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TITLE:

A Blood-Free Diet to Rear Anopheline Mosquitoes

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KEYWORDS:

artificial meal, mosquito, *Anopheles* diet, rearing, fresh blood substitutes, oogenesis

SUMMARY:

A protocol is presented for formulation of a blood-free artificial diet to feed *Anopheles* mosquitoes in captivity. This diet has a similar performance to vertebrate blood and triggers oogenesis and egg maturation and produces viable adult progeny.

ABSTRACT:

Malaria research requires large-scale breeding and production conditions for mosquitoes (*Anopheles* spp.) in captivity. The sustainable and reliable production of mosquitoes is currently inhibited by the supply of fresh vertebrate blood. Alternatives to blood are required to promote efficient control strategies for malaria and other vector borne diseases that are transmitted by blood feeding insects. With this in mind, artificial liquid diets were formulated as substitutes for fresh vertebrate blood. Herein we report a blood-free artificial liquid diet that delivers feeding rates similar to blood and mimics the physiological effects of a fresh vertebrate blood meal. The diet induces ovarian and egg maturation of *Anopheles* mosquitoes and also produces good larval survival and development of functional adults. The formulated blood-free liquid diet is an important advance towards sustainable mosquito breeding in captivity and will reduce the maintenance costs of mosquito colonies and eliminate the need for fresh vertebrate blood.

INTRODUCTION:

Vector-borne diseases affect several million humans worldwide and cause millions of deaths each year. They are transmitted by insects infected with disease-producing microorganisms

(protozoan, viruses) acquired when they feed on blood from an infected host. Subsequently, the infected vector will transmit the pathogen to a new host during the next blood meal. Malaria is the deadliest vector-borne disease that is transmitted by several different species of *Anopheles* mosquito and affects 40% of the world's population¹. The malaria protist parasite is responsible for more than 400,000 deaths every year, of which most are children under 5 years of age (World Health Organization). The female *Anopheles* mosquito transmits the malaria parasite of the *Plasmodium* genus between humans and other animals when it feeds on vertebrate blood, a necessary step for egg production and development².

Current strategies for eradication of malaria and other emerging deadly mosquito vector-borne diseases rely on the development of innovative mosquito control strategies³⁻⁵, which include the release into the wild of large numbers of mosquitoes bred in insectaries. However, a crucial limiting factor is the dependency on a supply of fresh blood for effective mosquito rearing and breeding. The variable composition of vertebrate blood can negatively impact mosquito fertility and progeny fitness and can limit the reliability and sustainability of captive breeding colonies. Mosquito release and control programs require large-scale mosquito production systems and a regular supply of large quantities of vertebrate blood. This is a major obstacle for mosquito production and raises a series of ethical issues associated with the use of live animals and logistical limitations caused by the associated demanding safety regulations. This makes the costs of maintenance and security of mosquito colonies high and challenges the sustainability of current mosquito rearing practices particularly in low income countries where the threat of malaria is far greater.

Recently research has been focused on the development of blood substitutes that mimic a vertebrate blood meal but so far, only limited success has been achieved⁶⁻⁹. A successful artificial diet needs to (1) provoke full female mosquito engorgement, (2) trigger vitellogenin production, (3) produce large batches of viable eggs, and (4) generate healthy progeny¹⁰. In addition, artificial diets have a standard composition and thus are more reliable for production of mosquitoes for research and control purposes. Successful blood-free diets have been developed for *Aedes* mosquitoes (reviewed by Gonzales and Hansen¹¹) but not for *Anopheles* spp. Existing artificial diets contain a phagostimulant (e.g., ATP¹⁰), a protein source for egg maturation^{6,12}, carbohydrates as a source of energy, and amino acids (aa)¹³ that are fundamental for egg production and are a major limiting factor for mosquito fertility¹⁴. An artificial blood free diet also needs to provide cholesterol¹⁵, which improves egg production. Here we describe an artificial blood-free diet for female *Anopheles* mosquitoes and demonstrate that it has a consistent and equivalent performance to a high-quality vertebrate blood meal.

PROTOCOL:

Mice were obtained from the IHMT animal house. Animal experiments were conducted in strict accordance with the Portuguese law and guidelines for the use of laboratory animals. The Direção-Geral de Veterinária, Ministério da Agricultura do Desenvolvimento Rural e das Pescas, Portugal approved all the study protocols (id approvals: 023351 and 023355).

NOTE: Perform all feeding assays at room temperature (~21 °C).

1. Mosquitoes

1.1. Maintain *Anopheles coluzzii* (former *Anopheles gambiae* M form) Yaoundé strain mosquitoes at 26 ± 1 °C, 75% humidity under a 12 h:12 h light:dark cycle. House mosquitoes using standard insectary conditions to guarantee mating.

1.2. Collect mosquito pupae into a small water container. Place the container inside a mosquito cage. Let adult mosquitoes emerge and mate. Provide 10% glucose feeding solution. Three days after emergence collect the necessary number of mosquitoes from the stock cage using an aspirator.

1.3. One day before the feeding trials, remove the 10% glucose feeding solution.

NOTE: Three-day-old mosquitoes were used throughout the experiments.

2. Mosquito feeding

2.1. Preparation of artificial liquid diets

2.1.1. Prepare the artificial liquid diets under sterile conditions in a laminar flow cabinet. Prepare the rich liquid diet (r-liq_diet) by adding the following to the initial liquid diet (i-liq_diet; Dulbecco's modified Eagle's medium [high glucose with L-glutamine], see **Table 1**): 0.55 g/L ATP, 1 g/L cholesterol, and 200 g/L bovine serum albumin (BSA). Mix all ingredients thoroughly and filter using a 0.45 µm microfilter.

NOTE: Do not store the diets; prepare diets freshly from the stock solutions for each experiment as they lose quality when stored. Components of the diets are described in **Table 1**.

2.2. Mouse blood collection

2.2.1. Anesthetize 6–8-week-old CD1 female mice (*Mus musculus*) with ketamine (120 mg/kg) and xylazine (16 mg/kg) using the intraperitoneal route.

2.2.2. Perform cardiac puncture (**Figure 1**) when the mouse displays no muscle reaction in response to different physical stimuli (e.g., toe and tail pinches).

2.2.3. Collect blood using a sterile 1 mL syringe with a 27 G x ½" (0.4 x 12 mm) needle that contains 100 µL of 1 mg/mL heparin (sodium salt) to prevent formation of blood clots. Maintain blood at 37 °C using a water bath. Euthanize mice by inhalation of 100% CO₂.

2.3. Artificial feeding

2.3.1. Collect approximately 30 female mosquitoes from the stock cage using an aspirator.

2.3.2. Anesthetise the female mosquitoes within the aspirator chamber by placing them at -20 °C for 30 s.

2.3.3. Transfer the female mosquitoes to 500 mL paper cups and cover with a fine mosquito net mesh so they cannot escape. Apply a glass feeder connected to plastic tubes to maintain a constant water flow to the top of each cup (**Figure 2**). Provide a constant water flow to the cylindrical tubing and feeder so the temperature within is kept at 37.5 °C.

NOTE: A standard glass bell artificial feeding apparatus¹⁶ was used to supply the formulated diets to female mosquitoes.

2.3.4. Stretch paraffin film membrane across the mouth of the glass feeder to contain the meal.

2.3.5. Pre-warm the i-liq_diet and r-liq_diet at 37 °C using a water bath. Apply 1 mL into the glass feeder. Feed the mosquitoes for 60 min in the dark with either i-liq_diet, r-liq_diet or fresh mouse blood. Perform assays at room temperature (~21 °C).

2.4. Evaluation of the feeding rate

2.4.1. After artificial feeding, cold-anesthetize the mosquitoes at -20 °C for 30 s. Place the mosquitoes in a refrigerated Petri dish.

2.4.2. Record the number of fully engorged (**Figure 3**) female mosquitoes.

NOTE: The percentage of fed mosquitoes is used as a proxy for feeding success.

3. Life history traits

3.1. Egg production and fertility

3.1.1. Transfer the fully engorged females using a brush to individual cages (20 cm x 20 cm x 20 cm) with a humidified filter paper at the bottom for egg laying (**Figure 4**).

3.1.2. Keep the mosquitoes at 26 ± 1 °C, 75% humidity and a 12 h:12 h light:dark cycle with 10% glucose *ad libitum*.

3.1.3. Count the eggs at 48 h and 72 h post feeding using a handheld magnifying glass. Flood the filter paper with distilled water to collect the eggs.

3.2. Larvae mortality

3.2.1. Collect the eggs into trays (23 cm x 15 cm x 6 cm) filled with distilled water (**Figure 5**). Maintain the water level in the trays constant during the experiments.

3.2.2. Feed the larvae daily with approximately 13 mg of ground fish food per tray. Apply a similar feeding regime to all the replicate trays.

3.2.3. Remove dead pupae and larvae daily. Finalize the experiments when all pupae have developed into adults and count the number of adult males and females.

3.2.4. Register the dates of hatching and death and calculate the mortality rates.

3.3. Longevity

3.3.1. Collect 15 adult males and 15 adult females from the F1 generation of each diet group. Keep males and females in the same cage.

3.3.2. Feed adults with a 10% glucose solution *ad libitum*. Remove the dead adults daily.

3.3.3. Maintain the mosquitoes at the same temperature, humidity, light cycle conditions and sugar feeding regime as described above.

3.3.4. Register the death dates and calculate the longevity.

3.4. Wing length measurement

3.4.1. Cold-anesthetize five-day-old F1 adult mosquitoes (male and female) from each diet group at -20 °C for 90 s.

3.4.2. Under a stereoscope, gently grasp the thorax of each mosquito with forceps and place them ventral side up.

3.4.3. Collect both wings using a scalpel and place them on a clean microscope slide containing a dried drop of mounting medium. With the aid of a 20 G needle add extra mounting media to the borders of the slide and slowly lower the coverslip onto the wings.

3.4.4. Measure the wing length (**Figure 6**) with a stereoscope using a micrometer.

REPRESENTATIVE RESULTS:

The results described below compare the performance of female *Anopheles* mosquitoes fed with the formulated rich artificial meal (r-liq_diet) and mosquitoes fed on the initial liquid diet (i-liq_diet) or a fresh blood meal. The diet was tested following the schematic protocol depicted in **Figure 7**. The r-liq_diet described herein is part of a patent (PCT/IB2019/052967).

Percentage of fully engorged females

The number of engorged female mosquitoes fed with the r-liq_diet (60%) was significantly higher than the number of engorged females fed on blood (44%) (**Figure 8**).

Fecundity and fertility

Female fecundity and fertility for the first gonotrophic cycle was used to evaluate the nutritional quality of the i-liq_diet and r-liq_diet. An average of 24 ± 11 eggs was laid by females fed on fresh vertebrate blood, whereas females fed on r-liq_diet laid an average of 25 ± 5 eggs (**Table 2**). No egg laying was observed by females fed on the i-liq_diet.

F1 mortality

The fitness of F1 mosquitoes was evaluated between colonies fed on vertebrate blood or the r-liq_diet. Larvae, pupae and adult mortality was recorded. Variability (standard error of the mean [SEM]) was higher in the blood-fed mosquitoes (**Figure 9** and **Table 2**) relative to mosquitoes fed the r-liq_diet. The F1 generation of mosquitoes fed on either the blood or the r-liq_diet had comparable mortality and survival rates.

F1 life expectancy

It is estimated by the Center for Disease Control and Prevention that wild adult female mosquitoes live for up to a month but probably do not survive for more than 1–2 weeks and that males live for about a week and feed exclusively on nectar and other sugar sources. It is noted that differences in parental food intake may affect the survival of the mosquito progeny¹⁷. In our experiment, adult females and males in the blood (female 24.5 ± 6.8 ; male 18.5 ± 6.9) and r-liq_diet (female: 22.5 ± 8.1 ; male: 11.9 ± 6.9) groups had similar average life expectancies average (**Table 3**) and females showed an increased life span relative to males.

F1 body size

Wing length was used as an indicator of adult body size. When compared to other species, *Anopheles* adults are small- to medium-sized mosquitoes with a wing length of between 2.8 to 4.4 mm wing length¹⁸. Adult body size of F1 *Anopheles* mosquitoes fed the r-liq_diet was within the expected range and was similar to blood-fed insectary mosquitoes (**Figure 10**).

Statistical analysis

The data presented represents the mean of at least three independent experiments (unless otherwise stated). Error bars represent the SEM. When data followed a Gaussian distribution, independent groups were compared using the Student's t test, otherwise the Mann-Whitney test was applied. Differences between the artificial diet-fed groups were analyzed using the Fisher's exact test and were considered to be significant at $P \leq 0.05$.

FIGURE AND TABLE LEGENDS:

Figure 1: CD1 mouse blood collection by intracardiac puncture.

Figure 2: Standard artificial feeding apparatus. The glass feeder contains r-liq_diet that is being fed to female *Anopheles* mosquitoes.

Figure 3: *Anopheles* mosquitoes after artificial feeding. From left to right: a fully engorged female that was offered r-liq_diet, non-engorged female that was offered r-liq_diet, male, and fully engorged female that was offered mouse blood.

Figure 4: *Anopheles* eggs laid 48 h post-feeding of females with the r-liq_diet.

Figure 5: L2 larvae stages that developed from the eggs and were collected on filter paper and placed in trays containing distilled water.

Figure 6: Right wing from an F1 generation of an *Anopheles coluzzii* female mosquito.

Figure 7: Schematic protocol of artificial diet testing.

Figure 8: Feeding rates of the female mosquitoes fed either the artificial diets or blood. Asterisks indicate significant differences between the mosquitoes fed the r-liquid and i-liquid diets and the blood-fed control group. Two-sided Fisher's exact test: **** $P \leq 0.0001$ (relative risk: 0.4828, 95% confident Level [CL]: 0.3776 to 0.6194) for r-liquid diet versus blood, * $P = 0.0335$ (relative risk: 1.379, 95% CL: 1.044 to 1.836) for blood versus i-liquid diet. Blue: unfed; red: fed.

Figure 9: Effect of the formulated blood-free meal on the mortality and male/female ratio of F1 *Anopheles coluzzii* mosquitoes. Three independent experiments were performed, each using 30 mosquitoes per diet. An unpaired t-test showed no significant differences between the blood-fed group and the r-liq_diet fed group (P values varied from 0.5047 to 0.8491).

Figure 10: Wing length. The distance from the axial incision to the R4+5 vein excluding the fringe seta was used to determine the wing length. Size was evaluated for 5 females and 5 males from each dietary group (mean \pm SEM). Values are represented as the mean \pm SEM. Salmon: r-liq_diet; red: vertebrate blood. Unpaired t-test; female left wing: $t = 1.300$, $df = 8$, $P = 0.2298$; male left wing: $t = 2.400$, $df = 8$, $P = 0.0432$; female right wing: $t = 1.300$, $df = 8$, $P = 0.2298$; male right wing: $t = 2.277$, $df = 7$, $P = 0.0569$.

Table 1: The composition of the i-liquid diet and r-liquid diet.

Table 2: Egg batches produced by *Anopheles coluzzii* females. Three independent experiments were performed for each experimental diet using 30 female mosquitoes in each.

Table 3: Life expectancy of F1 *Anopheles* mosquitoes. Longevity of F1 mosquitoes from artificially fed F0 was assessed by recording the dates of birth and death of each mosquito coming from the same dietary group (15 females and 15 males were followed). The results are represented as the mean mosquito life span per diet group.

DISCUSSION:

The success of our formulated blood-free diet is likely the result of the synergistic physiological effect of all the components added to the i-liq_diet (rich in sugar, amino acids, vitamins and microelements): BSA (protein source), ATP (phagostimulant) and cholesterol (lipid source). Supplementation of the r-liq_diet with the individual components alone was not effective in stimulating egg production (data not shown). One drawback of the protocol might be the cost of some of the components, such as cholesterol. Even so, its presence is fundamental, as insects are unable to synthesize it¹⁹ and this molecule is the precursor of the ecdysteroid hormones that regulate yolk synthesis and egg maturation in arthropods²⁰. Lower amounts of cholesterol should be tested in order to optimize the quantity needed with the aim of reducing costs and increasing the benefits of the artificial diet.

Another limitation of the method is that the artificial diet has to be freshly prepared from stock solutions, as once prepared in its final liquid form it loses quality after storage. In the future our formulated diet could be prepared as a dried power, similar to SkitoSnack^t, an artificial blood meal replacement for *Aedes aegypti* mosquitoes²¹.

Beside supplying the necessary nutrients, an artificial meal needs to attract and stimulate female mosquitoes to feed in the same way as when they feed on vertebrate fresh blood. The artificial blood-free diet herein described resulted in a 20% increase in fully engorged female mosquitoes when compared to the vertebrate blood fed group. This indirect measure of attraction could be further clarified by using olfactometers to confirm that the artificial diet is more attractive and more appealing to mosquitoes than fresh blood.

The highest impact of the diet on larval mortality was observed for larvae derived from mosquitoes fed on blood, suggesting that an artificial diet of stable composition can contribute to reduce mortality and improve mosquito breeding success when compared to fresh blood. The less predictable outcome of a blood meal may arise from host variations in composition¹⁷ and the presence in the blood of molecules that may interfere with mosquito physiology²². The preceding facts emphasize the advantages for high-quality mosquitoes rearing of fresh-blood-free diets.

Overall the average number of eggs laid in our study was low relative to those reported in some insectaries, but the mean number of oviposited eggs was comparable to the *A. gambiae* laboratory-reared strain fed on human blood (22.6 ± 5.5 eggs/female)²³. No significant statistical differences were observed between our experimental groups fed on either fresh blood or on the artificial meals (**Table 2**), suggesting that the implementation of an artificial membrane feeding systems with our formulated diet is sufficient to maintain and propagate *Anopheles* mosquito colonies in captivity.

Artificial blood-free meals can maintain *Aedes* colonies²², but when applied to *Anopheles* mosquitoes they are of limited or no success¹¹. Recently, a plasma-based artificial meal for *Anopheles* mosquitoes was described²⁴ but feeding rates and reproductive potential was low. Our results represent a substantial advance in the state-of-art (reviewed by Gonzales and Hansen¹¹) as our formulated r-liq_diet had a similar or better performance than the standard vertebrate blood meal. Further improvements on storage stability and cost should widen the scope of its application.

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DISCLOSURES:

The authors have nothing to disclose.

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Figure 1

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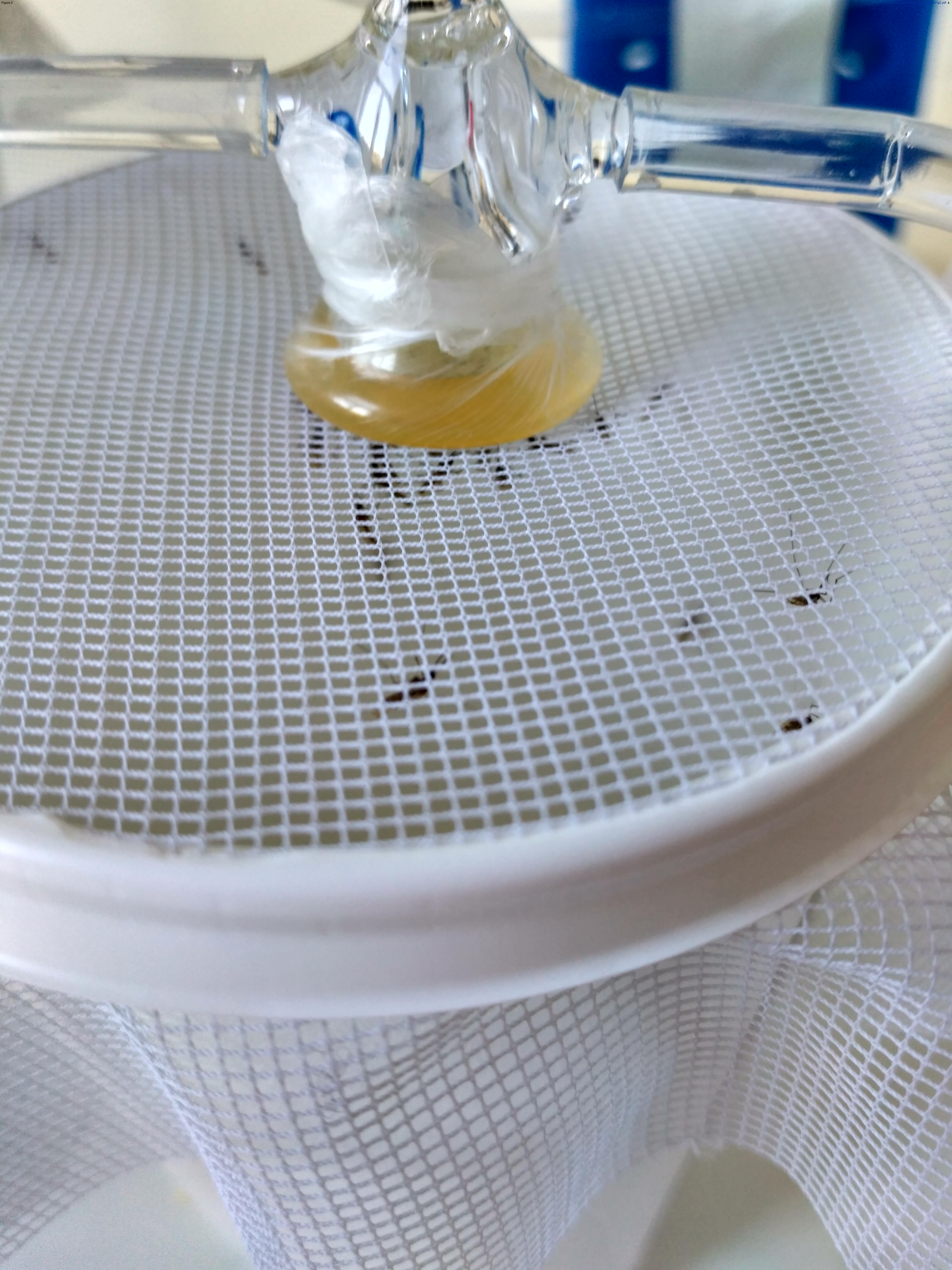
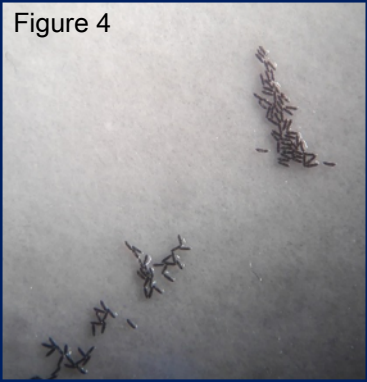


Figure 3

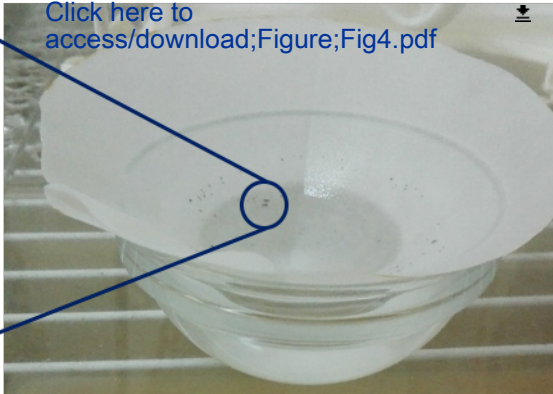
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Figure 4



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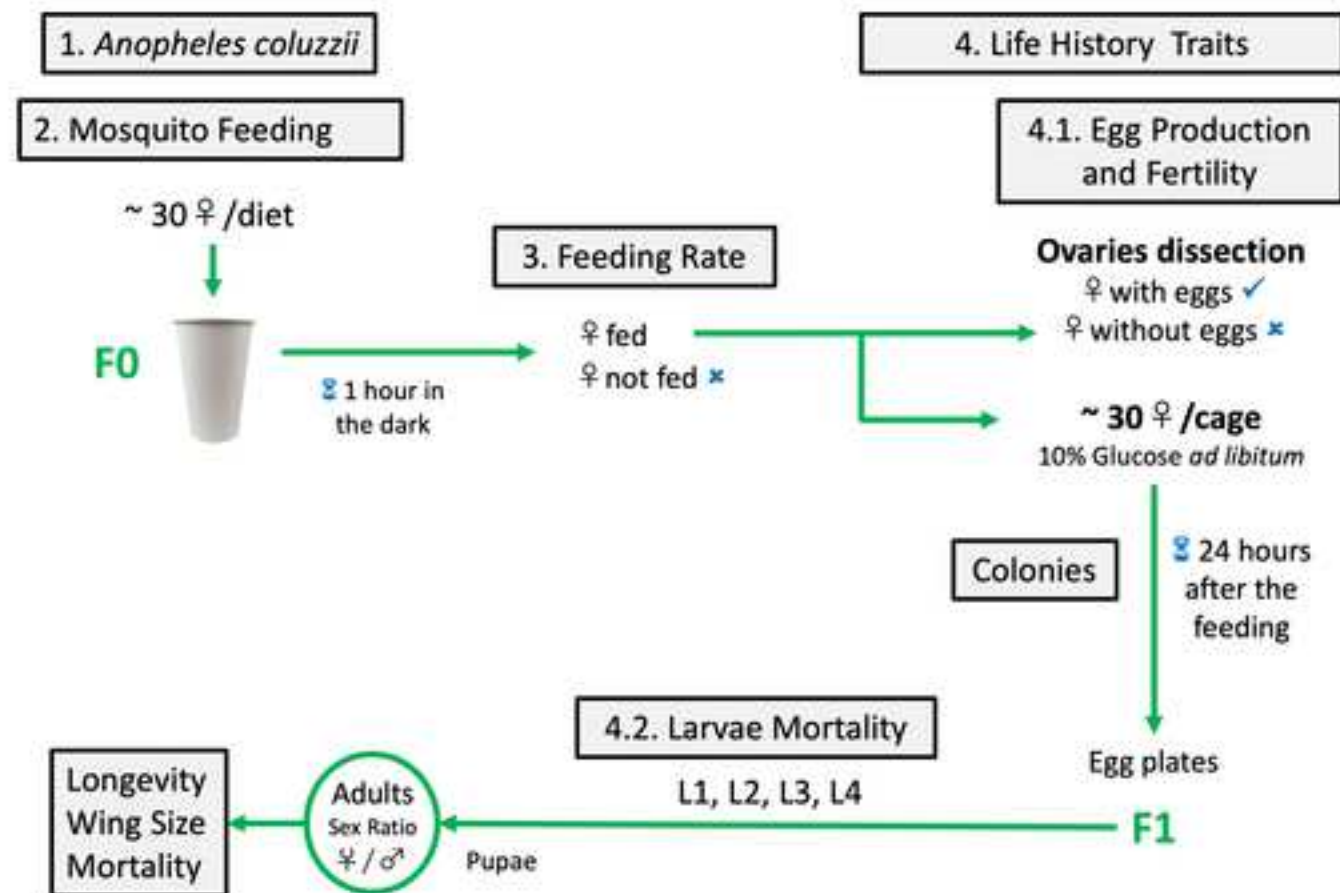
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DEL 144
EVALUATION

Figure 6

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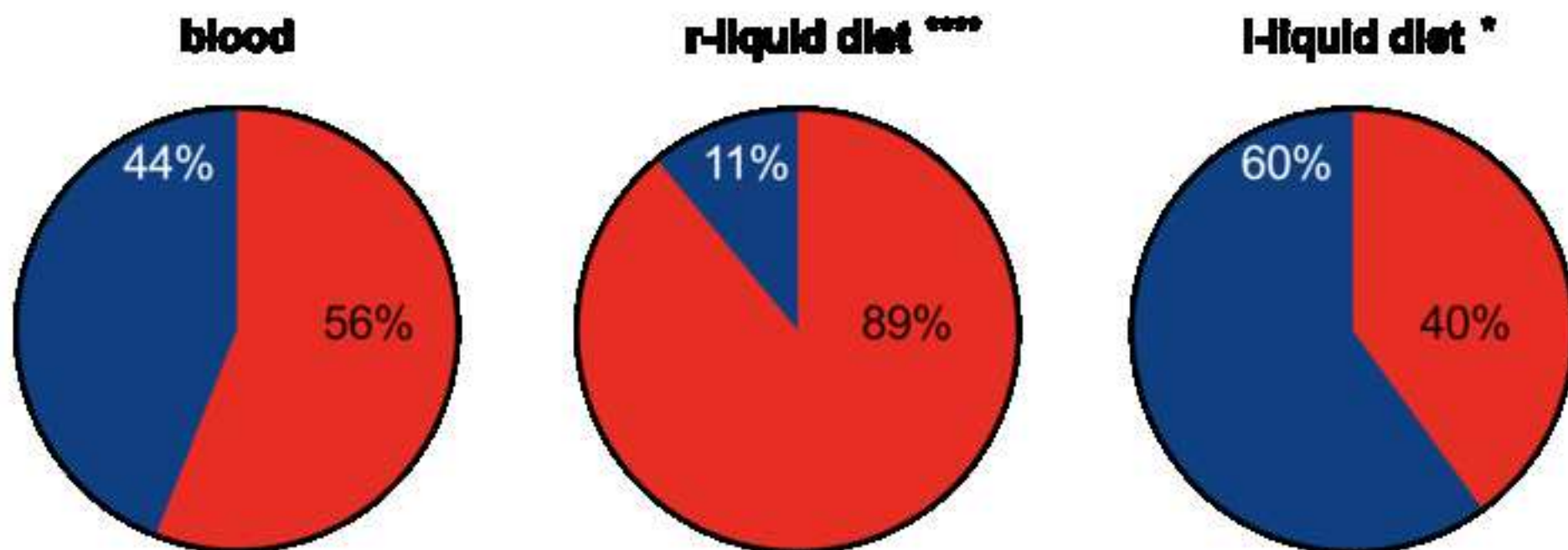
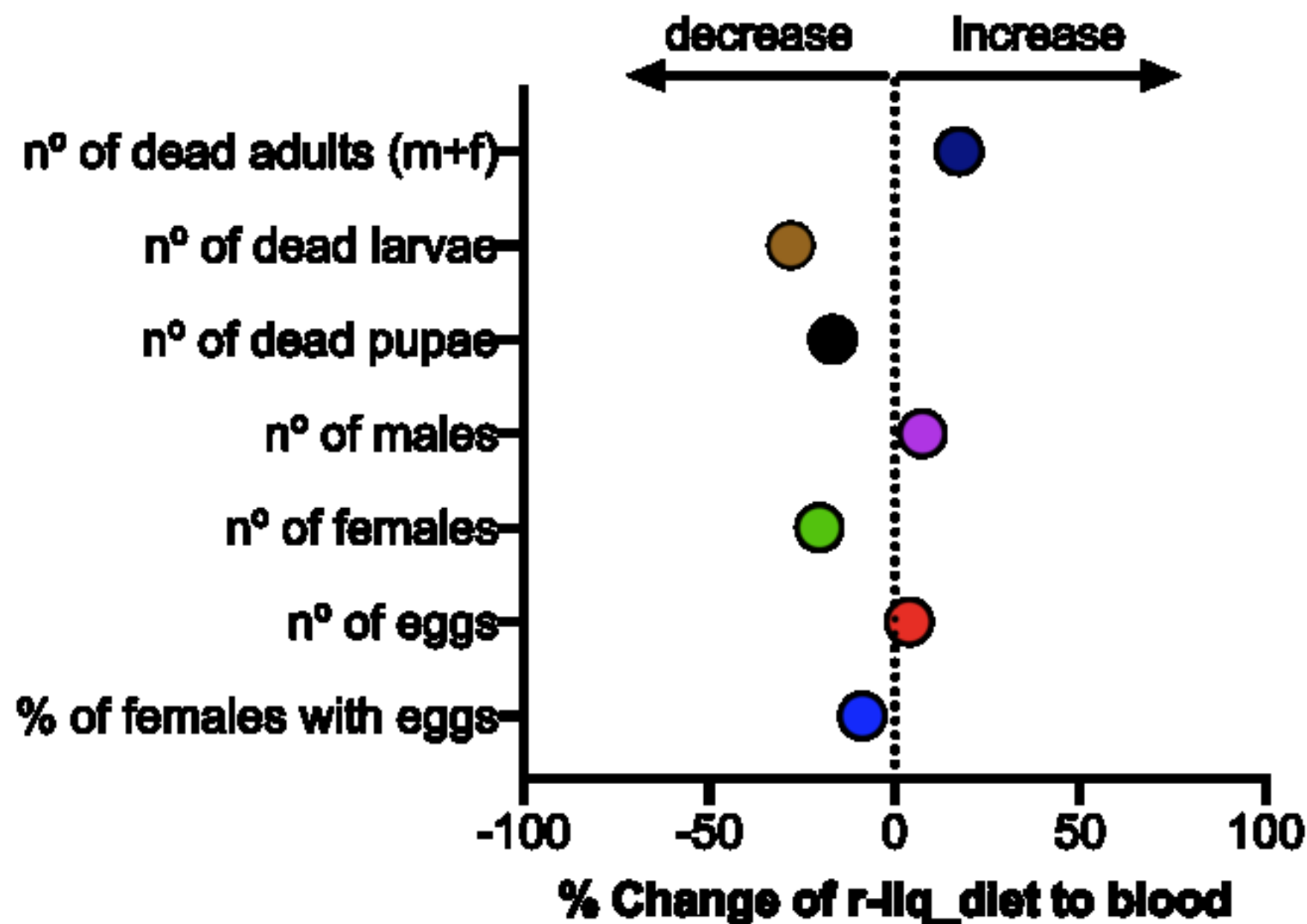
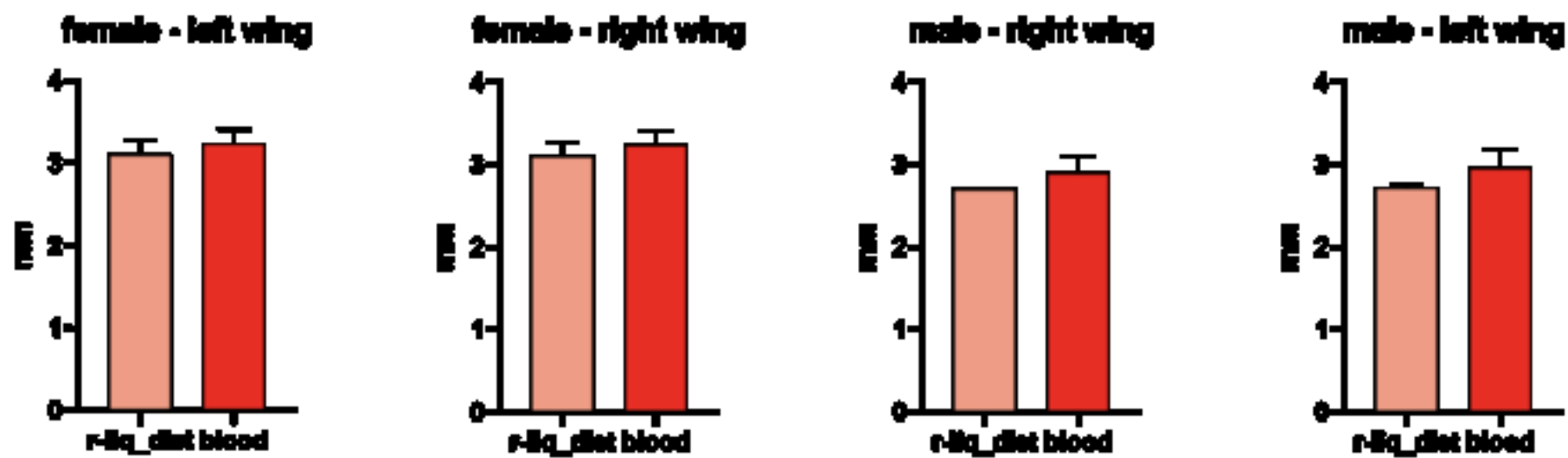


Figure 9





Components	g/L
*Adenosine Triphosphate	0.55
*Bovine Serum Albumin	200
*Cholesterol	1
Calcium chloride anhydrous	0.2
Choline chloride	0.004
D-calcium pantothenate (vitamin B5)	0.004
D-glucose anhydrous	4.5
Ferric nitrate nonahydrate	0.0001
Folic acid	0.004
Glycine	0.03
I-inositol	0.007
L-arginine monohydrochloride	0.084
L-cystine dihydrochloride	0.063
L-glutamine	0.584
L-histidine monohydrochloride monohydrate	0.042
L-isoleucine	0.105
L-leucine	0.105
L-lysine monohydrochloride	0.146
L-methionine	0.03
L-phenylalanine	0.066
L-serine	0.042
L-threonine	0.095
L-tryptophan	0.016
L-tyrosine disodium salt dihydrate	0.104
L-valine	0.094
Magnesium sulfate anhydrous	0.098
Niacinamide (nicotinamide)	0.004
Phenol red	0.015
Potassium chloride	0.4
Pyridoxine Monohydrochloride	0.004
Pyruvic acid sodium salt	0.011
Riboflavin (vitamin B2)	0.0004
Sodium bicarbonate	3.7
Sodium chloride	6.4
Sodium phosphate monobasic anhydrous	0.109
Thiamine monohydrochloride (vitamin B1)	0.004

*Only in r-liq_diet

	Total egg number (± SEM)	Eggs/female (± SEM)
Blood	733 ± 330	24 ± 11
r-liq_diet	763 ± 164	25 ± 5
i-liq_diet	0	0

	Females (days ± SEM)	Males (days ± SEM)
Blood	24.5 ± 6.8	18.5 ± 6.9
r-liq_diet	22.5 ± 8.1	11.9 ± 6.9

Name of Material/Equipment	Company	Catalog Number	Comments/Description
Adenosine 5'-triphosphate (ATP) disodium salt hydrate	Sigma Aldrich	A2383	
BSA-Bovine Serum Albumin	Sigma Aldrich	A790G	
Cholesterol	MP Biomedicals	199342	
glutamine)	Lonza Bioscience	BE12-604F	
Entellan mounting medium	Merck	1079610100	
Glassfeeder	Local glazier	by design	
	Pan Reac		
Heparin Sodium Salt	AppliChem	A3004,0001	
Imalgène 1000	Merial, Portugal	01MER122	
Needle 20 G x 1" 0.9 x 25 mm needle	Terumo Europe	NN-2025R	
Parafilm	Sigma Aldrich	P6543-1EA	
Rompun	Bayer, Portugal	7427831	
Sterilization Millex-HV 0,45	Millipore	SLHVR25KS	
Syringe, 1ml, 27 G x ½" 0.4 x 12 mm needle	Terumo Europe	BS-NIN2713	
		403073310095	
Teich Mix Astra Pond	Astra	7	
		400421874264	
Tetra Goldfish Flakes	Tetra	2	



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Blood-free diet to rear Anopheline mosquitoes

Author(s):

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Dear Editor,

Editorial comments were addressed as follows. All changes to the manuscript were made using track changes.

Editorial comments:

1. Please take this opportunity to thoroughly proofread the manuscript to ensure that there are no spelling or grammar issues. The JoVE editor will not copy-edit your manuscript and any errors in the submitted revision may be present in the published version.

[The MS has been revised. All alterations were made using track changes](#)

2. Please note that the editor has formatted the manuscript to match the journal's style. Please retain the same. The updated manuscript is attached and please use this version to incorporate the changes that are requested.

[Format was maintained. Automatic numbering might not be corrected. Please confirm. My word version does not always changes the document sub-heading numbering.](#)

3. Please note that the numbering of protocol steps has been updated according to JoVE guidelines. Please review for accuracy.

[Some changes were made please see comment above.](#)

4. Figure 3 is missing. Please upload it to your Editorial Manager account.

[Figure 3 was uploaded.](#)

5. Figure 6: Please include a space between the number and its unit (i.e., 1 mm).

[Figure 6 was altered](#)

6. Figure 7: Please update it to match the numbering of the protocol in the manuscript text.

[Figure 7 was altered](#)

7. Table 1: Please use exponential notation (instead of E notation) for numbers; i.e., change 4.00E-03 to 4×10^{-3} , etc.

[Number format was changed in Table 1](#)

8. Please address specific comments marked in the attached manuscript.

[All comments have been addressed, see below and MS for details.](#)

1. This is contradictory to line 139 (Perform assays at 27 °C). Please revise to be consistent.

[The temperature was changed to match, that in line 148](#)

2. How to obtain three-day-old mosquitoes? Please describe here in the imperative tense.

[The explanation was added. The points 1.1., 1.2. and 1.3. were altered for clarification \(line 93-103\)](#)

3. The editor rephrased this step. Please review for accuracy and make changes if necessary.

OK

4. Please spell it out.

Done (line 112)

5. Please specify age and gender.

Done (line 120)

6. Please describe how. Specify all surgical tools used.

The only tools used are a needle and a syringe. This procedure is specified on 2.2.3. (line 126)

7. Are they in a container? Please specify.

More information was added to line 136

8. With what, a pair of forceps?

No. Directedly one container dumped into the other

9. How large are the paper cups? 30 mosquitoes in one paper cup?

More information was added to line 136

10. What kind of net?

More information was added to line 136

11. Please describe how.

More information was added to line 146

12. This is contradictory to the previous statement.

The temperature was changed to match that on line 89

13. With what? Forceps?

More information was added to line 163

14. Can this be moved to step 3.1.1?

The sentence was removed out as it was a repetition of 2.4.2. and 3.1.1.

3.1.1. was also modified for clarity

15. Please specify the type of cage.

Information added to 3.1.1.

16. For how long is this done?

Experimental mosquitoes were kept till the last mosquito died

17. Are these males and females housed together or separately in different cages?

More information was added to line 188

18. Figure 3 is missing.

The figure 3 was uploaded

19. Please spell it out. Confidence level?

Done

20. Do you mean reference 11?

Yes. Changed accordingly

Other Changes:

1. Sentences were moved within the protocol section for better flow of the protocol.
2. Changes on the subtitles were made for clarify of the protocol and to increase consistency with the section "representative results". The following subtitles were renamed:

Line 191: "3. Egg production", was changed to "3. Life history traits"

Line 193: "3.1. Egg production", was changed to "3.1. Egg production and fertility".

Line 204: "3.2. Fertility evaluation" to "3.2. Larvae mortality"