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TITLE:

Transcutaneous Auricular Vagus Nerve Stimulation (taVNS) in Humans

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SUMMARY:

A methodological description of the technique, potential targets, and proper administration of transcutaneous auricular vagus nerve stimulation (taVNS) on the human ear is described.

ABSTRACT:

Non-invasive vagus nerve stimulation (VNS) may be administered *via* a novel, emerging neuromodulatory technique known as transcutaneous auricular vagus nerve stimulation (taVNS). Unlike cervically-implanted VNS, taVNS is an inexpensive and non-surgical method used to modulate the vagus system. taVNS is appealing as it allows for rapid translation of basic VNS research and serves as a safe, inexpensive, and portable neurostimulation system for the future

treatment of central and peripheral disease. The background and rationale for taVNS is described, along with electrical and parametric considerations, proper ear targeting and attachment of stimulation electrodes, individual dosing *via* determination of perception threshold (PT), and safe administration of taVNS.

INTRODUCTION:

Cranial nerve X, better known as the vagus nerve, is a large nerve tract that originates in the brainstem of the central nervous system and travels throughout the periphery, targeting every major organ in the thorax and abdomen (**Figure 1**)¹. Vagus nerve stimulation (VNS) involves surgical implantation of bipolar electrodes around the left cervical branch of the vagus nerve. Electrical pulses are delivered to the vagus nerve *via* an implanted pulse generator (IPG) surgically implanted in the chest². Although VNS is currently FDA-approved for epilepsy, refractory depression, and chronic obesity, it is a costly procedure requiring a hospital visit and surgery. Long-term safety of VNS is well established, and the majority of safety considerations regard current intensity related side effects (hoarse voice, throat pain) without serious stimulation-related adverse effects over the past 25 years of its clinical use³.

Recently, a noninvasive form of VNS known as transcutaneous auricular vagus nerve stimulation (taVNS) has emerged⁴. taVNS delivers electrical stimulation to the auricular branch of the vagus nerve (ABVN), an easily accessible target that innervates the human ear⁵. Over the last decade, several groups have demonstrated the safety and tolerability of this method⁶⁻⁸, including central and peripheral nervous system effects^{9,10}, and behavioral effects^{7,11-13} in neuropsychiatric populations. taVNS is also being explored in individuals as a promising enhancer of cognitive^{14,15} and social functioning¹⁶⁻¹⁸. As taVNS is becoming established, it offers the ability for researchers and clinicians to rapidly translate the promising VNS research that has been described in various disorders ranging from neurological and psychological trauma¹⁹⁻²¹, addiction²², inflammation²³, and tinnitus^{24,25}.

In principle, taVNS is methodologically similar to conventionally administered transcutaneous electrical nerve stimulation (TENS) used to treat musculoskeletal pain disorders²⁶. The difference is that taVNS is delivered to specific anatomical ear targets that are believed to be innervated by the ABVN⁵. The field is still determining optimal stimulation targets²⁷, although the two most common placements are the anterior wall of the outer ear canal (tragus) and the cyma conchae. Sham stimulation may be conducted by stimulating the earlobe of the ear, an area believed to have minimal ABVN innervation (**Figure 2**). Alternatively, sham may be delivered via a passive control method in which electrodes are attached to active sites, but no stimulation is delivered. Stimulation parameters may vary between groups, however according to the literature, stimulation is delivered in a pulsatile fashion (pulse width: 250-500 μ s, frequency: 10-25 Hz) and delivered at an individualized constant current (<5 mA). Stimulation current varies by individual and experimental protocol, with many groups exploring various intensities as a function of an individual perceptual threshold (PT). The PT is defined as the minimum amount of current eliciting a perceived sensation at the target site and is usually determined *via* parametric estimation by customized sequential testing (PEST) software described in this report.

taVNS is a safe technique that may be administered in the laboratory or clinical setting. Side-effects of taVNS are minimal, with skin irritation or redness being the most common side-effect. Most taVNS studies explore stimulation of the left ear, as it is believed to be safer, although data in a large trial (Badran *et al.* 2018) reveal that right-sided stimulation has no increases in the risk of adverse events. Due to the wealth of literature in unilateral-left side stimulation, we will illustrate the typical taVNS set-up for laboratory studies investigating the use of left-sided taVNS as an intervention.

PROTOCOL:

This experimental protocol illustrates a typical taVNS set-up for use in a laboratory or clinical setting in which we target stimulating the anterior wall of the auditory canal (tragus) in a supine posture with an 8mm diameter round metal electrode. These methods can be mimicked for alternative active treatment sites by simply changing electrode position to the cymba concha. All methods and procedures have been IRB approved by the Human Research Protection Program (HRPP) at City College New York.

1. Materials

1.1. Ensure all materials required to administer taVNS are prepared (**Figure 3**). The taVNS stimulator may be either a battery driven device that meets local safety regulations or powered from a conventional electrical outlet with built-in safety mechanisms that prevent unintended electrical surges. A constant current (current controlled) stimulator with a maximum output of 5 mA is required.

1.2. For taVNS, use stimulation electrodes made of a round conductive metal (tin, Ag/AgCl, gold) combined with a conductive medium such as electrolyte gel or conductive paste (see table of materials). Alternatively, use conductive electrodes made with flexible conductive carbon electrodes and conductive gel that may or may not be adhesive. Never place electrodes directly on the skin without a conductive medium, as this may pose unnecessary risk to the participant and can cause discomfort or pain.

1.3. Use computer running script software (see **Table of Materials**) that is programmed and used to control the stimulator and initiate stimulation with specific parameters. These parameters include current intensity (mA), pulse width (μ s), frequency (Hz), duty cycle (On/Off time, s), session duration (min).

1.4. Use alcohol preparation pads (70% isopropyl alcohol) to prepare the skin surface before attaching electrodes to the ear. This removes surface oils from the skin surface and reduces the resistance of the skin, ensuring stimulation is delivered at safe power levels.

2. Ear Targeting and Skin Preparation

2.1. Use the following general inclusion criteria for conducting taVNS in the research setting: Age 18-70, no facial or ear pain, no recent ear trauma, no metal implants including pacemakers, not pregnant.

2.2. In the experiments involving healthy participants in a laboratory setting, use the following exclusion criteria: personal or family history of seizure, mood, or cardiovascular disorders, dependence on alcohol or recent illicit drug use, on any pharmacological agents known to increase seizure risk.

2.3. Seat the participant on a comfortable bed or chair in a supine or other relaxed position with legs elevated and head supported.

2.4. Inspect the left ear of the participant. Ensure no jewelry is attached and all make-up and lotion are removed. Confirm there are no skin-related contraindications at the site of stimulation, including sun burn, cuts, lesions, open sores.

2.5. Find the stimulation target, landmarked by the anterior wall of the outer ear canal externally by finding the tragus. Stimulation will be delivered to the portion of the ear canal directly behind the tragus (**Figure 4**).

2.6. Use an alcohol prep pad to gently scrub the target site, both internally and externally, to decrease skin resistance and increase conductance.

3. Electrode Preparation and Placement

3.1. If using non-disposable electrodes, visually inspect electrodes to ensure clean, corrosion-free surface is exposed. Ensure that electrodes are disinfected to prevent spread of bacteria between subjects. This can be done using alcohol or sterilization wipes to scrub the electrodes. If using disposable electrodes, skip to step 3.2.

3.2. Spread a thin layer of conductive paste to the surface of the electrode evenly. This will distribute electricity to the stimulation site. For an 8 mm diameter round electrode, a pea-sized amount of paste is sufficient. Spread the paste using a narrow wooden applicator to form a thin layer <1 mm of paste on both electrodes.

3.3. Connect electrode cables to the stimulation device while the device is turned off and verify the polarity of the electrodes (red/positive electrode – anode, black/negative electrode - cathode). This is an important detail as targeting is polarity specific - the anode (red/positive terminal) is the electrode placed inside the ear canal and targeting the anterior wall of the outer ear canal. The cathode (black/negative terminal) sits on the outside of the ear attached to the tragus. For sham stimulation, the anode is placed on the anterior side of the ear.

3.4. Clip the spring electrode onto the tragus with the anode making contact with the anterior wall of the outer ear canal and the cathode contacting the anterior part of the tragus.

NOTE: If conducting sham stimulation, clip the electrode onto the earlobe (active control). Alternatively, sham stimulation may be delivered by attaching stimulation clips to active site and delivering no electrical current (passive control).

3.5. As subjects will feel the pressure of the electrodes clipped to their ear, ensure this pressure is not uncomfortable or disruptive to regional blood flow as demonstrated by pale white skin at clip site or physical pain sensed by the subject. After this point, determine perceptual threshold (PT) which will be described in the next procedural step.

4. Determination of Perceptual Threshold (PT)

NOTE: Perceptual threshold is a critical value used to determine the power of taVNS stimulation. This value is defined as the minimum amount of electricity required to perceive electrical stimulation on the skin described as a pricking or tingling sensation.

4.1. Determine the PT using a simple step-up and step-down binary parametric search. First turn on the stimulator and set the output to 3 mA. Deliver a 1 second train of taVNS stimulation at desired pulse width (typically 250 μ s – 500 μ s) and frequency (25 Hz, can vary based on application).

4.2. Ask the subject whether they felt the stimulation. Sensation is generally reported as a “tickle” or “pricking” sensation.

4.2.1. If YES, turn down stimulation intensity by 50% and repeat step 4.2. If NO, increase stimulation intensity by 50% and repeat step 4.2.

4.3. Repeat the process described in step 4.2 until recording a minimum of 4 “YES” responses in which the 4th YES response must come after a NO. The intensity (in mA) of the PT will be the value at which the subject says their fourth YES response to.

4.4. Use the example PT threshold finding is listed in **Table 1** to assist in PT determination.

5. Delivering Stimulation

5.1. Once the subject is comfortable stimulation electrodes with properly attached to the desired target, and the perceptual threshold determined at the desired pulse width and frequency, begin the stimulation.

5.2. Use a computer running a pulse generating GUI (e.g., stimDesigner, freeware included with this manuscript) connected to a data acquisition unit (DAQ) to drive the stimulation system. The software should output TTL pulses as programmable settings (**Figure 5**). The TTL pulses will be sent via a BNC cable to the stimulator “trigger in” port. This interface software/stimulator

interface allows modulation of frequency, duty cycle (on/off time), and session duration (**Figure 6**). The GUI used is attached as a free, open-source resource with this manuscript.

5.2.1. Ensure that stimulation is delivered at super-threshold levels, such as 200% of PT^{8,9}. For example, if the PT was determined to be 0.8 mA, stimulation will be delivered at 1.6 mA.

5.2.2. Ensure that the guidelines for duty cycles are followed when conducting long stimulation sessions. Typical duty cycles have 30-60 s “on” periods and 60-120 s “off” periods, or 20% -50% duty cycles.

5.2.3. Vary the length of stimulation session (total time). Studies suggest that 30-60 minute stimulation sessions at a 25% duty cycle is safe and free of any acute side effects or adverse events. These sessions can be repeated with 12-24 hours between sessions safely.

NOTE: taVNS safety is unclear for longer periods of stimulation sessions, larger percentage duty cycles (>40%), accelerated paradigms, and higher stimulation current doses.

6. After taVNS

6.1. When stimulation is completed, record objective data regarding the stimulation discomfort and side effects. Although taVNS, like implantable VNS, has limited safety concerns^{8,28}, monitor and record sensation, discomfort, and any adverse events on a rating from 0-10²⁹.

6.2. Remove the stimulation electrode from the ear and clean residual conductive paste from the subject’s ear using an alcohol prep pad.

6.3. Use alcohol to clean and disinfect the stimulation electrode immediately upon removing from the subject’s ear.

6.4. Inspect the ear for redness or irritation at the stimulation site and record any observations.

REPRESENTATIVE RESULTS:

When proper skin preparation is conducted, perceptual thresholds are inversely correlated with stimulation pulse width. As pulse width increases, the perceptual threshold decreases (**Figure 7**). Initial studies by this group exploring the effect of pulse width on PT in healthy individuals (meeting inclusion/exclusion criteria listed above), determined that the combined overall (n=15, 7 female, mean age 26.5 ± 4.99) PT at 100 µs = 3.92 ± 1.1 mA; 200 µs = 2.24 ± 0.74 mA; 500 µs = 1.24 ± 0.41 mA. These thresholds suggest that a constant current stimulator with capacity of delivering up to 5 mA of current is required for stimulation of 500 µs pulse width parameters, and a minimum of a 10mA stimulator is required for lower pulse widths (**Table 2**). Fine tuning of the current is required, with increments of 0.1 mA are necessary for precise stimulation.

Delivering stimulation at 200% PT is tolerable and relatively pain free as demonstrated by pain numerical rating scales (NRS) scales^{9,30}. The NRS scale is a rating system for pain from 0-10 in which individuals report pain or discomfort²⁹. Both Active and Sham stimulation rate similarly low pain levels (NRS <3 for all stimulation pulse widths. More specifically, the biologically-active pulse width of 500 μ s delivered at 25 Hz is reported on average to rate as Active = 1.98 ± 0.83 , Sham = 2.17 ± 1.27 (n=25, 9 female, mean age 25.16 ± 4.16 years) (Table 3). Pain ratings for other parameters are no more painful than the 25 Hz parameter and the details can be found in the groups' prior work³⁰.

Safety and tolerability of 30 min to 1 hour sessions at a 20-50% duty cycle has been widely reported in the literature with some studies delivering multiple sessions in the same day spread 12-15 hours apart^{12,31}. No serious adverse events have been reported from 60 subjects participating in several series of experiments with subjects participating from 1 to 8 repeated visits spread a minimum of 24 hours apart.

taVNS, when administered as reported in this manuscript, has been demonstrated to modulate the autonomic nervous system, induce functional brain activity changes as measured by fMRI BOLD, and piloted to treat neuropsychiatric disorders and aid in rehabilitation.

FIGURE AND TABLE LEGENDS:

Figure 1: Vagus Nerve Efferent Projections and Cross-section. A. Efferent projections of the vagus nerve target every major organ on the body with wide effects on bodily function B. Cross-section of the vagus nerve, demonstrating the inside anatomy of the nerve as a series of bundles of nerves all contained within one major pathway.

Figure 2: taVNS Ear Targets. Targeting the ABVN can be accomplished by stimulating the anterior wall of the outer ear canal, landmarked notably by the tragus (A1), or cymba conchae (A2). Sham stimulation is administered to the earlobe (S).

Figure 3: Key Components. The minimum required components for proper administration of taVNS are the following A) ear stimulation electrodes, B) conductive gel and alcohol prep pads, C) computer capable of sending and receiving TTL pulses to a D) constant current stimulator to trigger stimulation.

Figure 4: Example Setup. This photo shows an individual receiving taVNS of the left ear while in position to undergo an experimental paradigm.

Figure 5. Screenshot of the GUI used for stimulation.

Figure 6: Electrical Stimulation Waveform Manipulations. Direct square wave electrical current can be delivered at various parameters. This figure demonstrates key properties of the waveform that can be changed in order to achieve desired biologic effects.

Figure 7: Perceptual Threshold Values at Increasing Pulse Widths. As pulse width increases, perceptual threshold (PT) decreases. Most healthy individuals will have a PT within 2 standard deviations (SD) of these mean values.

Table 1: Example of How to Determine Perceptual Threshold (PT). This table shows an example sequence of Yes/No responses used to parametrically determine PT.

Table 2: Stimulation Current Levels. Values of stimulation current in mA (200% PT) for each pulse width (n=15).

Table 3: PT, Stimulation Current, and Pain Values for Suggested Stimulation Parameters. Values of stimulation current in mA (200% PT) for each pulse width (n=25).

Supplemental File: Freeware GUI used in this protocol.

DISCUSSION:

As in all novel modalities, all the described steps are critical in the safe administration of taVNS. Of ultimate concern is subject safety, which includes not only mitigating risks before taVNS *via* proper screening, but also monitoring subjects during stimulation for discomfort, pain, or adverse events. Here are the three most important considerations for administering taVNS. Screening for taVNS contraindications – contraindications are as follows: any current or past history of cardiovascular disorders, facial or ear pain, recent ear trauma, metal implants above the level of the neck. For proper subject skin preparation, removing any surface oils, dirt, or makeup from the surface of the skin with alcohol helps with conductivity of the electrodes, reduces stimulation voltage required to drive the stimulator, and ultimately results in a more tolerable and safe stimulation session. It is encouraged to use a stimulator and electrodes meeting Low Output Transcranial Electrical Stimulation (LOTES) guidelines³². LOTES sets guidelines and industry standards for electrical stimulators that are built for stimulation of the head and neck and it is encouraged for groups to read this document before building their own systems. It is recommended to use either a FDA-cleared plug-in stimulator (see **Table of Materials**), or a low voltage (<50 V), battery powered, constant current stimulator with appropriate safety measures built-in to avoid unintended over-delivery of current to the stimulation site. Ensure that electrodes are manufactured and assembled for specific use in taVNS. Ensure that the current manufacturing and engineering guidelines are followed as a reference if lab-made customized systems are used.

One consideration for taVNS is to ensure that the voltage output of the constant current stimulator can surmount the resistance of the skin and deliver the current required for stimulation. Ohm's law ($V=IR$) demonstrates the relationship between current (I) and skin resistance (R). A minimum of a 20 V tabletop stimulator is recommended to avoid an underpowered system. Heat generated from the scalp or environment may degrade the conductive paste. If this occurs, it is recommended to stop stimulation and re-prepare skin and electrodes with new conductive paste.

A limitation of taVNS is the vast parameter space. It is unknown as to which is more important – pulse width or frequency. There is a lacking data in recent taVNS trials that answer such questions. The various behavioral effects are derived from a variety of pulse widths, frequencies and stimulation currents^{13,33-39}.

At this time, it is suggested that the 500 μ s pulse width to be the most biologically active⁹. With respect to frequency, it has been demonstrated that 25 Hz is an effective frequency, although current investigations into optimal such as higher frequencies (>25 Hz), bilateral stimulation (left and right ears), and investigational burst paradigms are being conducted. Studies exploring different parameters of stimulation, alternative stimulation sites, and duty cycle optimization are needed to advance and refine the taVNS method.

taVNS is a promising non-invasive alternative to conventional VNS. taVNS provides an inexpensive (<\$5,000 in the demonstrated experimental setup, cost heavily dependent on type of stimulator used) and straightforward method that can be used to translate positive findings in animal models exploring the use of VNS on a variety of disorders, noninvasively modulate the autonomic nervous system, and potentially miniaturized and optimized for at-home neuromodulation for the treatment of neuropsychiatric and other disorders.

The future potential and possible applications of taVNS are vast. taVNS can serve as a promising adjunct or standalone treatment for neuropsychiatric disorders such as depression and epilepsy, taVNS-paired rehabilitation training to restore or accelerate learning of a behavior⁴⁰, decrease inflammatory response^{41,42}, and can potentially be used to enhance performance and autonomic function^{8,10}.

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DISCLOSURES:

The authors have nothing to disclose.

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502

Figure 1

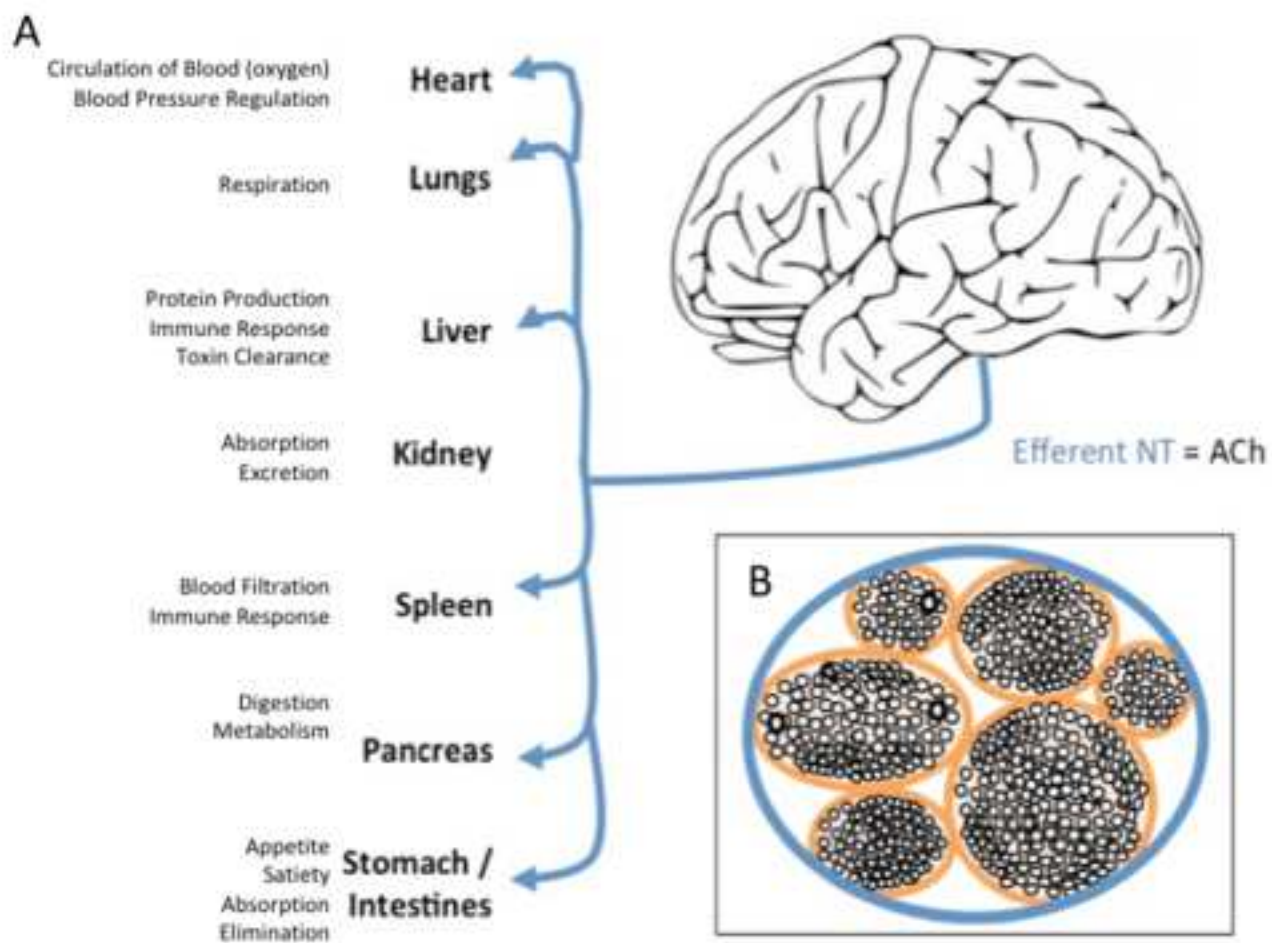


Figure 2

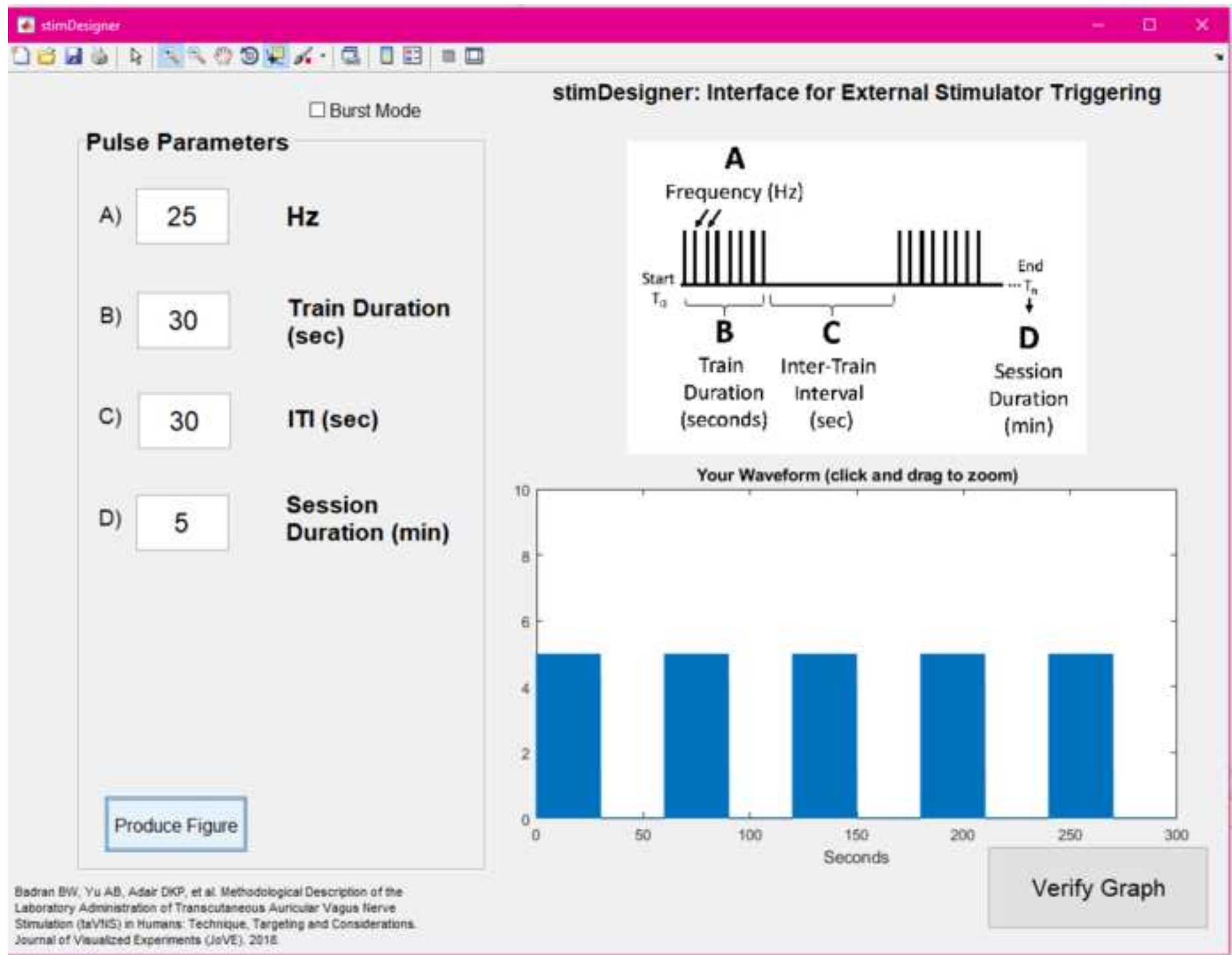


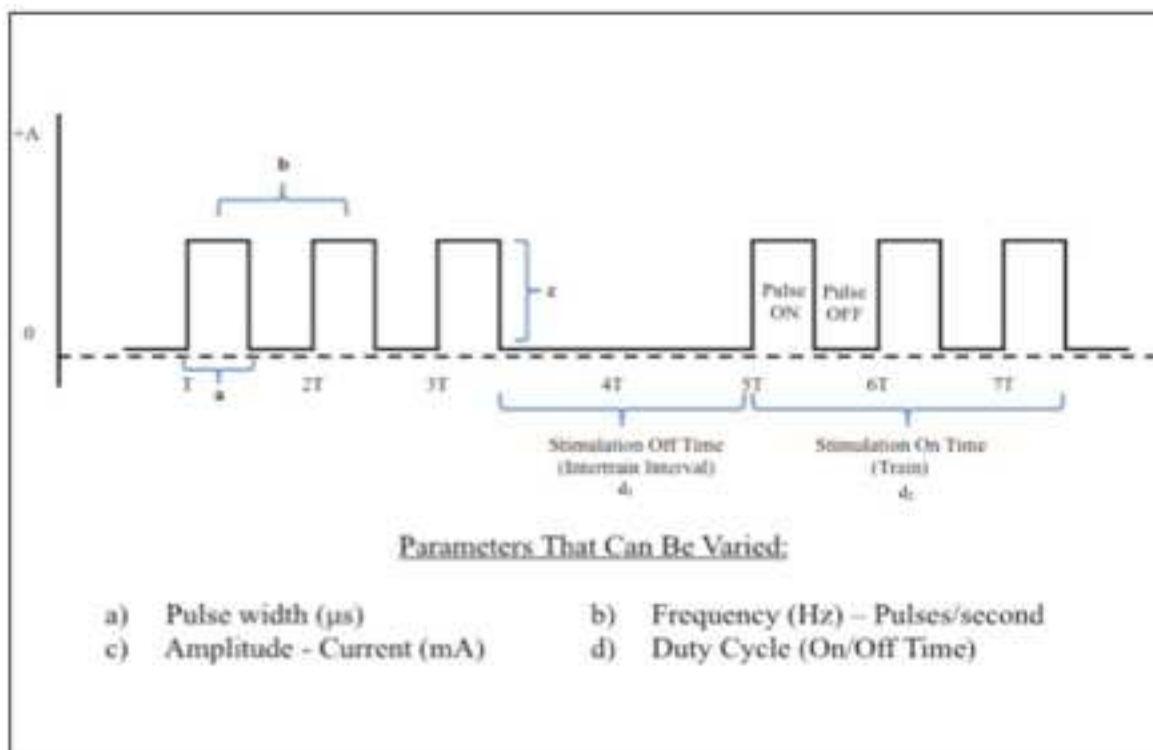
Figure 3

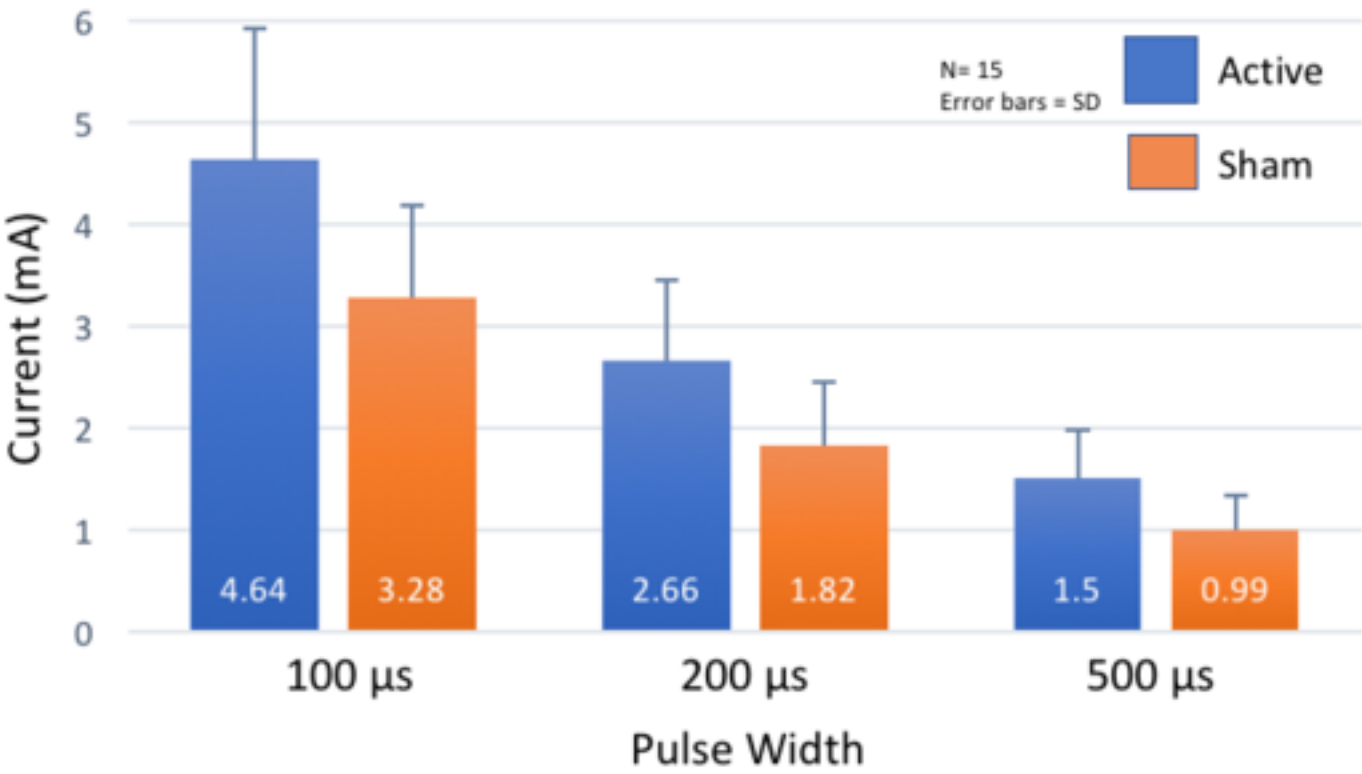



Figure 4












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stimDesigner.fig



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Stimulation Current Setting	3 mA	1.5 mA	0.75 mA	1.1 mA	0.55 mA	0.825 mA
Response	Yes	Yes	No	Yes	No	Yes

	Stim. Current ± SD (mA)		
Pulse Width	Tragus (Active)	Earlobe (Control)	Sig
100 μs	9.28 ± 2.56	6.57 ± 1.83	Yes
200 μs	5.32 ± 1.60	3.64 ± 1.26	Yes
500 μs	3 ± 0.93	1.97 ± 0.71	Yes

	PT ± SD (mA)		Sig	Stimulation Current ± SD (mA)		Sig
	Active	Control		Active	Control	
500 ms 25 HZ	0.82 ± 0.41	0.99 ± 0.39	No	1.37 ± 0.68	1.83 ± 0.73	Yes

Pain ± SD		Sig
Active	Control	
1.98 ± 0.83	2.17 ± 1.27	No

Name of Material/ Equipment	Company	Catalog Number
70% Isopropyl Alcohol Wipes	Any	N/A
Constant Current Stimulator (Triggerable)	Soterix Medical	N/A
Disposable Conductive Electrodes	Custom Built	N/A
Matlab Software w/ Stimulation GUI	MathWorks	N/A
Ten20 Conductive Paste	Weaver and Company	N/A

Comments/Description

Any alcohol preparation pads used for skin in appropriate.

Stimulator manufactured for custom use by Soterix Medical

Stimulation electrodes are custom built at the City College Neural Engineering Lab (Badran/Bikson)

MATLAB used for programing pulse pattern

Conductive paste used for administration of stimulation

n)

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Title of Article:	Methodological Description of the Laboratory Administration of Transcutaneous Auricular Vagus Nerve Stimulation (taVNS) in Humans: Technique, Targeting and Considerations
Author(s):	*Bashar W. Badran ^{1,2,3} , *Alfred B. Yu ² , Devin K.P. Adair ¹ , Georgia Mappin ³ , William H. DeVries ³ , Dorothea D. Jenkins ⁴ , Mark S. George ^{3,5,6} , Marom S. Bikson ¹

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Dear Editor,

Attached is our revised manuscript (revised title: Methodological Description of the Laboratory Administration of Transcutaneous Auricular Vagus Nerve Stimulation (taVNS) in Humans: Technique, Targeting and Considerations).

We have included a tracked changes and clean version along with responses to all reviewer comments.

Sincerely,

A handwritten signature in black ink, appearing to read "BWB", with a stylized flourish underneath.

Bashar W. Badran
e. basharwbadran@gmail.com
p. 843-792-1006

Editorial comments:

Changes to be made by the Author(s) regarding the written manuscript:

1. Please take this opportunity to thoroughly proofread the manuscript to ensure that there are no spelling or grammar issues.

OK, Completed

2. Please revise the title to reflect the method and its application.

Revised

3. Please provide an email address for each author.

Added to cover page

4. Keywords: Please provide at least 6 keywords or phrases.

Corrected.

5. Please spell out each abbreviation the first time it is used.

Corrected.

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Corrected. Thank you.

7. Please include an ethics statement before the numbered protocol steps, indicating that the protocol follows the guidelines of your institution’s human research ethics committee.

Added.

8. Please revise the protocol text to avoid the use of any personal pronouns (e.g., "we", "you", "our" etc.).

Corrected.

9. Please revise the protocol to contain only action items that direct the reader to do

something (e.g., “Do this,” “Ensure that,” etc.). The actions should be described in the imperative tense in complete sentences wherever possible. Avoid usage of phrases such as “could be,” “should be,” and “would be” throughout the Protocol. Any text that cannot be written in the imperative tense may be added as a “Note.” Please include all safety procedures and use of hoods, etc. However, notes should be used sparingly and actions should be described in the imperative tense wherever possible.

Corrected.

10. 1.1-1.4: The Protocol should contain only action items that direct the reader to do something. Please either write the text in the imperative tense as if telling someone how to do the technique (e.g., “Do this,” “Ensure that,” etc.), or move the materials and equipment information to the Materials Table.

Corrected

11. 2.1: What are the inclusion/exclusion criteria for recruiting the participants?

Added.

12. 2.3, 3.1, 3.5, 4.1, 4.2, 4.5, 5.3-5.6, 6.1, etc.: Please write the text in the imperative tense. Any text that cannot be written in the imperative tense may be added as a “Note.”

Corrected.

13. Please add more details to your protocol steps. There should be enough detail in each step to supplement the actions seen in the video so that viewers can easily replicate the protocol. Please ensure you answer the “how” question, i.e., how is the step performed? Alternatively, add references to published material specifying how to perform the protocol action. See examples below:

3.2: What is used to apply the conductive Ten20 paste?

3.3: How to verify the polarity of electrodes?

5.2: Please specify the pulse generating GUI used in this step.

Added more descriptions to the procedures.

Note- We would like to make the GUI open access. How do we do this? Can you host it as supplemental material?

14. Please upload each Figure individually to your Editorial Manager account as a .png, .tiff, .pdf, .svg, .eps, .psd, or .ai file.

Corrected.

15. Please upload each Table individually to your Editorial Manager account as an .xls or .xlsx file.

Corrected.

16. Figure 1: Please use capitalized letters A and B for panel labels.

Corrected.

17. Figure 2: Please note that the sham stimulation site is labeled as S, not B. Please revise to be consistent.

Corrected.

18. Figure 3: Please remove the picture containing commercial language (Ten20 in panel B).

Fixed.

19. Figure 6: Please include a space between numbers and their units (100 μ s, 200 μ s, 500 μ s). Please define SD in the figure legend.

Fixed.

20. Tables 1 and 3: Please include a space between numbers and their units (3 mA, 1.5 mA, 500 μ s, etc.). Should 0.825 in Table 2 be followed by mA?

Corrected.

21. Please number the tables in the sequence in which you refer to them in the manuscript text.

Corrected.

22. Please remove the embedded Table of Materials from the manuscript.

Removed.

23. References: Please do not abbreviate journal titles.

Corrected.

Reviewers' comments:

Reviewer #1:

Manuscript Summary:

This is a useful protocol that may extend the application of the tVNS, the following are my comments.

Major Concerns:

1. My main concern is the location of the stimulation. Based on a recent manuscript published in brain stimulation (2018) by Andreas M. Burger, there may be no vagus nerve distribution at tragus. Thus, before new evidence come up demonstrating that there is vagus nerve distribution at tragus, it may be not a good idea to put electrode at tragus as shown in Figure 2.

The reviewer is correct in addressing a current debate as to localization of stimulation. Our group, and others have chosen to stimulate the tragus as an active stimulation, whereas others stimulate the cymba conchae.

This was addressed in the introduction (lines 86-89)
The field is still determining optimal stimulation targets²¹, although the two most common placements are the anterior wall of the outer ear canal (tragus) and the cymba conchae. Sham stimulation may be conducted by stimulating the earlobe of the ear, an area believed to have minimal ABVN innervation (Figure 2).

We have added a second active site on Figure 2 to clarify this statement.

2. Individuals' s response to electrical stimulation varies significantly. I would suggest the authors do not use the term such as "stimulation current is determined as 200% of an individual perceptual threshold (PT)". There is no evidence suggest that 200% is better than 150% or 250%.

Great point. We agree. We have changed some of the language in the introduction and softened to language in methods.

3. It seems to the reviewer the threshold selection is too complicated to apply in clinical setting, I would suggest just increasing intensity gradually till the maximal intensity the subject can tolerate, then reduce intensity gradually till subjects feel comfortable.

The perceptual threshold finding algorithm is similar to the one used in rTMS that was developed and validated over the past 2 decades. This is considered the gold-standard for threshold finding. The authors believe this is an important

dosing parameter and respectfully will be maintaining it as a method for finding PT in the laboratory and clinical trial setting to maintain carefully controlled studies in the infancy of taVNS as a modality.

4. The Figure 4 is confusing, please just include the tVNS related part, delete all other materials that are unrelated to tVNS.

We have changed figure 4 to a more participant-focused image of a laboratory-based taVNS setup,

Reviewer #2:

Introduction

Paragraph 1: There is reference to '(C)' - unclear what this is referring to. Good mention of the financial cost of VNS, would also be worthwhile mentioning the safety profile of VNS, especially as safety and tolerability of taVNS is a key part of the paper.

Great catch. Typo deleted.

Added the safety profile of VNS to the introduction.

Paragraph 2: Mention could be made to the work by Colzato and colleagues- cognitive and social effects of taVNS.

We have added these. Thank you for the suggestion.

Paragraph 3: First sentence - should be 'is' instead of 'in'.

Fixed. Thanks.

Evidence needed for the tragus and cymba concha being the most popular areas of taVNS administration and for the earlobe having minimal ABVN innervation. Further explanation of why the tragus and cymba concha are innervated would be good.

Figure 2 is mentioned for sham stimulation, but not for stimulation on the tragus. Would help to make it clear in this paragraph that Figure 2 also shows where the tragus is. The text also mentions the cymba concha, but this is not illustrated on Figure 2. Another common target is the concha, but this is not mentioned - perhaps worth mentioning?

- R: A literature review reveals these two as the most common treatment positions for taVNS with the earlobe as the most common sham. Intervention is listed in citation 5.

Authors (and the field likely) concede that the field is operating under the auspices that the sole anatomical dissection study – Citation 5. We have made a statement to acknowledge this.

Peuker, E.T. & Filler, T.J. The nerve supply of the human auricle. *Clinical Anatomy*. **15** (1), 35-37 (2002).

We have fixed figure 2 to reflect two active sites and described in the figure legend.

May also be worth mentioning that sham can also consist of attaching electrodes to the same part of the ear which active stimulation is applied, but without passing current despite volunteers believing they are receiving active stimulation.

Interesting. Passive control added to the sham section.

Paragraph 4: Are there any reported side-effects of taVNS? - Left vs. right stimulation. Protocol

Well-detailed and clear. Can see this being useful for others. May be worth mentioning from the outset that this protocol seems to be specific for targeting the tragus. Would need to modify how the ear is targeted if administering taVNS to the concha and cymba concha.

Great point. Clarified in protocol and enhanced figure 2.

One concern is about the kit: does not look very portable or very user-friendly compared to others. Looks to be more used in experimental settings. Perhaps this is worth some consideration in the discussion.

Correct. This is a laboratory, non-portable setting that is used for parametric optimization and flexibility with opensource matlab script.

Electrode preparation and placement section:

Good detail provided regarding anode and cathode placement for the tragus. May be worthwhile being completely clear for anode and cathode placement for sham on the ear lobe.

Great point. Added.

Determination of perceptual threshold section is very good - provides a much more controlled way of determining PT.

May be worthwhile providing more detail about what to do if volunteers find taVNS too painful or uncomfortable.

Thank you. We have added a brief statement regarding this concern.

Delivering stimulation section:

Figure 5: 'd' could be more clearly labelled on the diagram.

Labeled d on the ON and OFF time. Duty cycle is hard to describe in a figure like this so we labelled d_1 and d_2 on the On/Off times.

If stimulation is typically administered at 200% of PT, reference is needed (5.3).

Fixed and noted.

After taVNS section:

Paragraph 1: References needed for first sentence to show that discomfort and side effects of taVNS are 'commonly' reported.

Added.

Representative results

It is interesting that for the same pulse width, a lower current is needed to sham site (earlobe) than tragus stimulation. From the values presented in Table 2, some of these active vs. sham differences may be statistically significant, would be worthwhile making this clear in this table.

Added. Thanks for pointing out this oversight.

Would also be of interest to see the pain rating for some of the other stimulation parameters for active and sham stimulation.

Pain ratings for other parameters can be found in the citation added to the manuscript.

Paragraph 1: There seems to be a slight mix-up with the numbering of the tables. More information about the participants would be helpful (e.g. number of males/females, age range, exclusion criteria).

Fixed.

Paragraph 2: NRS should be written out in full here (numeric rating scale). References are needed to support the first sentence. Check grammar of third sentence. More information about the participants would be helpful (e.g. number of males/females, age range, exclusion criteria).

Fixed.

Paragraph 3: Sentence 1 needs references.

Added.

Paragraph 4: Check grammar and references are needed.

Fixed typos as a result of merged edits. Thank you.

Discussion

Critical steps in the section:

Bullet point 1: What about mental health conditions? Why screen for cardiovascular disorders? Are there any particular cardiovascular disorders which may prevent participation in a taVNS study?

Inclusion/exclusion criteria included in the protocol. We had accidentally omitted this section in the initial submission.

Bullet point 2: Is there any evidence that their proposed skin preparation improves safety of taVNS? It is already a safe stimulus, so would be interesting to know how this further enhances safety.

Great question. Skin prep reduces resistance at the skin/electrode interface. This allows for less voltage needed to drive the required current through the ear.

Generally, we like to use low-current and low-voltage stimulators, to minimize the electricity needed to stimulate the nerve.

Bullet point 3: 'LOTES guidelines' should be expanded and further explained. Also, the group's published guidelines should be referenced.

Lines 47-0481 expand on LOTES and it is cited (#29).

Modifications and troubleshooting of the method section:

What about negative effects encountered by participants?

In our series of studies with over 150 individuals receiving stimulation over 4 studies, we have yet to have a negative effect encountered by participants.

Reviewer #3:**Manuscript Summary:**

The manuscript provides a relatively clear description of how to apply a new and promising technique that allows for a non-invasive stimulation of the vagus nerve

Minor Concerns:

1) It may be useful to extend the introduction to mention some of the recent studies that have successfully used taVNS to alter cognitive and social functioning in healthy participants.

We have added a brief bit in the introduction about this. Thank you for the suggestion.

2) the main differences between the authors' setup and the stimulation set-ups used by other groups should be pointed out

Noted. We have discussed different active placements with citation in the introduction and modified figure 2 to reflect other active position some groups use.

We also softened language regarding perceptual thresholds and current intensities as parameters are not yet fully established

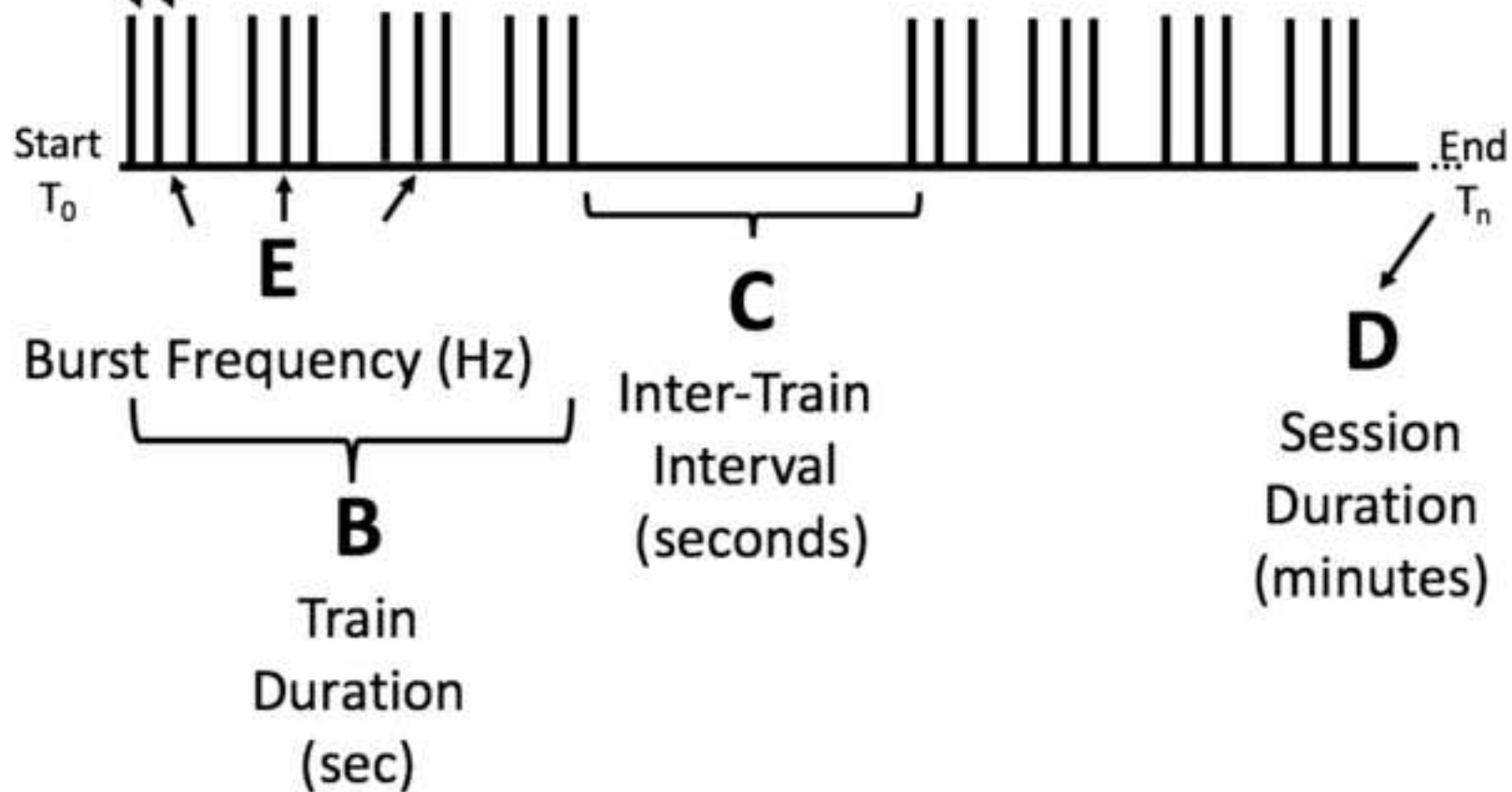
3) It would be useful to share the script used to control the stimulator

We will be including the GUI as an open-source resource with this manuscript. Thank you for the suggestion.

A & F

A) Pulse Frequency (Hz)

F) Number of Pulses in Burst (#)



READ ME

Please

Hi Researcher :)

Thank you for using stimDesigner. This software is used to trigger stimulators (electrical, magnetic, etc) by delivering individually designed waveforms custom built by you.

STEP 1: Enter parameters

This software will initiate a series of TTL pulses that drives your stimulator. In order to do this, you must input your desired waveform parameters into the GUI.

These parameters are visualized in the demo waveform figure on the right hand side of the GUI and listed below:

- 1- Frequency (Hz). This is the number of pulses delivered per second.
- 2- Train Length - This is the "ON" time of each stimulation train (Input in Seconds)
- 3- Inter-train interval - This is the "OFF" time between train. (Input in Seconds)
- 4- Total Duration- This is the length of your desired experimental stimulation period. (Input in MINUTES)

Advanced users may choose to do burst patterns. In that case, please check "burst mode" and the parameters will change to include burst parameters.

STEP 2: Inspect your waveform.

After inputting desired parameters into the GUI, click "PRODUCE FIGURE" to visualize you waveform. This will appear below the demo waveform on the right side of the GUI.

To zoom and inspect your waveform use the magnifier button on the very top tool bar (top left) of the GUI. After clicking this button, use your cursor to drag and inspect the waveform.

STEP 3: Initiate Stimulation

After verifying your waveform is correct in step 2, click "verify" to confirm and subsequently press "ok" in the dialog box. Only after these two steps are complete can you then click "START"

Key Features:

- 1) There is a running clock that starts as soon as you hit the START button. This counts up to your total session duration.

2) There is a STOP button to stop the triggers. You cannot restart after hitting start, but ensure that you have noted the time you stopped stimulation, exit the GUI, and restart.

Thank you for using stimDesigner. Go forth and stimulate safely. Please cite our manuscript listed below when you use our software. It helps us out!

Badran BW, Yu AB, Adair DKP, et al. Methodological Description of the Laboratory Administration of Transcutaneous Auricular Vagus Nerve Stimulation (taVNS) in Humans: Technique, Targeting and Considerations. Journal of Visualized Experiments (JoVE). 2018.

FAQs and Considerations

In order to properly use stimDesigner, you will need the following software and hardware:

Software:

- 1- Matlab (tested and built on v2017 but will likely work on other modern versions post-2014)
- 2- Data Acquisition Toolbox (Matlab toolbox)
- 3- Update drivers from National Instruments (These drivers are to ensure proper computer/DAQ connectivity)
- 4- stimDesigner.m & stimDesigner.fig (please keep all files in the root stimDesigner folder)

Hardware

- 1- PC Computer that can run Matlab (This has not been tested on Linux/Unix/Mac)
- 2- DAQ (our group uses the NI-DAQmx from national instruments)
- 3- Stimulator that accepts TTL pulses in for triggering (eg: Digitimer, Grass, Soterix, Magventure, etc)
- 4- USB cable (PC to DAQ) and a BNC cable (DAQ to Stimulator)

-> Please note this is a research investigational tool for IRB approved investigational studies in the research setting.

-> Test your output waveform before delivering it to any research participants. Please follow all local guidelines for safe administration of any form of brain stimulation.

Refer to all stimulation safety guidelines to assist in determination of safe stimulation parameters.

Triggering information:

Voltage of the TTL pulse is 5V rising edge (5 microsecond pulse width)

Need Help?

This GUI was designed by Devin Adair of the Neural Engineering Group at CCNY. Feel free to contact her with questions:
dadair@gradcenter.cuny.edu

