuir–Blodgett (LB) method is widely used to form lipid monolayers at the water/air interface where hydrophilic heads of the lipids are oriented toward the water while the hydrophobic tails are oriented toward the air. Different from the LB method, by doping with CuPc molecules, our method permitted hybrid membranes with a bilayer structure to be formed

at the water/air interface in one step. In the hybrid membrane, it is assumed that the hydrophobic CuPc molecules are located between the hydrophobic "tails" of lipid molecules, forming an intriguing lipid/CuPc/lipid sandwich structure. We confirmed this assumption by conducting fluorescence resonance energy transfer (FRET) measurements¹⁸.

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In addition, we repeated the film formation process using several different molar ratios of DPhPC to CuPc following the same protocol described above. Generally, a low CuPc ratio (e.g., molar ratio of 20:1) resulted in a hybrid membrane with a lighter color and a smaller area than one prepared using a high CuPc ratio (e.g. 3:1 and 10:1). It appears that the CuPc molecules assist in the bilayer formation and also act as an adhesive, resulting in the formation of membranes with large areas. In addition, in the absence of lipid molecules, CuPc molecules tend to aggregate on both the water surface²⁰, and on the solid substrate after the solvent evaporation²¹. However, in the case of a hybrid membrane, XRD results indicated that the CuPc molecules did not aggregate to form small crystals in the hybrid membrane¹⁸. This suggests that the aggregation of CuPc molecules was prevented by the interaction of hydrophobic "tails" of lipids with CuPc. However, when more CuPc molecules were used to prepare a lipid solution (such as a molar ratio of 1:1), the hybrid membranes not only show a darker blue color but CuPc molecules are also visibly aggregated in the membranes. Considering that the size of the CuPc molecule (1.7 nm) is slightly larger than the diameter of the head group of the lipid molecule (about 1 nm), a molar ratio higher than 3:1 tends to result in films that contain aggregated CuPc molecules. The molar ratio of 10:1 that was used in the representative experiments is a tradeoff between the membrane area and undesirable aggregation.

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Following the described protocol, hybrid lipid membranes with a sandwich structure were formed at the water/air interface. By doping with CuPc molecules, the hybrid lipid membrane would possess some of functionalities of semiconducting molecules, including optoelectronic and photocatalytic properties, which would significantly expand the applications of lipid bilayer structures. It should be also noted that the doping material is not limited to CuPc molecules. We have also formed similar hybrid membranes with lipid/Pc/lipid sandwich structures by using 2,9,16,23-tetra-tert-butyl-29H,31H-phthalocyanine (H₂PC) and zinc 2,9,16,23-tetra-tert-butyl-29H,31H-phthalocyanine (ZnPC) as the doping materials. Other groups have demonstrated that the surface-modified Au-nanoparticles, graphene nanosheets, and fullerenes could stabilize inside the lipid bilayer^{7,9,12}. Therefore, it is possible to dope a lipid bilayer with other hydrophobic molecules and nanomaterials, which would further expand the range of applications of hybrid lipid membranes.

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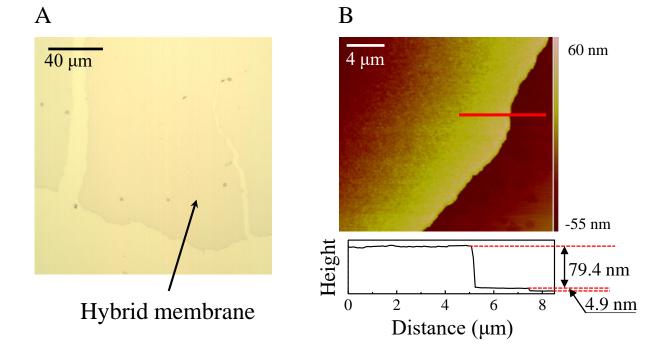
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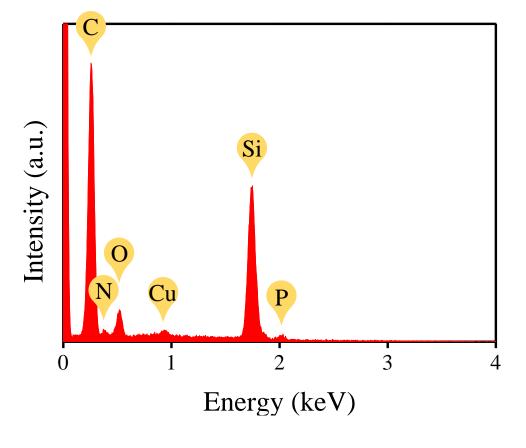
The authors have nothing to disclose.

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Name of Reagent/Material	Company	Catalog Number	Comments
Chloroform	Wako Chemicals	033-08631	
CuPc	Sigma-Aldrich	423165	
DPhPc	Avanti Polar Lipids	850356C	
Glass vials with screw cap	Nichiden-Rike Glass Co., Ltd	6-29801	
Hexane	Wako Chemicals	084-03421	
Membrane filters	Merck Millipore Ltd.	R8CA42836	
Micro-syringe	Hamilton	80530	
Peristaltic pump	Tokyo Rikakikai Co., Ltd.	11914199	
Vortex mixer	Scientific Industries, Inc.	SI-0286	

Dear Editor,

Thank you for your letter concerning our manuscript entitled "The self-assembly of hybrid lipid membranes doped with hydrophobic organic molecules at a water/air interface" (JoVE60957) which we submitted for publication in JoVE Video Journal.

We greatly appreciate the reviewers' insightful and suggestive comments. We have carefully revised our manuscript according to the reviewers' comments and issues pointed out by the editor. A list of supplies and equipment was also added. The response to the reviewer's comments is as follows:

(The corresponding corrections and revisions are marked with red color in the revised manuscript)

Reviewers' comments:

Reviewer #1:

Manuscript Summary:

This manuscript describes a method to prepare lipid bilayer-small molecule hybrids and their thin films in a relatively simple way, demonstrating a method recently published in JPhysChem.

Major Concerns:

There is an effort to suggest (Line 28, 39, 63) that the method is quite general for hydrophobic materials (in particular, extendable to nanoparticle hybrids) but generality of the method is not clear from the article, and would be fairly surprising. This may arise from calling the Cu-phtalocyanine complex as a nano-material (debatable, but acceptable in a broad sense), but for clarity, it shouldn't be referred to as a nanoparticle.

We are sincerely sorry for the improper definition of doping materials. All "nanoparticle" was replaced by "nanomaterial" in the revised manuscript. In the Discussion part of the revised manuscript, we have deleted the statement regarding the generality of the method. Examples showing that other nanomaterials could stabilize in the lipid bilayers are now added in the revised manuscript.

Minor Concerns:

Step 2.6. clarify if the hybrid solution is used directly from freezer or if returned to RT. Step 3.1 please provide more detail (in video or in manuscript) as to from where the water is removed.

As suggested by the reviewer, experimental details were added to Step 2.6 and 3.1.

Reviewer #2:

Manuscript Summary:

The authors describe a simple protocol for forming air-stable hybrid lipid bilayers with organic copper-phthalocyanine molecules on silicon substrates.

Minor Concerns:

The authors make the point that their method for forming the hybrid bilayers on silicon substrates is a one-step procedure which doesn't require expensive equipment or complicated processes, which apparently includes a langmuir trough. Using a langmuir trough to form conventional lipid bilayer films is indeed a difficult task, since they are not air stable, requiring the film to be submerged in aqueous solution at all times. However, the use of a langmuir trough to form these hybrid bilayers at the water-air interface would be no more difficult than for conventional langmuir monolayers, precisely because they are air-stable. The use of a langmuir trough would allow the user to monitor the interfacial tension and control the surface pressure of the film on the fly, and to monitor and optimize the transfer to the substrate by recording the transfer ratio in real time. I'm not suggesting that the authors must use a langmuir trough for this manuscript to be published, but they are missing an opportunity here by discounting out of hand the use of a langmuir trough.

We greatly appreciated the suggestion from the reviewer. As pointed out by the reviewer, using a langmuir trough allow us to monitor and optimize the transfer process, and would possibly improve the quality of transferred membrane. We are now planning to use langmuir trough to form the hybrid membrane and the results will be reported elsewhere.



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