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## Methods for short session high intensity interval training and treadmill assessment in aged mice --Manuscript Draft--

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**TITLE:**

Short Session High Intensity Interval Training and Treadmill Assessment in Aged Mice

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exercise, HIIT, aging, frailty, sarcopenia, gender, aerobic, anaerobic, endurance

**SUMMARY:**

Short session ( $\leq 10$  min) high intensity interval training (HIIT) is emerging as an alternative to longer exercise modalities, yet the shorter variants are rarely modeled in animal studies. Here, we describe a 10 min, 3 day a week, uphill treadmill HIIT protocol that enhances physical performance in male and female aged mice.

**ABSTRACT:**

High intensity interval training (HIIT) is emerging as a therapeutic approach to prevent, delay, or ameliorate frailty. In particular short session HIIT, with regimens less than or equal to 10 min is of particular interest as several human studies feature routines as short as a few minutes a couple times a week. However, there is a paucity of animal studies that model the impacts of short session HIIT. Here, we describe a methodology for an individually tailored and progressive short session HIIT regimen of 10 min given 3 days a week for aged mice using an inclined treadmill. Our methodology also includes protocols for treadmill assessment. Mice are initially acclimatized to the treadmill and then given baseline flat and uphill treadmill assessments. Exercise sessions begin with a 3 min warm-up, then three intervals of 1 min at a fast pace, followed by 1 min at an active recovery pace. Following these intervals, the mice are given a final segment that starts at the fast pace and accelerates for 1 min. The HIIT protocol is individually tailored as the speed and intensity for each mouse are determined based upon

initial anaerobic assessment scores. Additionally, we detail the conditions for increasing or decreasing the intensity for individual mice depending on performance. Finally, intensity is increased for all mice every two weeks. We previously reported in this protocol enhanced physical performance in aged male mice and here show it also increases treadmill performance in aged female mice. Advantages of our protocol include low administration time (about 15 min per 6 mice, 3 days a week), strategy for individualizing for mice to better model prescribed exercise, and a modular design that allows for the addition or removal of the number and length of intervals to titrate exercise benefits.

## INTRODUCTION:

Regular exercise is effective at preventing or delaying many age-associated diseases such as sarcopenia and frailty<sup>1-4</sup>. However, less than 15% of those 65 and older meet recommendations of 150 min a week of moderate intensity exercise plus strength training<sup>5,6</sup>. As the lack of time and lengthy sessions are common barriers to exercise, high intensity interval training (HIIT) is emerging as an alternative to traditional regimens. HIIT features multiple short bursts of intense activity that are interspersed with brief periods of active recovery, and there has been recent interest in identifying the shortest regimens that still yield beneficial outcomes. Such studies include 3 day a week regimens featuring total session times of 4 min<sup>7</sup>, 2-3 min<sup>8</sup>, 1.5 min<sup>9</sup>, a single min<sup>10</sup>, and even 40 s<sup>11</sup>.

Likewise, there has been substantial interest in HIIT animal models. A majority of studies used mice<sup>12-21</sup> or rats<sup>22-25</sup> and were performed using a treadmill, although a few others used swimming protocols<sup>26-28</sup>. A majority of these studies use VO<sub>2</sub>max to set the initial intensity of the exercise<sup>13,14,19,21,24</sup>. Additionally, although an often described benefit of HIIT is having shorter regimens, almost all of these identified studies feature regimens that last 30 min or longer<sup>11-15,18,19,21</sup>, with the exception of one with a slightly longer than 10 min regimen<sup>20</sup>, and another with 19 min across three different intensities<sup>16</sup>. To our knowledge, there are no reported animal studies that examine a 10 min or less HIIT regimen, or tailor the regimen to individual animals, with the exception of our published study<sup>17</sup> that serves as the basis for this protocol.

Here, we describe a protocol for HIIT in aged mice designed to model individualized, short session ( $\leq 10$  min) variants used recently in human studies<sup>7-11</sup>. The method includes a 10 min regimen on an inclined (25°) treadmill with a 3 min warm up, and four 1 min intervals at high intensity, interspersed with three 1 min active recovery segments. Advantages of the protocol include greater clinical relevance as it features strategies for tailoring intensity to individual animals, setting intensities that are not based on VO<sub>2</sub>max, thereby avoiding the need for metabolic treadmills, and modular design whereby the number of intervals and timings are easily adjustable. Additionally, within this protocol we provide instructions for two strategies for treadmill assessment, which include flat continuous and uphill interval, to examine endurance. Using these methods, we extend our previous findings that short session HIIT increased functional capacity in aged male mice<sup>17</sup>, and now demonstrate HIIT increases treadmill performance in aged female mice.

## PROTOCOL:

All studies and experimental protocols were approved by and in compliance with guidelines of the University at Buffalo and VA Western New York Animal Care and Use Committees.

### 1. Experiment setup and general advice

NOTE: A total of twenty-four female mice on a C57BL/6J background were used in this protocol starting at 23 months of age. The mice exhibited a conditional SIRT1 $\Delta$ ex4ERT2 mutation<sup>29</sup>, however, this was not induced in this experiment.

1.1. Ensure that mice receive permanent identifiers such as ear tags, RFID chips, or tail tattoos.

NOTE: It is recommended to also use temporary markings (e.g., permanent marker to mark tails) for quick identification during assessment periods.

1.2. Clean the treadmill with 0.25-0.5% bleach (v/v) or 70% (v/v) ethanol at the end of the day or to remove feces or urine between experimental trials. Dry solutions completely before initiating a new trial.

NOTE: Ethanol may increase wear to the treadmill belt. It is recommended that the treadmill be cleaned between each run if working with group-housed male mice in order to minimize in cage fighting. It is recommended to perform treadmill assessments at the same time of day at each time point in longitudinal studies<sup>30-32</sup>, and if other assessments are being performed, the order should remain the same. The investigator running the experiment should also be blind to the group designations of the mice.

### 2. Acclimatizing mice to treadmill apparatus

NOTE: Initiate acclimatization of mice 1 month prior to baseline experiments.

2.1. Set up the initial training program using the treadmill software (v3.4.7) in **Manual mode** (**Figure 1** and **Table 1**).

2.1.1 Open the treadmill software. Then, click on the file and open the experiment.

2.1.2 Input the values indicated in **Table 1** on the **Manual** tab (**Figure 1**) under **Acclimation**.

2.1.3 Input the session number as 1, the number of active channels as 1 to 6 depending on the number of mice, the number of visits to the grid as 10, and the number of shocks as 20.

NOTE: Shock intensity for this initial run is set at level 1 (0.46 mA).

2.2. Set the inclination of the treadmill to 0° (flat).

2.3. Hold mice by the tail when placing in the treadmill and place mice directly on the belt to avoid starting mice on the shock grid.

2.4. Use a brush or tongue depressor to keep mice away from the shock grid when training or assessment begins. Nudge mice to begin running to avoid unintended shocks.

2.5. After each mouse is given the initial training program, repeat the program as above with the shock intensity increased to level 2 (0.59 mA) with at least 15 min between trainings.

2.6. Administer the uphill treadmill acclimatization, one to two days after the initial training program.

2.6.1 Open the treadmill software. Then click on the file and then open the experiment.

2.6.2 Input the values on the **Basic** tab as indicated in **Table 2** under **Acclimation**.

2.6.3 Under **Shock Detection**, input the session number as 1, the number of active channels as 1 to 6 depending on the number of mice, the number of visits to the grid as 10, and the number of shocks as 20.

2.6.4 Click on the **Profile Mode** tab (**Figure 2**) and for Step 1, input a start speed of 0 m/min and an end speed of 5 m/min with a period of 5 s.

2.6.4.1. Add a warm up step 2 with a start speed of 5 m/min and end speed of 5 m/min for 30 seconds. Add a transition step that starts at 5 m/min and an end speed of 6 m/min for 5 s.

2.6.4.2. Add a test speed step that starts at 6 m/min and ends at 6 m/min for 20 s. Add a transition step that starts at 6 m/min and ends at 5 m/min.

2.6.4.3. Add a recovery interval that starts at 5 m/min and ends at 5 m/min for 20 s (**Figure 2**).

2.6.5 Repeat step 2.6.4 to add test speed steps as indicated in **Table 2** under acclimation.

2.7. Two weeks prior to baseline assessments, continue acclimatization by providing two treadmill flat aerobic capacity assessments as described in section 3, given on consecutive days.

2.8. One to two days after the second endurance training, provide two treadmill uphill anaerobic capacity assessments as described in section 4, given on the same day with a minimum of 30 min between trainings.

### 3. Flat continuous treadmill assessment

3.1. Create a treadmill program as described in step 2.1, using values indicated in **Table 1** under **Assessment**. Under **Shock Detection**, set the session number as 1, the number of active channels as 1 to 6 depending on the number of mice, the number of visits to the grid as 10, and the number of shocks as 20.

3.2. Remove mice from the instrument that reach endpoint criteria during the trial.

NOTE: This step is to avoid mice inadvertently touching the shock grid of neighboring mice and affecting data. Parameters include the total time on belt, the distance travelled, and the shocks to visits ratio. It is recommended to perform two assessments at each time point, separated by at least one day between assessments.

#### 4. Uphill interval treadmill assessment

4.1. Create a treadmill program as described in step 2.6, using values indicated in **Table 2** under **Assessment**. Under **Shock Detection**, set the session number as 1, the number of active channels as 1 to 6 depending on the number of mice, the number of visits to the grid as 5, and the number of shocks as 10.

4.2. Set the interval field in the treadmill program to 0.5.

NOTE: This allows for data points to be collected every second instead of every minute, which aids in identifying the speed at endpoint for each mouse. The program will automatically stop after the 50 m/min stage due to software limitations. Parameters include the time on belt, the distance travelled, and the test speed of the last successfully completed stage before endpoint. The latter is used for determining baseline intensity for the HIIT regimen described in step 5. It is recommended to perform two assessments per time point, separated by at least 30 min.

#### 5. Short session high intensity interval training

5.1. Set the treadmill uphill (25°) and remove the plastic cover.

5.2. Determine the intensity for each mouse. Use the speed of the last successfully completed stage (Step 4.5) and find the intensity group and corresponding **Base**, **Sprint**, and **Dash** speeds using **Table 3**.

5.3. Open the treadmill software and click on file to create a new program.

5.3.1 Click on the **Profile Mode** tab and for Step 1, input a start speed of 0 m/min and an end speed as indicated under **Base** speed as indicated in **Table 3** with a period of 5 s.

5.3.2 Add a warm up step 2 with a start speed at **Base** speed and end speed at **Base** speed for 180 s.

5.3.3 Input 3 intervals each with 1) a transition step that goes from **Base** to **Sprint** for 5 s, 2) an step at **Sprint** speed for 60s, 3) a transition step that goes from “sprint” to **Base** over 5 s, and 4) a step for a recovery interval at **Base** speed for 60 s.

5.3.4 Add a transition step that goes from **Base** to **Sprint** speed over 5 s and a final step that goes from **Sprint** to **Dash** speed over 60 s.

NOTE: Programs can be saved, modified, and reloaded.

5.4. Perform exercise three days a week with at least one day of rest between sessions (i.e., Monday, Wednesday, Friday).

5.5. Rewrite exercise programs to increase **Base**, **Sprint**, and **Dash** speeds 1 m/min every two weeks.

NOTE: No electric shocks are given during the exercise.

5.6. Motivate mice to run using a makeup brush (recommended) or tongue depressor to lightly motivate mice that fall near or on the shock grid. If a mouse does not respond retry in 5 s, then 10 s, and then retry every 30 s or during a recovery interval until the end of the session.

5.7. Move mice that cannot be motivated to complete the first three intervals in two consecutive exercise sessions to lower intensity groups (**Table 3**).

5.8. Move mice that do not require motivation in two consecutive exercise sessions to higher intensity groups (**Table 3**).

5.9. Take sedentary control mice in cages place beside the treadmill as it runs. Alternatively, sedentary controls can be placed in the lanes of a non-moving treadmill for 10 min.

## REPRESENTATIVE RESULTS:

A total of twenty-five female mice were bred and aged in house. The C57BL/6J background mice exhibited a SIRT1 $\Delta$ ex4ERT2 mutation<sup>29</sup>; however, the conditional mutation was not induced and therefore all mice exhibited full length Sirtuin1 (data not shown). At 24 months of age, mice were assessed for treadmill endurance and uphill sprint capacity prior to and after the administration of two months of HIIT exercise ( $n = 14$ ), or remaining cage sedentary ( $n = 11$ ). Our data show all 14 mice in the HIIT group increased treadmill time on belt compared to 7 of 11 of the SED mice (**Figure 4A**, paired student's T-Test  $p < 0.0001$  for HIIT and  $p < 0.14$  for SED). In total HIIT mice exhibited greater improvement in time on belt based on the better of two trials (HIIT:  $18.2 \pm 10.5$  min to  $31.8 \pm 13.7$  min, delta  $13.6 \pm 7.5$  min versus SED:  $19.9 \pm 10.0$  min to  $25.3 \pm 7.3$  min, delta  $5.3 \pm 11.4$  min, unpaired student's T-test  $p < 0.0391$ ).

Additionally, we detected greater increase in uphill treadmill capacity as 12 of 14 HIIT mice increased maximal speed while we observed decline in 8 of 11 SED mice (**Figure 4B**, paired

student's T-Test  $p < 0.0022$  for HIIT and  $p < 0.85$  for SED). HIIT mice also demonstrated greater improvement in maximal speed compared to SED mice based on the better of two trials (HIIT:  $14.6 \pm 4.3$  m/min to  $17.6 \pm 5.5$  m/min, delta  $3.0 \pm 3.0$  m/min versus SED:  $16.9 \pm 3.8$  m/min to  $16.5 \pm 5.0$  m/min, delta  $0.5 \pm 5.0$  m/min, unpaired student's T-test  $p = 0.0441$ ). Shock tolerance was similar between the two groups of mice (Shock to grid visit ratio baseline - SED:  $1.2 \pm 0.1$  versus HIIT:  $1.4 \pm 0.4$ ,  $p=0.24$ ; endpoint - SED:  $1.2 \pm 0.4$  versus HIIT:  $1.3 \pm 0.4$ ,  $p=0.46$ ).

#### FIGURE AND TABLE LEGENDS:

**Figure 1. Treadmill software manual mode parameters for treadmill endurance assessment.**

The treadmill software manual mode allows adjustment for protocols involving continuous belt acceleration.

**Figure 2. Treadmill software profile mode program parameters for uphill sprint assessment.**

The treadmill software profile mode, used in conjunction with the options on the manual mode tab (**Figure 1**), allows the ability to create protocols with custom speed intervals.

**Figure 3. High Intensity Interval Training regimen schematic.** Base, Sprint, and Dash speeds are indicated in **Table 3**. Speeds increase by 1 meter/min every two weeks.

**Figure 4. Impacts of HIIT on treadmill performance in aged mice.** Twenty-four month old female mice were given HIIT ( $n=14$ ) or remained sedentary (SED,  $n=11$ ) for 2 months. Treadmill endurance (**A**) was assessed before and after the exercise intervention as a best of two trials given on consecutive days on a flat treadmill. Uphill sprint capacity (**B**) was assessed as the best of two trials administered with no less than 30 min of rest on an inclined ( $25^\circ$ ) treadmill. Lines indicate individual mice. B is baseline and E is endpoint, \*\*\* indicate a  $p$ -value  $< 0.001$  and \*\*\*\* indicates  $p < 0.0001$  as determined by a paired student's T-test comparing within group change between baseline and endpoint.

**Table 1. Program parameters for treadmill endurance acclimatization and assessment.**

**Table 2. Program parameters for uphill sprint acclimatization and assessment.**

**Table 3. Intensity designations for HIIT exercise mice.**

#### DISCUSSION:

Benefits from short sessions are a key aspect of high intensity interval training that captures scientific and public interest. However, animal studies rarely investigate HIIT regimens that are 10 min or less. Here, we describe a protocol for a 10 min short session HIIT uphill treadmill exercise regimen that enhanced treadmill performance in aged female mice and which we previously have shown to increase physical performance in aged male mice<sup>17</sup>. A strength of our protocol is that in addition to the protocol being only 10 min in length, the design is modular such that the number of intervals can be added or removed to make protocols that are even one min in length. Additionally, the length of the intervals can be modified providing multiple strategies to titrate exercise and evaluate the impacts.



Another strength of this protocol is that we include a system to tailor the exercise regimen to the physical capabilities of individual mice, which to our knowledge only one other study<sup>33</sup> aside from our report in aged male mice<sup>17</sup> has used. Although individually tailoring an exercise intervention introduces variability into the experimental design, an important advantage of this method is that it better models the administration of prescribed exercise in human clinical settings. Furthermore, animals do not train at intensities that are too easy or difficult, which could be a significant factor in experiments where the animal population exhibits diversity in exercise capacity (i.e. during aging). One other advantage of this methodology is that it does not require the use of metabolic treadmills to determine VO<sub>2</sub>max as the exercise intensity is tailored to the performance of the animal, a concept that is further examined by Picoli *et al.*<sup>34</sup>.

This protocol includes two assessments for treadmill performance, in the form of a slowly accelerating flat treadmill test and an inclined treadmill incrementing interval test, respectively. There are multiple protocols that have been published for determining continuous treadmill performance in mice using treadmills<sup>16,34,35</sup>, including a JoVE article<sup>36</sup>. Multiple parameters differ across these studies and across the literature, including: inclination (usually 0° or 5° for endurance testing), shock intensity (ranging from 0.25-1.12 mA), and rules for defining exhaustion. It was also noticed that 5 consecutive shocks were commonly used to define exhaustion across multiple studies. Although this strategy may induce greater exhaustion than the rules we have used in our study and this protocol, this system also assumes all mice have similar pain thresholds, which may not be true depending on experimental conditions. Our definition of exhaustion of 10 visits or 20 total shocks provides a framework to assess if there are different pain thresholds. Ultimately, there are advantages and disadvantages to both strategies and the decision for defining the endpoint should be best aligned with the goals of the study. Additionally, some treadmill assessment protocols for mice have been designed without the use of shocks as a stimulus<sup>37</sup>. Although there are advantages to this approach<sup>37</sup>, some drawbacks to consider include the use of a determination of exhaustion that is subjective and the potential difficulty of assessing multiple mice simultaneously.

Additionally, others have used work and power as parameters to describe treadmill performance<sup>35,38,39</sup>. The speed of the belt and rate of acceleration are also diversely represented in the literature. As most protocols feature increasing speed, the result will yield a blend of aerobic and anaerobic capacity in the mouse, with slower protocols providing more focus on the aerobic component. We therefore designed our uphill treadmill interval test to provide greater focus on the anaerobic component<sup>17</sup>. To achieve this, our protocol features short test intervals with active recovery between tests, allowing for mice to achieve higher speeds and therefore greater anaerobic utilization relative to our flat continuous treadmill endurance assessment. For our protocol we used an active recovery period of 20 s between intervals, the half time for re-oxygenation of muscle tissue in human tissue<sup>40</sup>. However, we note a limitation of this method is that the specific impacts on anaerobic metabolism have yet to be elucidated, and including an examination of anaerobic parameters such as glycolytic metabolism, phosphocreatine utilization, and lactate kinetics would strengthen this method. In addition, new studies that investigate the impacts of changing stage and active rest durations,

inclination, and different speed increments would also strengthen this method.

The methodology described in this protocol, including the two treadmill assessments, are designed for older animals in particular, and as such includes greater time for acclimation of mice to the protocols. Proper acclimation is a critical step to experimental design to ensure endpoints are due to exhaustion and not insufficient learning by the mouse. Improper acclimation is likely apparent in longitudinal studies - yet, importantly, may not be noticed in cross-sectional studies. Although, shorter acclimation protocols might be possible for younger cohorts, in our experience older mice require greater acclimation and a published protocol for exercise training in aged rats initiates acclimation 1 month prior with 10 total days given for treadmill acclimatization<sup>41</sup>. However, we agree with Castro et al. that providing excessive acclimation may impact the behavior of and impacts on the mice<sup>35</sup> and his recommendation of 3 to 10 days total<sup>35</sup>, for which our acclimatization protocol is in line with these recommendations. Furthermore, older animals also exhibit greater diversity in exercise performance and for this reason we include two trials per assessment time point. Younger animals display greater homogeneity in performance and therefore a single trial may be sufficient. Furthermore, the diversity of aged animals may make direct cross-sectional comparisons of an intervention difficult to interpret, and more success may be achieved by comparing the change from baseline as was done in the analysis of the aged female mice. We also indicate as a caveat that we have not tested our short session HIIT protocol in younger mice, where it is possible a ceiling effect in performance may mask the benefits of our short session HIIT protocol. The protocol was also used only in the C57BL/6J mouse strain, and therefore the impact of this exercise in other mouse strains remains to be elucidated. Additionally, the C57BL6/J mice used in this experiment exhibited a SIRT1 $\Delta$ ex4ERT2 mutation<sup>29</sup>, that was not induced. These mice were aged in house from birth; however, insufficient numbers were available at 24 months of age to power 4 groups. We therefore focused the experiment on just HIIT and sedentary groups. We note that both the present cohort of aged female mice and our previously published cohort of aged male mice<sup>17</sup> both improved in physical performance using this short session HIIT protocol.

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#### DISCLOSURES:

The authors have nothing to disclose.

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# Figure 1

Manual	Profile Mode	Inclination Setup
<b>Interval</b> <input type="text" value="1"/> min		
<b>Duration</b> <input type="text" value="90"/> min		
<b>Treadmill</b>		
<input type="checkbox"/> Enable Profile Mode		
<b>Accel Step</b> <input type="text" value="1.0"/> m/m		
<b>Accel Int</b> <input type="text" value="120"/> sec		
<b>Max Speed</b> <input type="text" value="35.0"/> m/m		
<b>S_Speed</b> <input type="text" value="5.0"/> m/m		
<b>Belt Speed</b> <input type="text" value="0.0"/>		
		
<b>Shock Rep_Rate</b>		
<input type="radio"/> Off <input type="radio"/> 2Hz		
<input checked="" type="radio"/> 1Hz <input type="radio"/> 3Hz		
<b>Shock Intensity</b> <input type="text" value="3"/>		
<b>Shock Detection</b>		
<b>Session Number</b> <input type="text" value="1"/>		
<b>Number of Active Channels</b> <input type="text" value="1"/>		
<b>Number of Visits to Grid (1-999)</b> <input type="text" value="10"/>		
<b>Number of Shocks (1-999)</b> <input type="text" value="10"/>		

## Figure 2. Uphill Sprint Assessment Schematic

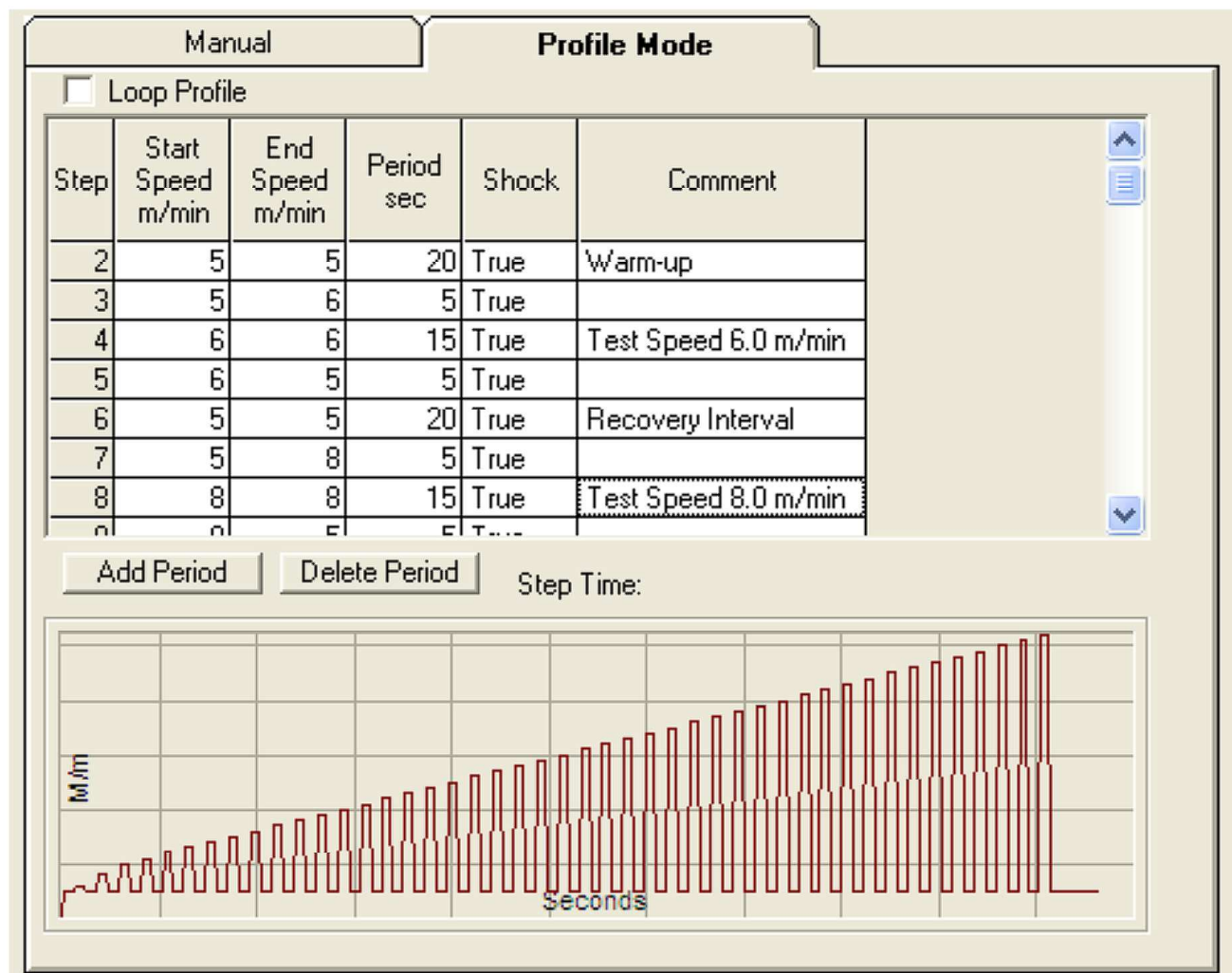


Figure 3. HIIT Schematic

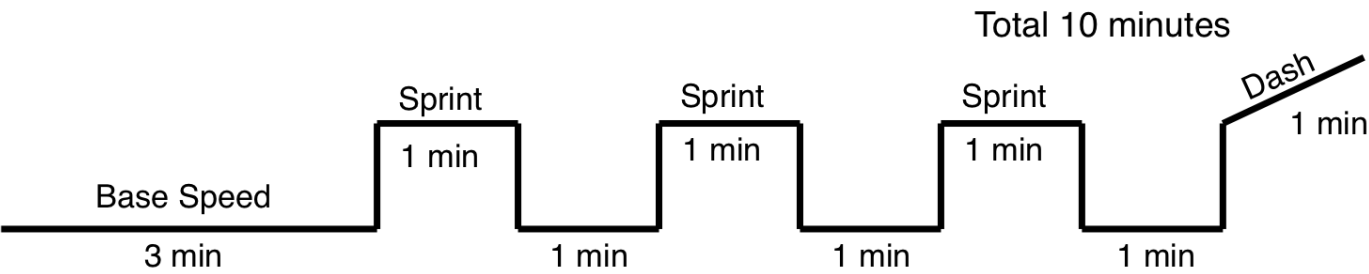
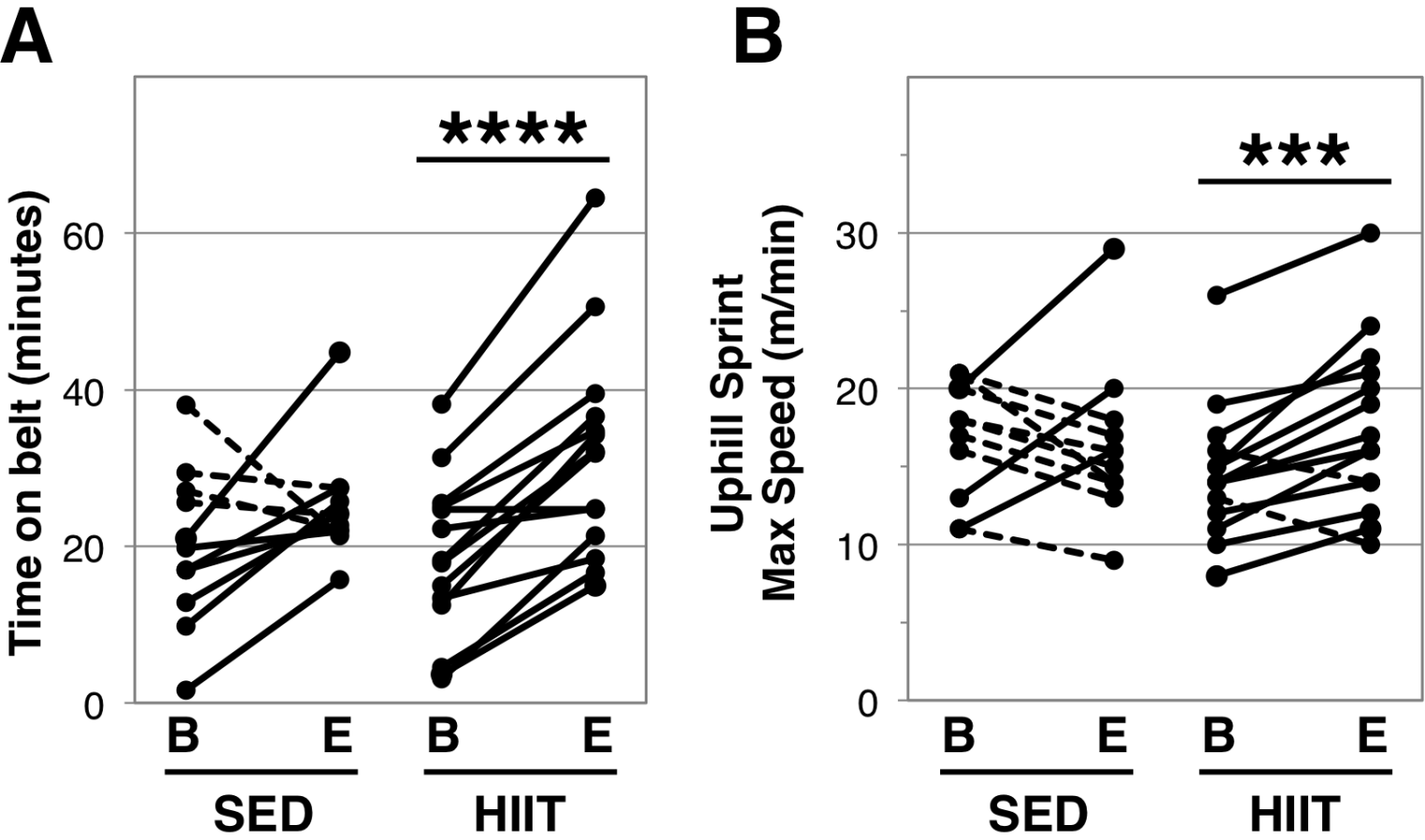




Figure 4. Treadmill impacts of HIIT



Program parameter	Acclimation	Assessment
Interval	1	1
Duration	5	90
Enable Profile Mode	Unchecked	Unchecked
Accel Step	1.0 m/min	1 m/min
Accel Int	60 seconds	120 seconds
Max Speed	10 m/min	35 m/min
S_speed	5 m/min	5 m/min
Shock Rep Rate	1 hz	1 hz
Shock Intensity	1 then 2	3

Program parameter	Acclimation	Assessment
Interval	0.5	1
Duration	10	90
Enable Profile Mode	Checked	Checked
Accel Step	N/A	N/A
Accel Int	N/A	N/A
Max Speed	15 m/min	35 m/min
S_speed	5 m/min	5 m/min
Shock Rep Rate	1 hz	1 hz
Shock Intensity	2	3
PROFILE MODE (Refer to Figures 1 and 2 to set up program parameters)		
Test intervals	8, 10, 10, 10 m/min	6, 8, 10, 11, 12 ... 51, 52 m/min

Intensity Group	Baseline Uphill Sprint Score	Initial Exercise Speeds		
		Base	Sprint	Dash
0	< 8 m/min	5 m/min	7 m/min	10 m/min
1	8-16 m/min	5 m/min	10 m/min	15 m/min
2	17-25 m/min	8 m/min	13 m/min	18 m/min
3	>25 m/min	11 m/min	16 m/min	21 m/min

Name of Material/ Equipment	Company	Catalog Number
Exer-3/6 Open Treadmill w/ Shock, Detection, auto-calibration and PC Interface/Software	Columbus Instruments	1055-SDRM
Bleach	Varies	Varies
Ethanol	Varies	Varies
Make-up Brush (large)	Varies	Varies

### **Comments/Description**

The Columbus Instruments 3/6 treadmill allow up to 6 mice or 3 rats simultaneously.

The device comes with controllers to allow manual control of treadmill belt speed and shock intensity, or connections to a computer and software to run and control these elements.

0.25-0.5% Bleach solution (V/V) is used to clean the treadmill belt between sessions

70% ethanol solution (V/V) can alternatively be used to clean treadmill belt between runs and sessions.

A make-up brush provides a soft surface and ample length to motivate mice to continue exercise.



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