

Journal of Visualized Experiments

Bilateral assessment of the corticospinal pathways of the ankle muscles using navigated transcranial magnetic stimulation --Manuscript Draft--

Article Type:	Invited Methods Article - JoVE Produced Video
Manuscript Number:	JoVE58944R1
Full Title:	Bilateral assessment of the corticospinal pathways of the ankle muscles using navigated transcranial magnetic stimulation
Keywords:	Transcranial magnetic stimulation; corticospinal tracts; tibialis anterior; soleus; tonic voluntary activation; neurophysiology; corticomotor response; brain neuronavigation
Corresponding Author:	Charalambos C Charalambous, Ph.D New York University School of Medicine New York, NY UNITED STATES
Corresponding Author's Institution:	New York University School of Medicine
Corresponding Author E-Mail:	cccharalam@gmail.com
Order of Authors:	Charalambos C Charalambous, Ph.D Jing Nong Liang Steve A. Kautz Mark S. George Mark G. Bowden
Additional Information:	
Question	Response
Please indicate whether this article will be Standard Access or Open Access.	Standard Access (US\$2,400)
Please indicate the city, state/province, and country where this article will be filmed . Please do not use abbreviations.	Charleston, SC, USA



Department of Neurology,
New York University Langone Health
222 E 41st St, 14th Floor, 14-025G
New York, NY 10017, USA
Office: (929) 455-5116

Editorial Board
Journal of Visualized Experiments

September 25, 2018

Dear Editors,

With this letter we are resubmitting our invited manuscript, entitled “Bilateral assessment of the corticospinal pathways of the ankle muscles using **navigated** transcranial magnetic stimulation” for review (**NOTE: we have slightly revised the title**). We appreciate the thoughtful comments of the editor and reviewers; the changes we have made in response to their comments have strengthened the paper.

As before, I will serve as the corresponding author for this manuscript.

If you have questions or need any further information, please contact me.

Sincerely,

A handwritten signature in black ink that reads 'Charalambos C. Charalambous'.

Charalambos C. Charalambous, PhD
Postdoctoral Researcher
Email: charalambos.charalambous@nyulangone.org

TITLE:**Bilateral Assessment of the Corticospinal Pathways of the Ankle Muscles Using Navigated Transcranial Magnetic Stimulation****AUTHORS AND AFFILIATIONS:**

Charalambos C. Charalambous^{1,2}, Jing Nong Liang^{3,4}, Steve A. Kautz^{2,5}, Mark S. George^{5,6}, Mark G. Bowden^{2,5,7}

¹Department of Neurology, New York University School of Medicine, New York, NY, USA

²Department of Health Sciences and Research, Medical University of South Carolina, Charleston, SC, USA

³Department of Physical Therapy, University of Nevada Las Vegas, Las Vegas, NV, USA

⁴Department of Health Professions, Medical University of South Carolina, Charleston, SC, USA

⁵Ralph H. Johnson VA Medical Center, Charleston, SC, USA

⁶Department of Psychiatry, Medical University of South Carolina, Charleston, SC, USA

⁷Division of Physical Therapy, Medical University of South Carolina, Charleston, SC, USA

Corresponding Author:

Charalambos C. Charalambous

charalambos.charalambous@nyulangone.org

Tel: (929)-455-5116

Email Addresses of Co-authors:

Jing Nong Liang (jingnong.liang@unlv.edu)

Steve A. Kautz (kautz@musc.edu)

Mark S. George (georgem@musc.edu)

Mark G. Bowden (bowdenm@musc.edu)

KEYWORDS:

Transcranial magnetic stimulation, corticospinal tracts, tibialis anterior, soleus, tonic voluntary activation, neurophysiology, corticomotor response, brain neuronavigation

SUMMARY:

The present protocol describes the simultaneous, bilateral assessment of the corticomotor response of the tibialis anterior and soleus during rest and tonic voluntary activation using a single pulse transcranial magnetic stimulation and neuronavigation system.

ABSTRACT:

Distal leg muscles receive neural input from motor cortical areas via the corticospinal tract, which is the main motor descending pathway in humans and can be assessed using transcranial magnetic stimulation (TMS). Given the role of distal leg muscles in upright postural and dynamic tasks, such as walking, a growing research interest in the assessment and modulation of the corticospinal tracts relative to the function of these muscles has emerged in the last decade. However, methodological parameters used in previous work have varied across studies making the interpretation of results from cross-sectional and longitudinal studies less robust. Therefore,

use of a standardized TMS protocol specific to the assessment of leg muscles' corticomotor response (CMR) will allow for direct comparison of results across studies and cohorts. The objective of this paper is to present a protocol that provides the flexibility to simultaneously assess the bilateral CMR of two main ankle antagonistic muscles, the tibialis anterior and soleus, using single pulse TMS with a neuronavigation system. The present protocol is applicable while the examined muscle is either fully relaxed or isometrically contracted at a defined percentage of maximum isometric voluntary contraction. Using each subject's structural MRI with the neuronavigation system ensures accurate and precise positioning of the coil over the leg cortical representations during assessment. Given the inconsistency in CMR derived measures, this protocol also describes a standardized calculation of these measures using automated algorithms. This protocol is not conducted during upright postural or dynamic tasks. However, this protocol can be used to assess bilaterally any pair of leg muscles, either antagonistic or synergistic, in both neurologically intact and impaired subjects.

INTRODUCTION:

The tibialis anterior (TA) and soleus (SOL) are ankle antagonistic muscles located in the anterior and posterior compartment of the lower leg, respectively. Both muscles are uniarticular, while the main function of TA and SOL is to dorsiflex and plantarflex the talocrural joint, respectively¹. Furthermore, TA is more functional for long muscle excursions and less important for force production, whereas SOL is an antigravity muscle designed to generate high force with small excursion of the muscle². Both muscles are especially relevant during upright postural and dynamic tasks (*e.g.*, walking)^{3,4}. Regarding neural control, the motor neuron pools of both muscles receive neural drive from the brain via the motor descending pathways^{5,6}, in addition to varying degrees of sensory drive.

The main motor descending pathway is the corticospinal tract, which originates from the primary and supplementary motor areas and terminates in the spinal motor-neuron pools^{7,8}. In humans, the functional state of this tract (corticomotor response – CMR) can be feasibly assessed using transcranial magnetic stimulation (TMS), a non-invasive brain stimulation tool^{9,10}. Since the introduction of TMS and given their functional significance during upright postural task and walking, CMR of TA and SOL have been assessed in various cohorts and tasks¹¹⁻³².

In contrast to the assessment of CMR in upper-extremity muscles³³, no universal TMS protocol has been established for the assessment of CMR in lower-extremity muscles. Due to the lack of an established protocol and the large methodological variability across the previous studies (*e.g.*, type of coil, use of neuronavigation, level of tonic activation, testing side and muscle, use and calculation of CMR measures, *etc.*), the interpretation of results across studies and cohorts can be cumbersome, complicated, and inaccurate. As the measures are functionally relevant in various motor tasks, an established TMS protocol specific to lower extremity CMR assessment will allow motor neuroscientists and rehabilitation scientists to systematically assess the CMR in these muscles across sessions and various cohorts.

Therefore, the objective of this protocol is to describe the bilateral assessment of TA and SOL

CMR using single pulse TMS and neuronavigation system. In contrast to previous work, this protocol aims to maximize rigor of the experimental procedures, data acquisition, and data analysis by employing methodological factors that optimize the validity and duration of the experiment, and standardize the CMR assessment of these two lower extremity muscles. Given that the CMR of a muscle depends on whether the muscle is fully relaxed or is partially activated, this protocol describes how the TA and SOL CMR can be assessed during rest and tonic voluntary activation (TVA). The following sections will thoroughly describe the present protocol. Finally, representative data will be presented and discussed. The protocol described here is derived from that in Charalambous *et al.* 2018³².

PROTOCOL:

All experimental procedures presented in this protocol have been approved by the local Institutional Review Board and are in accordance with the Declaration of Helsinki.

1. Consent Process and Safety Questionnaires

1.1. Prior to any experiment, explain to each subject the aim of the study, the main experimental procedures, and any potential risk factors associated with participating in the study. After answering any questions or concerns that subjects may have, ask subjects to acknowledge the consent process and sign the informed consent form.

1.2. Administer MRI³⁴ and TMS³⁵ safety-screening questionnaires to ensure subjects' safety and qualification for both MRI and TMS testing. Exclude all subjects who don't meet all safety criteria from both MRI and TMS assessments.

2. MRI and Preparation of the Neuronavigation System

2.1. Administer the MRI assessment prior to TMS assessment³². Have subjects lie in a supine position with a cushion placed under their knees to ensure a comfortable posture. Instruct subjects to keep still in the scanner.

2.1.1. Provide ear protection to the subjects to attenuate the loud noise of the scanner. Preferentially use ear plugs over ear muffs due to the use of bilateral supratragic notch for subject-image registration in the neuronavigation system (see 5.2).

2.1.2. Obtain high-resolution T-1 weighted anatomical brain images (minimum requirements: 1 mm slice thickness and full brain and cerebellar coverage), either as NIFTI or DICOM files. Ensure that nose is fully included in the images due to the use of subject's tip of the nose for subject-image registration in the neuronavigation system (see 5.2).

2.2. Upload MRI files into a neuronavigation system. Co-register manually each subject's MRI to anterior and posterior commissures, so the subject's MRI can be mapped using the Montreal Neurological Institute atlas.

2.2.1. Reconstruct the skin and full curvilinear brain model by adjusting the bounding box around the skull and brain tissue, respectively. Identify four anatomical landmarks (tip of the nose, nasion - bridge of the nose, and supratragic notch of the right and left ear) using the skin model (see **Figure 1A**).

2.2.2. Place a rectangular grid over leg motor cortical area at each hemisphere (see **Figure 1B**). Position the centered row of the grid at the center and over the gyrus of the leg motor cortical area where the corticospinal tracts that innervate leg motor pools originate³⁶. Position the medial column of the grid parallel and adjacent to the medial wall of the ipsilateral hemisphere.

2.2.3. Use a cortex based approach in which error in orientation has a negligible effect on the stimulation site³⁷ instead of using a scalp based target approach in which any error in orientation can alter the stimulation site. Use this grid to find the hot spot. For motor mapping, use larger grids either by adding more spots and/or increasing the distance between spots (*e.g.*, 10 mm).

3. Subject Preparation and Placement

3.1. Measure the electrophysiological responses by single pulse TMS using a total of 4 surface EMG electrodes. For the preparation and placement of the electrodes, use published guidelines^{38,39} and complete placement while the subject is in a standing position.

3.1.1. Prepare the area over which each electrode would be placed by shaving and lightly exfoliating any dead skin cells and oils using alcohol swabs.

CAUTION: For subjects on blood thinners (*e.g.* people post-stroke), use caution during skin preparation due to high risk of bleeding.

3.1.2. Attach electrodes bilaterally on TA. While in the standing position, ask subjects to lift their toes upwards and then place the electrode at the upper third of the line between the head of the fibula and medial malleolus (*i.e.*, muscle belly immediately lateral to the tibial crest).

3.1.3. Attach electrodes bilaterally on the lateral SOL. While in the standing position, ask the subject to perform heel raise and then place the electrode at the lower third of the line between the lateral femoral condyle and lateral malleolus.

3.1.4. Attach the ground reference passive electrode either on the patella or lateral malleolus. Depending on the EMG acquisition unit, place the ground electrodes either bilaterally or unilaterally.

3.2. Test the electrodes' placement and quality of the signal.

3.2.1. Test the electrodes' placement (*e.g.*, for clear visually detectable EMG bursts) by asking the subject to either dorsiflex or plantarflex the ankle in an upright posture while displaying the

raw EMG signal of all muscles tested on a computer screen. In the case of a misplaced electrode, remove and replace it until there is clear visually detectable EMG bursts with minimal background noise. An adequate signal to noise ratio is critical in detecting a motor response ($> 50 \mu\text{V}$).

3.2.2. Test the quality of the signal (*e.g.*, for baseline noise) by discharging the TMS units for a few times while the TMS coil is held away from the seated subject and with the muscles at rest. Check that the baseline signal for each EMG channel is close to zero (*i.e.*, the peak-to-peak amplitude should be less than $50 \mu\text{V}$ and there is no baseline noise, such as 50 or 60 Hz power line hum). If baseline noise is present in a channel, remove the corresponding electrode and repeat the skin preparation procedures. If the noise is still present (*i.e.*, peak-to-peak amplitude $> 50 \mu\text{V}$), adjust the reference electrode's position and replace the electrolyte gel.

3.3. Secure all electrodes using light foam pre-wrap tape. Periodically throughout the experiment, check to ensure that electrodes are securely attached and that the signal has good quality.

3.4. Seat the subject in a chair. To ensure consistent feet placement across subjects, secure both feet in walking boots (*i.e.*, ankle foot orthosis) that allow the ankle ROM to be adjusted to a specific position and provide resistance during TVA testing. Adjust both hip and knee angles to avoid subject discomfort. Instruct the subject to keep still throughout the experiment. Use a forehead rest attached to the chair to keep subjects still during TMS application, if available.

4. TVA Testing

4.1. Determine bilaterally the maximum voluntary isometric contraction (MVIC) of each muscle. For each motion (*i.e.*, dorsiflexion and plantarflexion), instruct subjects to maximally contract the contralateral examined muscle (*e.g.*, right TA) 4 times (~ 5 s contractions separated by 60 s of rest) while subject is seated in the posture described above.

4.2. Calculate the maximum muscle activity value during each MVIC (*i.e.*, the average within a 100 ms window centered around the maximum rectified and smoothed EMG) of the last three trials, the average of the three values, and the 15% and 5% of each muscle's average MVIC.

CAUTION: A larger % MVIC can be used, but it may not be feasible in clinical cohorts (*e.g.*, people post-stroke).

5. Registration in Neuronavigation System

5.1. Place the subject tracker, either a headband or glasses, with reflective markers on the subject's head at the opposite side from the stimulated hemisphere so the tracker does not obstruct positioning of the coil during the stimulation of each grid spot.

CAUTION: In the case that a headband is used, ensure that it is snug on subject's head, yet not overly tight because it may cause a headache after an extended period of time.

5.2. Verify the proper position of the motion capture camera by placing the subject tracker, the pointer, and the coil tracker in its capture volume space. Perform the subject-image registration by placing the tip of the pointer on the 4 anatomical landmarks (see **Figure 1A**).

5.3. Once all anatomical landmarks are sampled, verify whether registration occurred accurately by placing the tip of the pointer on several spots over the subject's skull (*i.e.*, validation stage). If the distance from the tip of the pointer to the reconstructed skin is less than 3 mm, proceed to TMS experiment; otherwise, repeat the subject-image registration until the desired error values are obtained. During the experiment, repeat registration if the subject tracker is accidentally moved.

6. TMS

6.1. Use the same methodological parameters during rest and TVA.

6.1.1. Apply single pulse stimuli on the optimal site (*i.e.*, hot spot; see next paragraph for further details) of the examined muscle. Apply reach stimulus randomly every 5-10 s to avoid stimulus anticipation and to minimize the carry-over effects of the previous pulse to the subsequent one⁴⁰.

6.1.2. In case that two TMS units are simultaneously used, set the units at either the standard or simultaneous mode⁴¹. The standard mode applies a weaker pulse than a single unit, whereas the simultaneous mode applies a stronger pulse than a single unit. The use of either one could be based on the needs of the protocol and the total number of stimuli.

6.1.3. Use a double cone coil to induce a posteroanterior intracranial current. If necessary, use the neuronavigation system to control the coil manually and correct its position in relation to the desired stimulated spot prior to each stimulus.

6.1.4. Across sessions and subjects, randomize the order of the examined muscle and hemisphere. Always administer the TVA condition after the rest condition to avoid any interference with testing at rest (*e.g.*, fatigue of the descending pathways due to TVA testing).

6.2. Determine bilaterally the hot spot of both muscles.

6.2.1. Find the suprathreshold intensity, which will be used during hot spot hunting, by applying a single stimulus over the centered spot next to the interhemispheric fissure (see blue and red squares in **Figure 1B**). Use this spot because it is located at the locus of the leg motor area^{36,42}.

6.2.2. Start at low intensity (*e.g.*, 30% maximum stimulator output; MSO) and gradually increase the TMS intensity by 5% increments, until reaching the intensity that elicits a motor evoked potential (MEP) with a peak-to-peak amplitude greater than 50 μ V in all contralateral examined muscles for 3 consecutive stimuli.

6.2.3. Determine immediately after each stimulus whether a MEP has been elicited based on both the raw waveforms and peak-to-peak amplitudes (search window: 20-60 ms post-TMS onset) of all examined muscles.

6.2.4. Apply one TMS pulse on each spot of the grid (total 36 stimuli). After the completion of the hot spot protocol, transfer the amplitude and latency values of each spot for all contralateral muscles in a spreadsheet and sort amplitude from high to low and latency from low to high. Identify the hot spot of contralateral TA and SOL as the location in the grid with the largest amplitude and the shortest latency⁴³.

CAUTION: If the largest amplitude and shortest latency are not at the same spot, define the hot spot using the largest amplitude.

6.3. Determine bilaterally each muscle's resting motor threshold (RMT).

6.3.1. Select the grid spot in the neuronavigation system that corresponds to the examined muscle's hot spot.

6.3.2. Use an adaptive threshold-hunting method for RMT determination of the examined muscles⁴⁴. Set the initial intensity and step size at 45 and 6% MSO, respectively³². Run the RMT hunting twice for each muscle and use the average for the subsequent CMR assessment.

6.4. Assess bilaterally TA and SOL CMR during rest.

6.4.1. Select the grid spot in the neuronavigation system that corresponds to the examined muscle's hot spot. Apply 10 single TMS pulses at 1.2 RMT of the examined muscle.

6.4.2. Prior to each stimulus, instruct the subject to stay still and relax the examined muscles bilaterally and monitor the activity of all muscles using a real time visual feedback displaying on a computer screen. In case any muscle is active before or after TMS, discard that trial and apply an additional single pulse. Repeat until 10 waveforms for each contralateral examined muscle at rest have been collected.

6.5. Assess bilaterally the TA and SOL CMR during TVA.

6.5.1. Select the grid spot in the neuronavigation system that corresponds to the examined muscle's hot spot.

6.5.2. Ask subjects to contract the examined muscle at $15\% \pm 5\%$ MVIC and apply 10 single TMS pulses at 1.2 RMT. Instruct subjects to keep the smoothed moving line (root mean square amplitude of 0.165 s) of the examined muscle, either TA or SOL, within the two horizontal cursors (MVIC range: $15\% \pm 5\%$) and sustain that contraction at that level for few seconds.

6.5.3. When TA is the examined muscle, ask subjects to pull slightly up against the bootstraps on

their contralateral leg (*i.e.*, the leg with the examined muscle contralateral to stimulated hemisphere). When SOL is the examined muscle, ask subjects to push slightly down against the boot on the contralateral leg.

6.5.4. Monitor the muscle activity of the active examined muscle and the remaining resting muscles using a real time visual feedback display on a computer screen. Discard that stimulus and apply an additional single pulse again in case the examined muscle's activity is either below or above the predetermined range or any other muscle is activated. Collect 10 trials while the examined muscle is activated at the predetermined range.

7. Data Analysis

7.1. For all CMR measures except RMT, calculate the value of each measure from each MEP sweep (the total duration should be at least 500 ms with minimum 100 ms pre-stimulus duration) for all muscles and then average these 10 values to get a single value (*i.e.*, mean)³². Amplitude and cortical silent period (CSP) are proxy excitability measures of CMR, whereas latency is a proxy connectivity measure of CMR. For both rest and TVA, normalize latency relative to each subject's height, as latency is influenced by distance to the examined muscle⁴⁵.

7.2. Calculate MEP amplitude and latency during rest.

7.2.1. Calculate amplitude (μV) from the raw EMG as the largest difference between positive and negative peaks (*i.e.*, peak-to-peak) of the MEP. For these two particular muscles, search for peak-to-peak within a time window of 20-60 ms after TMS onset.

CAUTION: Though the MEP search window of 20-60 ms may work for neurologically intact subjects and people post-stroke, wider MEP search windows (*e.g.*, 20-75 ms) might be required for other neurological populations (*e.g.*, multiple sclerosis).

7.2.2. Calculate latency (ms) from the rectified EMG as the time between TMS onset and MEP onset (*i.e.*, the time when a rectified EMG trace first crosses a predetermined threshold - mean plus three standard deviations of the 100 ms pre-stimulus EMG)^{32,46}.

7.3. Calculate MEP amplitude, latency, and CSP during TVA.

7.3.1. Calculate amplitude (μV) from the raw EMG as the largest difference between positive and negative peaks (*i.e.*, peak-to-peak) of the MEP. For these two particular muscles, search for peak-to-peak within a time window of 20-60 ms after TMS onset.

7.3.2. Calculate latency (ms) from the rectified EMG as the time between TMS onset and MEP onset.

7.3.2.1. Calculate the MEP onset differently in TVA than in rest. Calculate MEP onset and offset by finding the two time points that the rectified EMG trace crosses the predetermined threshold

set to the level of 100 ms pre-stimulus mean EMG. Then, find the peaks that are at least greater than the mean of the pre-stimulus EMG plus three standard deviations and between those two time points. Then, search from the first peak to 50 data points (sampling rate of 5000 Hz) before that peak for the time that the rectified EMG trace first crosses the threshold of the mean pre-stimulus EMG. Define that time as the MEP onset³².

7.3.3. Calculate CSP (ms) from the rectified EMG as the time between the MEP offset and EMG resumption (*i.e.*, absolute CSP: exclusion of MEP duration)⁴⁷. Search from the last peak to 200 data points (sampling rate of 5000 Hz) after that peak for the time that the rectified EMG trace last crossed the threshold of the mean pre-stimulus EMG; define that time as the MEP offset. Then, calculate the resumption of baseline EMG, which is the time that the rectified EMG trace last crosses 25% of the mean pre-stimulus EMG³².

REPRESENTATIVE RESULTS:

Figures 2-4 present data from a representative neurologically intact 31 year old male with height and weight of 178 cm and 83 kg, respectively.

Figure 2 presents the bilateral hot spots and RMT of each ankle muscle. Using the spot located on the center of the leg area in each hemisphere (see squares in **Figure 1B**), the intensity of 45% MSO was bilaterally used for the hot spot hunting. The hot spot location for each muscle differed between hemispheres, yet as anticipated all four hot spots were located in the leg motor cortical areas. This finding indicates that TA and SOL may not share the same hot spot; therefore, CMR of each muscle should be assessed using each muscle's hot spot as opposed to using the same hot spot for both muscles. Bilateral RMT was determined for each muscle, using an adaptive threshold-hunting method. The number of stimuli applied for RMT determination ranged from 6 to 22 stimuli. The difference between the two RMT values of each muscle ranged from 1% to 3% MSO. Combining these results suggests that using an adaptive threshold-hunting method can be an efficient approach to determine the RMT of an ankle muscle with low variability. Furthermore, all RMTs were lower than the intensity used for hot spot hunting (dashed line in **Figure 2**). This finding indicates that using the spot located on the leg motor area (see squares in **Figure 1B**) to determine a "true" suprathreshold intensity is feasible.

Figure 3 presents the bilateral responses of TA and SOL when the hot spot of each muscle was stimulated during rest. For all bilateral stimulated hot spots, contralateral MEPs were elicited in both TA and SOL. However, the responses and latencies were always larger and shorter in TA than in SOL, respectively, regardless which muscle's hot spot was stimulated. Ipsilateral responses were present mainly in TA and when the stimulated hot spot was proximal to interhemispheric fissure (see **Figure: 2A – TA hot spot, 2B – both muscles hot spot**). Conversely, ipsilateral responses were absent in both muscles when the stimulated side was further lateral from the interhemispheric fissure (>10 mm) (see **Figure 2A – SOL hot spot**).

Figure 4 presents the bilateral responses of TA and SOL when the hot spot of each muscle was stimulated during TVA. As in rest, contralateral MEPs were elicited in both TA and SOL for all

bilateral stimulated sites during a $15\% \pm 5\%$ MVIC. Only the examined muscle was activated; therefore, the remaining three muscles were at rest. CSP was present only in the examined activated muscle, both TA and SOL. As in rest, TMS over right TA and left SOL hot spots also elicited ipsilateral responses; those responses were present only in the ipsilateral TA (see **Figure 4A,D**). Conversely, TMS over the right SOL and left TA hot spots elicited only contralateral MEPs. Interestingly, late responses in contralateral SOL was present only when TA was activated; those responses were present bilaterally, occurred between 80-100 ms post-TMS, and had larger amplitudes than the MEPs (see \dagger in **Figure 4A,C**). These late responses with range 70-100 ms post-TMS have previously been reported to be prevalent in SOL only with TA TVA (0-40% MVIC)^{48,49}.

Both resting and TVA conditions were similar in that ipsilateral responses were elicited when certain hot spots were stimulated. The presence of ipsilateral responses could potentially be the result of stimulation of an oligosynaptic pathway (*e.g.*, cortico-reticulo-spinal tract) or the spread of the pulse's current. An approach to distinguish between the two possible causes is to calculate the difference in latency between the contralateral and ipsilateral responses. Previous TMS studies have speculated that an ipsilateral response of > 3 ms delay relative to the contralateral response is an ipsilateral MEP (iMEP), and the potential pathway could be the cortico-reticulo-spinal tract (*i.e.*, oligosynaptic pathway)⁵⁰⁻⁵⁴. Conversely, any ipsilateral response with a shorter delay might be the result of the pulse's current; therefore, such a response may not be an iMEP. During rest, ipsilateral responses had similar latencies as contralateral responses (see **Figure 3A, C and D**). Thus, these responses were most likely not iMEPs, but were likely elicited due to the spread of the pulse's current applied adjacent to the interhemispheric fissure. When the right TA and the left SOL were activated during TVA, ipsilateral responses were only elicited in the TA and were delayed by > 3 ms compared to the contralateral response (see **Figure 4A,D**). These responses might be iMEPs, which may indicate stimulation of the cortico-reticulo-spinal tract. In summary, ipsilateral responses are common when the leg motor area is stimulated⁵⁵; therefore, caution should be taken when these responses are interpreted as iMEPs.

FIGURE AND TABLE LEGENDS:

Figure 1: Reconstructed Skin and Curvilinear Brain Models. (A) A skin model with four anatomical landmarks (tip of the nose, nasion, and supratragic notch of the right and left ear) is used to calculate the subject-image registration during the assessment by placing the tip of a pointer on each landmark. (B) A 4 x 9 rectangular grid placed bilaterally over the leg motor cortical area. Squares denote the spots used to determine the suprathreshold intensity used for the hot spot hunting.

Figure 2: Bilateral TA and SOL Hot Spots and RMT. In both hemispheres, the star symbol denotes the hot spot of each muscle. Bar plots present the mean RMT of two assessments (open white circle) for each muscle, while the values below each circle denote the number of stimuli applied to determine the RMT using an adaptive threshold-hunting method. The dashed line indicates the intensity used for the hot spot hunting (45% MSO). (A) Hot spots and RMTs of the right/contralateral TA and SOL while TMS was applied over the left hemisphere. TA hot spot was

over the leg motor area and proximal to the interhemispheric fissure whereas SOL hot spot was 10 mm lateral to TA hot spot. The number of stimuli used to determine TA and SOL RMT ranged 6-21 and 9-11, respectively. **(B)** Hot spots and RMTs of the left/contralateral TA and SOL while TMS was applied over the right hemisphere. As in the left hemisphere, TA hot spot was over the leg motor area and proximal to the interhemispheric fissure. SOL hot spot was 7.1 mm posterior-lateral to TA hot spot. The number of stimuli used to determine TA and SOL RMT were in the ranges 10-22 and 10-11, respectively.

Figure 3: Bilateral TA and SOL CMR Assessment - Rest. For the stimulation of each hot spot, the EMG of the bilateral resting TA and SOL were collected while the average waveform of each muscle is presented (total duration 500 ms; 100 ms pre-TMS). The √ and X symbols denote that MEP was either present ($>50 \mu\text{V}$) or absent ($\leq 50 \mu\text{V}$), respectively. In case of MEP's presence, the values of the peak-to-peak amplitude (μV) and latency (ms) are presented. **(A)** Stimulation of the right/contralateral TA hot spot on left hemisphere. MEPs were elicited in both right/contralateral ankle muscles, with right TA having larger amplitude and shorter latency than right SOL. Given that the stimulated hot spot is located by the interhemispheric fissure and proximal to the leg motor area on left hemisphere, MEP on the left/ipsilateral ankle muscles was also elicited (only TA). **(B)** Stimulation of the right/contralateral SOL hot spot on left hemisphere. MEPs were elicited only on the right/contralateral ankle muscles; however TA had larger MEP amplitude and shorter latency than SOL. **(C)** Stimulation of the left/contralateral TA's hot spot on right hemisphere. MEPs were elicited in both left/contralateral and right/ipsilateral ankle muscles with both TA having larger MEP amplitudes and shorter latencies than both SOL. This bilateral MEP elicitation is mainly due to the location of the stimulated hot spot and suprathreshold intensity. **(D)** Stimulation of the left/contralateral SOL hot spot on right hemisphere. MEPs were elicited in left/contralateral ankle muscles and right/ipsilateral TA.

Figure 4. Bilateral TA and SOL CMR Assessment - TVA. For the stimulation of each hot spot, the EMG of the bilateral TA and SOL were collected while the examined contralateral muscle was activated at $15 \pm 5\%$ MVIC. The average waveform of each muscle is presented (total duration 500 ms; 100 ms pre-TMS). The √ and X symbols denote that MEP was either present ($>50 \mu\text{V}$) or absent ($\leq 50 \mu\text{V}$), respectively. In case of MEP's presence, the values of the peak-to-peak amplitude (μV), latency (ms), and CSP (ms) are presented. **(A)** Stimulation of the right/contralateral TA hot spot on left hemisphere. Right TA MEP was followed by CSP. MEP was elicited in contralateral/right SOL in which a late response (†) was also elicited (amplitude: $563 \mu\text{V}$; latency: 82.8 ms). MEP was also elicited in left/ipsilateral TA, whose latency is delayed by 5.2 ms compared to the right/contralateral TA's latency. **(B)** Stimulation of the right/contralateral SOL hot spot on left hemisphere. Right/contralateral SOL MEP was followed by CSP, and MEP was also elicited in contralateral/right TA. No left/ipsilateral MEPs were elicited. **(C)** Stimulation of the left/contralateral TA hot spot on right hemisphere. Left TA MEP was followed by CSP. MEP was elicited in left/contralateral SOL in which a late response (†) was also elicited (amplitude: $465 \mu\text{V}$; latency: 96.3 ms). No MEPs were elicited in right/ipsilateral muscles. **(D)** Stimulation of the left/contralateral SOL hot spot on right hemisphere. Left SOL MEP was followed by CSP. MEPs were elicited in left/contralateral SOL and right/ipsilateral TA, whose latency is delayed by 4.7 ms compared to the left/contralateral TA's latency. No MEP was elicited in right/ipsilateral SOL.

DISCUSSION:

Given the emerging interest in how the motor cortex contributes to the motor control of leg muscles during dynamic tasks in various cohorts, a standardized TMS protocol that describes the thorough assessment of these muscles is needed. Therefore, for the first time, the present protocol provides standardized methodological procedures on bilateral assessment of two ankle antagonistic muscles, SOL and TA, during two muscle states (rest and TVA) using a single pulse TMS with neuronavigation.

The findings described in the representative results section points out several critical steps which should be considered. First, CMR assessment of these muscles, as well as other leg muscles, should be conducted using a neuronavigation system in which each subject's MRI should be used and each muscle's hot spot should be determined. Neuronavigation can guide precise TMS stimulation over the target motor area, and when the subject's MRI is used, the target motor area can be stimulated accurately^{56,57}. Previous work examined the effects of using neuronavigation during a TMS assessment of upper extremity muscles⁵⁸⁻⁶⁰; findings from those studies were mixed. Yet, no study examined this effect for a lower extremity muscle. Given the location of the motor cortical areas of TA and SOL (*i.e.*, adjacent to the interhemispheric fissure at approximately 3-4 cm below the scalp surface)^{36,42,61}, hunting for the "true" hot spot of each muscle using a grid placed on each subject's anatomy increases the probability of feasibly eliciting an MEP in either muscle, especially in SOL. Using the same protocol presented here, we have recently shown that MEPs were successfully elicited in both TA and SOL in nearly all subjects (N = 21)³². The second critical step is the bilateral assessment of each muscle. In contrast to upper extremity motor areas, the two leg motor areas are adjacent to each other, and when a pulse is applied over one area the opposite area might be stimulated due to current spread. Therefore, any ipsilateral response in either muscle may indicate either the presence of an iMEP (a potential proxy of cortico-reticulo-spinal pathway)⁵⁰ or just a direct stimulation of the opposite leg motor area. In the past, ipsilateral TA responses were reported, yet the stimulated site was based on anatomical landmark (10 and 15 mm posterior and lateral to vertex)⁶². Using this protocol, the hot spot of each muscle can be determined separately, and depending on the hot spot's location either contralateral or bilateral responses can be elicited (see **Figure 3** and **Figure 4**). Whether the bilateral response is a result of multiple descending pathways or just stimulation of a single pathway requires further investigation.

The present protocol can be modified depending on the research design. While single pulse TMS is used in this protocol, paired pulse (test pulse is preceded by conditioning pulse)^{63,64} can also be used to assess the intracortical networks of these two ankle muscles. Similarly, after hot spot and RMT determination of each muscle, bilateral input-output curves of each muscle can be acquired to assess the relationship between TMS intensity (input) and the MEP amplitude (output). To assess the CMR of each muscle, 10 stimuli are applied on each hot spot during rest and TVA, yet recent reports have suggested that more than 10 stimuli should be used to assess reliably the CMR of a muscle^{65,66}. Similarly, more than one stimulus per spot can be applied during the hot spot hunting (*e.g.*, 2-5 stimuli/spot) compared to a single stimulus per spot used in this

protocol. By applying more than one stimulus per spot, the hot spot of each muscle might be more reliably determined. Recent study suggested that as few as two stimuli per spot might be sufficient for hot spot determination⁶⁷. Furthermore, compared to the most widely used threshold hunting method, the relative frequency method⁶⁸, which is based on the Rossini-Rothwell criterion^{69,70}, the adaptive threshold-hunting method is used in the present protocol. Though the adaptive threshold-hunting method is more efficient (*i.e.*, fewer stimuli are required to determine RMT) than the relative frequency method, both methods share similar precision⁷¹. It is important to remember that all aforementioned modifications increases the total count of stimuli applied. Lastly, the current protocol used the criterion of less than 50 μ V peak-to-peak amplitude to assess for baseline noise and for the “true” resting state. Discarding any EMG signal greater than 10 μ V (root mean square calculated over 100 ms) is an alternative approach.

This protocol has few methodological considerations. First, the assessment of these two muscles is in a seated position, either during rest or TVA. As previously mentioned, both TA and SOL are crucially important during upright postural tasks and walking. Although previous studies have examined TA and SOL CMR during upright postural task^{14,72-76} and walking^{20,22,77-79}, the assessment was only unilateral, and TMS was not guided by neuronavigation. Therefore, even if the present protocol is not used during these tasks, it can still provide a non-invasive window about the cortical drive of these two ankle muscles. Second, the active motor threshold (AMT) was not determined because there is not a well established methodology for that measure. Given that AMT is correlated with and is lower than RMT ($\sim 82\%$)⁸⁰, MEP can be elicited during TVA even when using a suprathreshold intensity of RMT. Third, use of structural MRI of each subject with the neuronavigation system may not be feasible in all settings due to high cost of obtaining MRI and the neuronavigation system. However, certain neuronavigation systems including the one used in this protocol, can be used without subject’s MRI; but an average MRI is used. In this case, the coil can be still precisely positioned over the stimulated site.

While previous work has examined TA and SOL CMR during various tasks in different cohorts, no study used a standardized protocol that examined these two muscles bilaterally using neuronavigation with each subject’s MRI. Use of each subject’s structural MRI combined with a neuronavigation system promotes the accuracy and precision of stimulation of the motor cortical representations of both muscles. This is crucially important for the leg motor cortical areas. Also, given that the CMR of a muscle depends on whether the muscle is fully relaxed or is partially activated, this protocol describes how the TA and SOL CMR can be assessed during rest and TVA. Additionally, each hemisphere is stimulated while the bilateral CMR of each muscle is simultaneously assessed. Furthermore, rather than using the same hot spot for assessing a single muscle’s CMR, each muscle’s hot spot is determined using a standardized grid, which was laid over the leg cortical representation, and is defined as the spot with the largest amplitude and shortest latency⁴³. Though the relative frequency method is widely used to measure the motor threshold of a muscle⁶⁸, this protocol uses an adaptive threshold-hunting method to reduce the experimental duration and total number of stimuli applied per session⁴⁴. Finally, to reduce the duration of data analysis and to standardize the calculation of CMR measures, an automated data analysis methodology is used.

Future studies can use this protocol to further elucidate the cortical control of TA and SOL in both neurologically intact and impaired cohorts. One such application of the present protocol is the mapping of these two muscles. Though few studies examined the motor cortical area of TA⁸¹⁻⁸⁴, only one study reported the motor cortical area of SOL from a single patient with focal cortical dysplasia⁸⁵. A common characteristic that all these studies share is the use of the same neuronavigated TMS system, which is different from the system used in this protocol. However, this system is extremely expensive, and it is usually found in clinical settings such as hospitals. By modifying the present protocol, future studies can systematically investigate and establish normative data of cortical mapping measures for TA and SOL in neurologically intact adults. Such findings will establish which motor mapping measures should be used to specifically quantify the motor representations of each muscle. Another potential application of the present protocol is the assessment of these two muscles before and after a surgery or an intervention (behavioral: exercise; neurophysiological: repetitive TMS, transcranial direct current stimulation - TDCS) and during the recovery period in athletic or clinical cohorts. This will allow rehabilitation scientists to determine how a surgery or an intervention may alter the cortical drive of these two muscles.

ACKNOWLEDGMENTS:

The authors thank Dr. Jesse C. Dean for helping with methodological development and providing feedback on a draft of the manuscript. This work was supported by a VA Career Development Award-2 RR&D N0787-W (MGB), an Institutional Development Award from the National Institute of General Medical Sciences of the NIH under grant number P20-GM109040 (SAK) and P2CHD086844 (SAK). The content does not represent the views of the Department of Veterans Affairs or the United States Government.

DISCLOSURES:

The authors have nothing to disclose.

REFERENCES:

- 1 Schünke, M., Schulte, E., Ross, L. M., Schumacher, U. & Lamperti, E. D. *Thieme Atlas of Anatomy: General Anatomy and Musculoskeletal System*. (Thieme, 2006).
- 2 Lieber, R. L. & Friden, J. Functional and clinical significance of skeletal muscle architecture. *Muscle Nerve*. **23** (11), 1647-1666, (2000).
- 3 Winter, D. A. *The biomechanics and motor control of human gait: Normal, Elderly and Pathological*. 2nd edn, (University of Waterloo Press, 1991).
- 4 Winter, D. A. *A.B.C. (anatomy, Biomechanics and Control) of Balance During Standing and Walking*. (Waterloo Biomechanics, 1995).
- 5 Nielsen, J. B. Motoneuronal drive during human walking. *Brain Research Reviews*. **40** (1-3), 192-201, (2002).
- 6 Nielsen, J. B. How we walk: central control of muscle activity during human walking. *Neuroscientist*. **9** (3), 195-204, (2003).
- 7 Davidoff, R. A. The pyramidal tract. *Neurology*. **40** (2), 332-339, (1990).

617 8 Nathan, P. W., Smith, M. C. & Deacon, P. The corticospinal tracts in man. Course and
618 location of fibres at different segmental levels. *Brain*. **113 (Pt 2)** 303-324, (1990).

619 9 Hallett, M. Transcranial magnetic stimulation and the human brain. *Nature*. **406** (6792),
620 147-150, (2000).

621 10 Hallett, M. Transcranial magnetic stimulation: a primer. *Neuron*. **55** (2), 187-199, (2007).

622 11 Brouwer, B., Ashby, P. & Midroni, G. Excitability of corticospinal neurons during tonic
623 muscle contractions in man. *Experimental Brain Research*. **74** (3), 649-652, (1989).

624 12 Advani, A. & Ashby, P. Corticospinal control of soleus motoneurons in man. *Canadian*
625 *Journal Physiology and Pharmacology*. **68** (9), 1231-1235, (1990).

626 13 Holmgren, H., Larsson, L. E. & Pedersen, S. Late muscular responses to transcranial cortical
627 stimulation in man. *Electroencephalography and Clinical Neurophysiology*. **75** (3), 161-
628 172, (1990).

629 14 Ackermann, H., Scholz, E., Koehler, W. & Dichgans, J. Influence of posture and voluntary
630 background contraction upon compound muscle action potentials from anterior tibial and
631 soleus muscle following transcranial magnetic stimulation. *Electroencephalography and*
632 *Clinical Neurophysiology*. **81** (1), 71-80, (1991).

633 15 Brouwer, B. & Ashby, P. Corticospinal projections to lower limb motoneurons in man.
634 *Experimental Brain Research*. **89** (3), 649-654, (1992).

635 16 Priori, A. *et al.* Transcranial electric and magnetic stimulation of the leg area of the human
636 motor cortex: single motor unit and surface EMG responses in the tibialis anterior muscle.
637 *Electroencephalography and Clinical Neurophysiology/Evoked Potentials Section*. **89** (2),
638 131-137, (1993).

639 17 Valls-Sole, J., Alvarez, R. & Tolosa, E. S. Responses of the soleus muscle to transcranial
640 magnetic stimulation. *Electroencephalography and Clinical Neurophysiology*. **93** (6), 421-
641 427, (1994).

642 18 Brouwer, B. & Qiao, J. Characteristics and variability of lower limb motoneuron responses
643 to transcranial magnetic stimulation. *Electroencephalography and Clinical*
644 *Neurophysiology*. **97** (1), 49-54, (1995).

645 19 Devanne, H., Lavoie, B. A. & Capaday, C. Input-output properties and gain changes in the
646 human corticospinal pathway. *Experimental Brain Research*. **114** (2), 329-338, (1997).

647 20 Capaday, C., Lavoie, B. A., Barbeau, H., Schneider, C. & Bonnard, M. Studies on the
648 corticospinal control of human walking. I. Responses to focal transcranial magnetic
649 stimulation of the motor cortex. *Journal of Neurophysiology*. **81** (1), 129-139, (1999).

650 21 Terao, Y. *et al.* Predominant activation of I1-waves from the leg motor area by transcranial
651 magnetic stimulation. *Brain Research*. **859** (1), 137-146, (2000).

652 22 Christensen, L. O., Andersen, J. B., Sinkjaer, T. & Nielsen, J. Transcranial magnetic
653 stimulation and stretch reflexes in the tibialis anterior muscle during human walking.
654 *Journal of Physiology*. **531** (Pt 2), 545-557, (2001).

655 23 Bawa, P., Chalmers, G. R., Stewart, H. & Eisen, A. A. Responses of ankle extensor and flexor
656 motoneurons to transcranial magnetic stimulation. *Journal of Neurophysiology*. **88** (1),
657 124-132, (2002).

658 24 Soto, O., Valls-Sole, J., Shanahan, P. & Rothwell, J. Reduction of intracortical inhibition in
659 soleus muscle during postural activity. *Journal of Neurophysiology*. **96** (4), 1711-1717,
660 (2006).

- 25 Barthelemy, D. *et al.* Impaired transmission in the corticospinal tract and gait disability in spinal cord injured persons. *Journal of Neurophysiology*. **104** (2), 1167-1176, (2010).
- 26 Barthelemy, D. *et al.* Functional implications of corticospinal tract impairment on gait after spinal cord injury. *Spinal Cord*. **51** (11), 852-856, (2013).
- 27 Beaulieu, L. D., Masse-Alarie, H., Brouwer, B. & Schneider, C. Brain control of volitional ankle tasks in people with chronic stroke and in healthy individuals. *Journal of Neurological Science*. **338** (1-2), 148-155, (2014).
- 28 Palmer, J. A., Hsiao, H., Awad, L. N. & Binder-Macleod, S. A. Symmetry of corticomotor input to plantarflexors influences the propulsive strategy used to increase walking speed post-stroke. *Clinical Neurophysiology*. **127** (3), 1837-1844, (2016).
- 29 Palmer, J. A., Needle, A. R., Pohlig, R. T. & Binder-Macleod, S. A. Atypical cortical drive during activation of the paretic and nonparetic tibialis anterior is related to gait deficits in chronic stroke. *Clinical Neurophysiology*. **127** (1), 716-723, (2016).
- 30 Palmer, J. A., Hsiao, H., Wright, T. & Binder-Macleod, S. A. Single Session of Functional Electrical Stimulation-Assisted Walking Produces Corticomotor Symmetry Changes Related to Changes in Poststroke Walking Mechanics. *Physical Therapy*. 10.1093/ptj/pzx008, (2017).
- 31 Palmer, J. A., Zarzycki, R., Morton, S. M., Kesar, T. M. & Binder-Macleod, S. A. Characterizing differential poststroke corticomotor drive to the dorsi- and plantarflexor muscles during resting and volitional muscle activation. *Journal of Neurophysiology*. **117** (4), 1615-1624, (2017).
- 32 Charalambous, C. C., Dean, J. C., Adkins, D. L., Hanlon, C. A. & Bowden, M. G. Characterizing the corticomotor connectivity of the bilateral ankle muscles during rest and isometric contraction in healthy adults. *Journal of Electromyography and Kinesiology*. **41** 9-18, (2018).
- 33 Kleim, J. A., Kleim, E. D. & Cramer, S. C. Systematic assessment of training-induced changes in corticospinal output to hand using frameless stereotaxic transcranial magnetic stimulation. *Nature Protocols*. **2** (7), 1675-1684, (2007).
- 34 Shellock, F. G. & Spinazzi, A. MRI safety update 2008: part 2, screening patients for MRI. *American Journal of Roentgenology*. **191** (4), 1140-1149, (2008).
- 35 Rossi, S., Hallett, M., Rossini, P. M. & Pascual-Leone, A. Screening questionnaire before TMS: an update. *Clinical Neurophysiology*. **122** (8), 1686, (2011).
- 36 Conti, A. *et al.* Navigated transcranial magnetic stimulation for "somatotopic" tractography of the corticospinal tract. *Neurosurgery*. **10 Suppl 4** 542-554; discussion 554, (2014).
- 37 Comeau, R. in *Transcranial Magnetic Stimulation*. 31-56 (Springer, 2014).
- 38 Cram, J. R. & Criswell, E. *Cram's Introduction to Surface Electromyography*. (Jones & Bartlett Learning, 2011).
- 39 Hermens, H. J., Freriks, B., Merletti, R., Stegeman, D., Blok, J., Rau, G., Disselhorst- & Klug, C., Hagg, G.,. *European Recommendations for Surface ElectroMyoGraphy: Results of the Seniam Project (SENIAM)*. 2nd edn, citeulike-article-id:5280603 (Roessingh Research and Development, 1999).
- 40 Awiszus, F. TMS and threshold hunting. *Supplements to Clinical Neurophysiology*. **56** 13-23, (2003).

705 41 Sinclair, C., Faulkner, D. & Hammond, G. Flexible real-time control of MagStim 200(2) units
706 for use in transcranial magnetic stimulation studies. *Journal of Neuroscience Methods*.
707 **158** (1), 133-136, (2006).

708 42 Alkadhi, H. *et al.* Reproducibility of primary motor cortex somatotopy under controlled
709 conditions. *American Journal of Neuroradiology*. **23** (9), 1524-1532, (2002).

710 43 Rossini, P. M. *et al.* Applications of magnetic cortical stimulation. The International
711 Federation of Clinical Neurophysiology. *Electroencephalography and Clinical*
712 *Neurophysiology Supplement*. **52** 171-185, (1999).

713 44 Borckardt, J. J., Nahas, Z., Koola, J. & George, M. S. Estimating resting motor thresholds in
714 transcranial magnetic stimulation research and practice: a computer simulation
715 evaluation of best methods. *Journal for ECT*. **22** (3), 169-175, (2006).

716 45 Livingston, S. C., Friedlander, D. L., Gibson, B. C. & Melvin, J. R. Motor evoked potential
717 response latencies demonstrate moderate correlations with height and limb length in
718 healthy young adults. *The Neurodiagnostic Journal*. **53** (1), 63-78, (2013).

719 46 Cacchio, A. *et al.* Reliability of TMS-related measures of tibialis anterior muscle in patients
720 with chronic stroke and healthy subjects. *Journal of Neurological Science*. **303** (1-2), 90-
721 94, (2011).

722 47 Saisanen, L. *et al.* Factors influencing cortical silent period: optimized stimulus location,
723 intensity and muscle contraction. *Journal of Neuroscience Methods*. **169** (1), 231-238,
724 (2008).

725 48 Ertekin, C. *et al.* A stable late soleus EMG response elicited by cortical stimulation during
726 voluntary ankle dorsiflexion. *Electroencephalography and Clinical*
727 *Neurophysiology/Electromyography and Motor Control*. **97** (5), 275-283, (1995).

728 49 Tarkka, I. M., McKay, W. B., Sherwood, A. M. & Dimitrijevic, M. R. Early and late motor
729 evoked potentials reflect preset agonist-antagonist organization in lower limb muscles.
730 *Muscle Nerve*. **18** (3), 276-282, (1995).

731 50 Ziemann, U. *et al.* Dissociation of the pathways mediating ipsilateral and contralateral
732 motor-evoked potentials in human hand and arm muscles. *Journal of Physiology*. **518** (Pt
733 **3**) 895-906, (1999).

734 51 McCambridge, A. B., Stinear, J. W. & Byblow, W. D. Are ipsilateral motor evoked potentials
735 subject to intracortical inhibition? *Journal of Neurophysiology*. **115** (3), 1735-1739, (2016).

736 52 Tazoe, T. & Perez, M. A. Selective activation of ipsilateral motor pathways in intact
737 humans. *Journal of Neuroscience*. **34** (42), 13924-13934, (2014).

738 53 Chen, R., Yung, D. & Li, J. Y. Organization of ipsilateral excitatory and inhibitory pathways
739 in the human motor cortex. *Journal of Neurophysiology*. **89** (3), 1256-1264, (2003).

740 54 Wassermann, E. M., Pascual-Leone, A. & Hallett, M. Cortical motor representation of the
741 ipsilateral hand and arm. *Experimental Brain Research*. **100** (1), 121-132, (1994).

742 55 Kesar, T. M., Stinear, J. W. & Wolf, S. L. The use of transcranial magnetic stimulation to
743 evaluate cortical excitability of lower limb musculature: Challenges and opportunities.
744 *Restorative Neurology and Neuroscience*. **36** (3), 333-348, (2018).

745 56 Lefaucheur, J. P. Why image-guided navigation becomes essential in the practice of
746 transcranial magnetic stimulation. *Neurophysiologie Clinique/Clinical Neurophysiology*.
747 **40** (1), 1-5, (2010).

748 57 Sparing, R., Hesse, M. D. & Fink, G. R. Neuronavigation for transcranial magnetic

- stimulation (TMS): where we are and where we are going. *Cortex*. **46** (1), 118-120, (2010).
- 58 Sparing, R., Buelte, D., Meister, I. G., Pauš, T. & Fink, G. R. Transcranial magnetic stimulation and the challenge of coil placement: a comparison of conventional and stereotaxic neuronavigational strategies. *Human Brain Mapping*. **29** (1), 82-96, (2008).
- 59 Gugino, L. D. *et al.* Transcranial magnetic stimulation coregistered with MRI: a comparison of a guided versus blind stimulation technique and its effect on evoked compound muscle action potentials. *Clinical Neurophysiology*. **112** (10), 1781-1792, (2001).
- 60 Jung, N. H. *et al.* Navigated transcranial magnetic stimulation does not decrease the variability of motor-evoked potentials. *Brain Stimulation*. **3** (2), 87-94, (2010).
- 61 Terao, Y. & Ugawa, Y. Basic mechanisms of TMS. *J Clin Neurophysiol*. **19** (4), 322-343, (2002).
- 62 Madhavan, S., Rogers, L. M. & Stinear, J. W. A paradox: after stroke, the non-lesioned lower limb motor cortex may be maladaptive. *European Journal of Neuroscience*. **32** (6), 1032-1039, (2010).
- 63 Kujirai, T. *et al.* Corticocortical inhibition in human motor cortex. *Journal of Physiology*. **471** 501-519, (1993).
- 64 Ziemann, U. Intracortical inhibition and facilitation in the conventional paired TMS paradigm. *Electroencephalography and Clinical Neurophysiology Supplement*. **51** 127-136, (1999).
- 65 Cavaleri, R., Schabrun, S. M. & Chipchase, L. S. The number of stimuli required to reliably assess corticomotor excitability and primary motor cortical representations using transcranial magnetic stimulation (TMS): a systematic review and meta-analysis. *Systematic Reviews*. **6** (1), 48, (2017).
- 66 Goldsworthy, M. R., Hordacre, B. & Ridding, M. C. Minimum number of trials required for within- and between-session reliability of TMS measures of corticospinal excitability. *Neuroscience*. **320** 205-209, (2016).
- 67 Cavaleri, R., Schabrun, S. M. & Chipchase, L. S. Determining the Optimal Number of Stimuli per Cranial Site during Transcranial Magnetic Stimulation Mapping. *Neuroscience Journal*. **2017** 6328569, (2017).
- 68 Groppa, S. *et al.* A practical guide to diagnostic transcranial magnetic stimulation: report of an IFCN committee. *Clinical Neurophysiology*. **123** (5), 858-882, (2012).
- 69 Rossini, P. M. *et al.* Non-invasive electrical and magnetic stimulation of the brain, spinal cord and roots: basic principles and procedures for routine clinical application. Report of an IFCN committee. *Electroencephalography and Clinical Neurophysiology*. **91** (2), 79-92, (1994).
- 70 Rothwell, J. C. *et al.* Magnetic stimulation: motor evoked potentials. The International Federation of Clinical Neurophysiology. *Electroencephalography and Clinical Neurophysiology Supplement*. **52** 97-103, (1999).
- 71 Silbert, B. I., Patterson, H. I., Pevcic, D. D., Windnagel, K. A. & Thickbroom, G. W. A comparison of relative-frequency and threshold-hunting methods to determine stimulus intensity in transcranial magnetic stimulation. *Clinical Neurophysiology*. **124** (4), 708-712, (2013).
- 72 Obata, H., Sekiguchi, H., Nakazawa, K. & Ohtsuki, T. Enhanced excitability of the corticospinal pathway of the ankle extensor and flexor muscles during standing in

humans. *Experimental Brain Research*. **197** (3), 207-213, (2009).

73 Tokuno, C. D., Taube, W. & Cresswell, A. G. An enhanced level of motor cortical excitability during the control of human standing. *Acta Physiologica (Oxf)*. **195** (3), 385-395, (2009).

74 Obata, H., Sekiguchi, H., Ohtsuki, T. & Nakazawa, K. Posture-related modulation of cortical excitability in the tibialis anterior muscle in humans. *Brain Research*. **1577** 29-35, (2014).

75 Remaud, A., Bilodeau, M. & Tremblay, F. Age and Muscle-Dependent Variations in Corticospinal Excitability during Standing Tasks. *PLoS ONE*. **9** (10), e110004, (2014).

76 Baudry, S., Collignon, S. & Duchateau, J. Influence of age and posture on spinal and corticospinal excitability. *Experimental Gerontology*. **69** 62-69, (2015).

77 Petersen, N. T. *et al.* Suppression of EMG activity by transcranial magnetic stimulation in human subjects during walking. *Journal of Physiology*. **537** (Pt 2), 651-656, (2001).

78 Schubert, M., Curt, A., Jensen, L. & Dietz, V. Corticospinal input in human gait: modulation of magnetically evoked motor responses. *Experimental Brain Research*. **115** (2), 234-246, (1997).

79 Schubert, M., Curt, A., Colombo, G., Berger, W. & Dietz, V. Voluntary control of human gait: conditioning of magnetically evoked motor responses in a precision stepping task. *Experimental Brain Research*. **126** (4), 583-588, (1999).

80 Ngomo, S., Leonard, G., Moffet, H. & Mercier, C. Comparison of transcranial magnetic stimulation measures obtained at rest and under active conditions and their reliability. *Journal of Neuroscience Methods*. **205** (1), 65-71, (2012).

81 Niskanen, E. *et al.* Group-level variations in motor representation areas of thenar and anterior tibial muscles: Navigated Transcranial Magnetic Stimulation Study. *Human Brain Mapping*. **31** (8), 1272-1280, (2010).

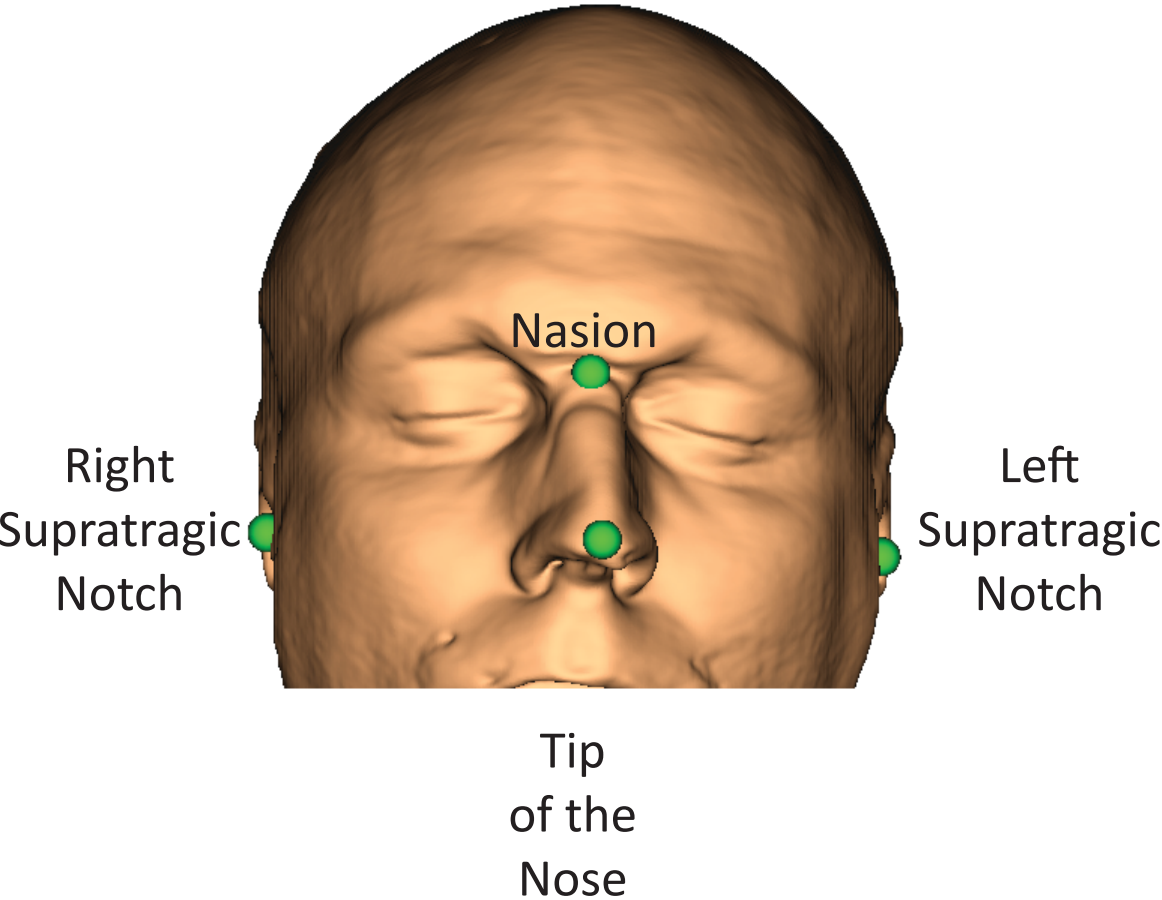
82 Thordstein, M., Saar, K., Pegenius, G. & Elam, M. Individual effects of varying stimulation intensity and response criteria on area of activation for different muscles in humans. A study using navigated transcranial magnetic stimulation. *Brain Stimulation*. **6** (1), 49-53, (2013).

83 Vaalto, S. *et al.* Long-term plasticity may be manifested as reduction or expansion of cortical representations of actively used muscles in motor skill specialists. *Neuroreport*. **24** (11), 596-600, (2013).

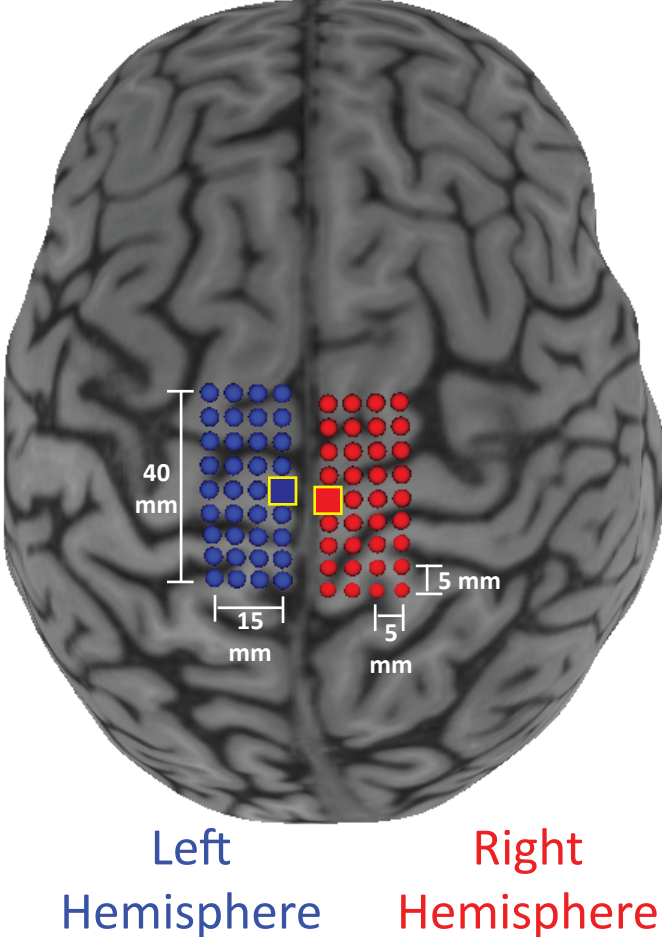
84 Forster, M. T., Limbart, M., Seifert, V. & Senft, C. Test-retest reliability of navigated transcranial magnetic stimulation of the motor cortex. *Neurosurgery*. **10 Suppl 1** 51-55; discussion 55-56, (2014).

85 Saisanen, L. *et al.* Non-invasive preoperative localization of primary motor cortex in epilepsy surgery by navigated transcranial magnetic stimulation. *Epilepsy Research*. **92** (2-3), 134-144, (2010).

A. 3D Skin Model

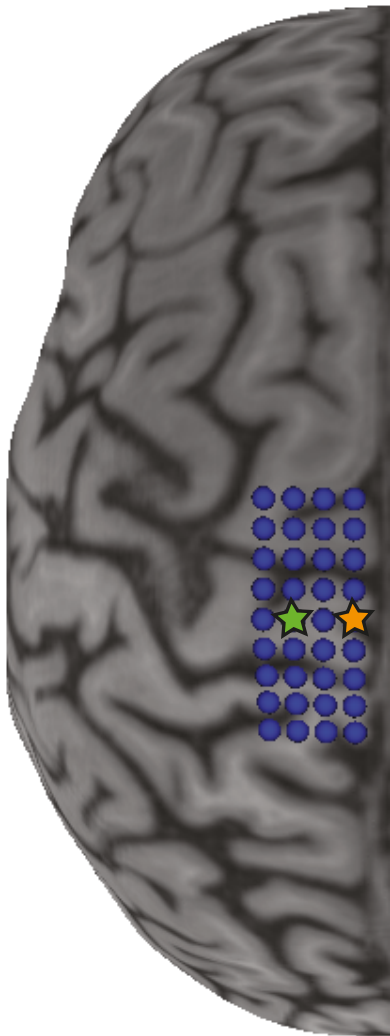


B. 3D Full Brain Curvilinear



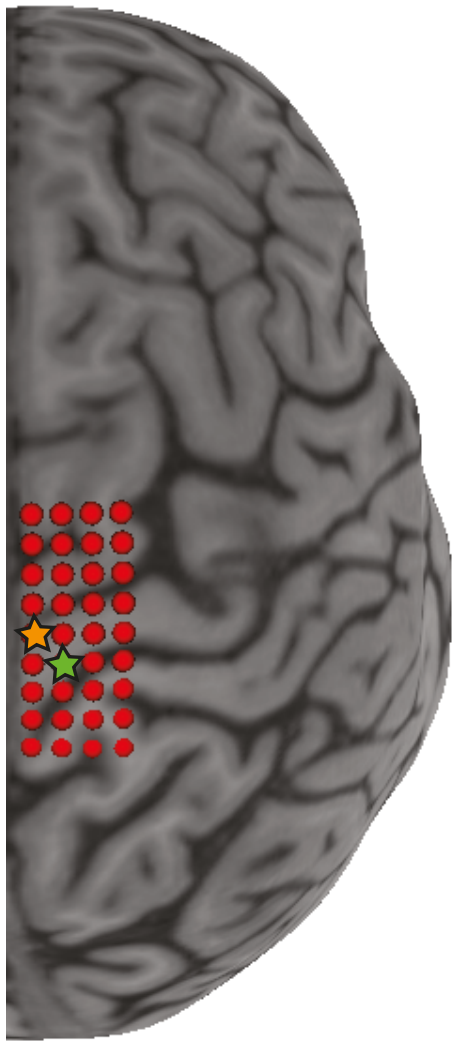
**A. TMS Over
Left Hemisphere -
Right/Contralateral**

Hot Spot ☆



**B. TMS Over Right
Hemisphere -
Left/Contralateral**

Hot Spot ☆

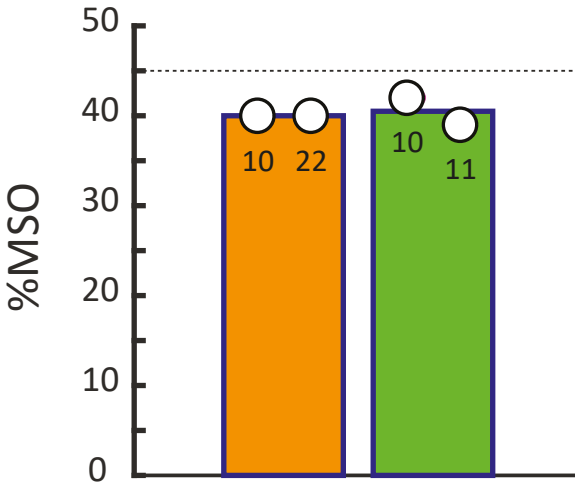
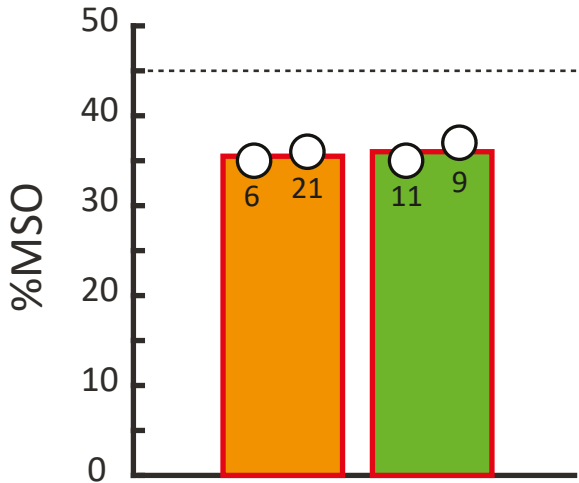


Tibialis
Anterior

RMT

Soleus

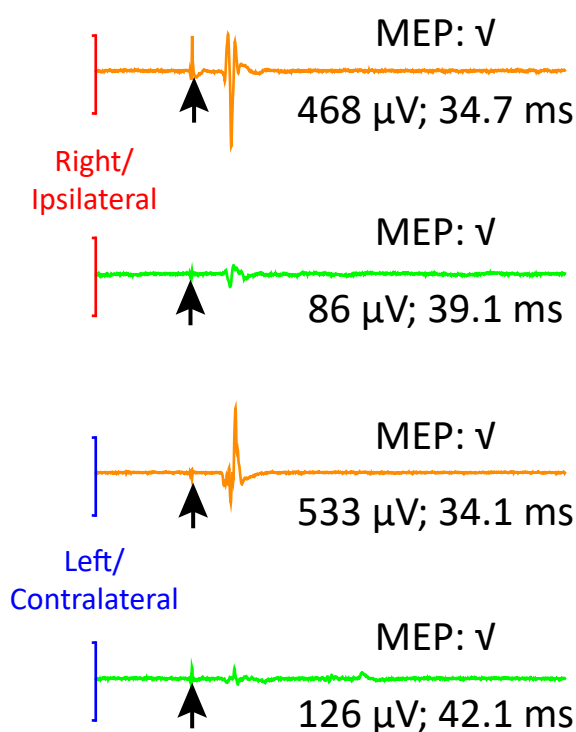
RMT



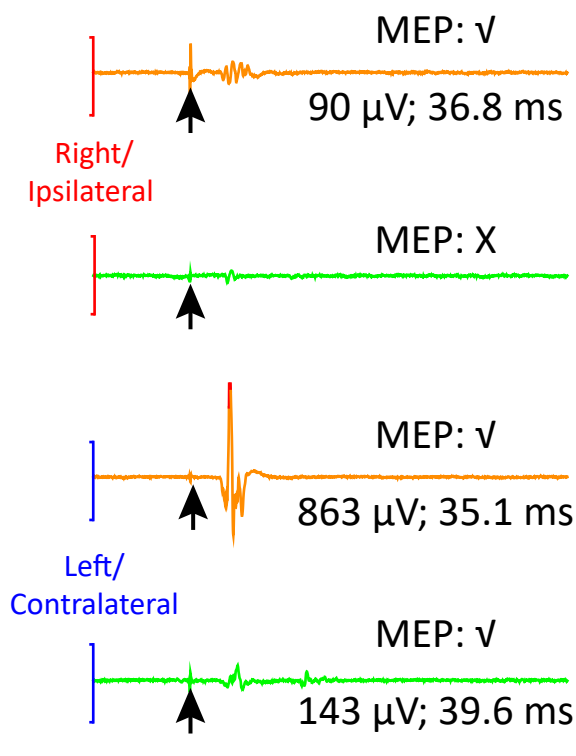
[Click here](#)
R1_Fig3.a

Right Hemisphere

C. Left/Contralateral TA Hot Spot (48% MSO)



D. Left/Contralateral SOL Hot Spot (49% MSO)



↑ TMS Onset

Figure 4

Suprathreshold (1.2 RMT) TMS During TVA

[Click here to access/download:Figure,JoVE-RN_Fig4.a](#)

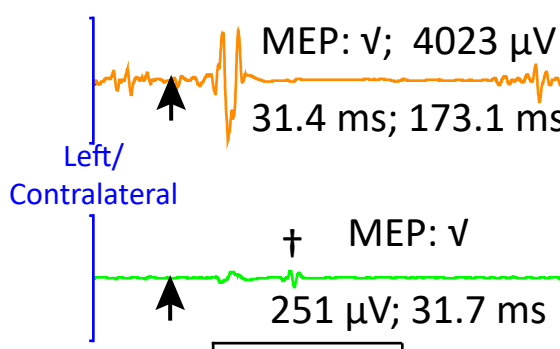
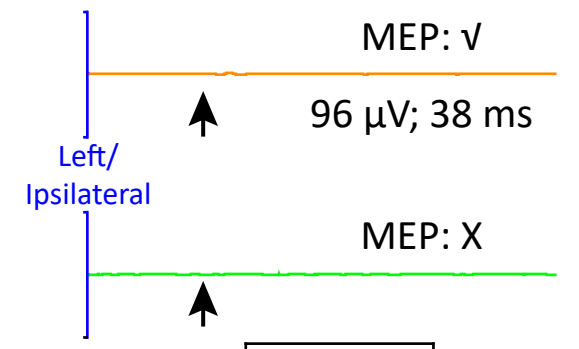
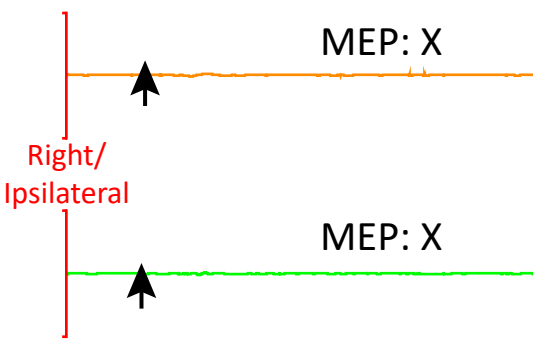
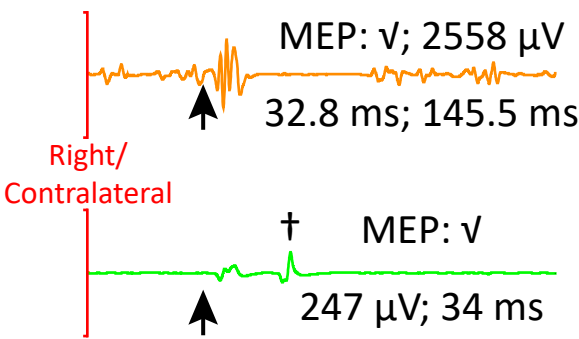


Left Hemisphere

Right Hemisphere

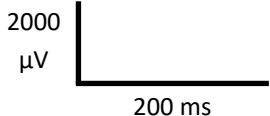
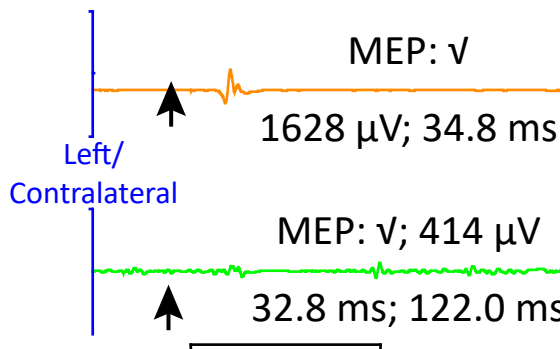
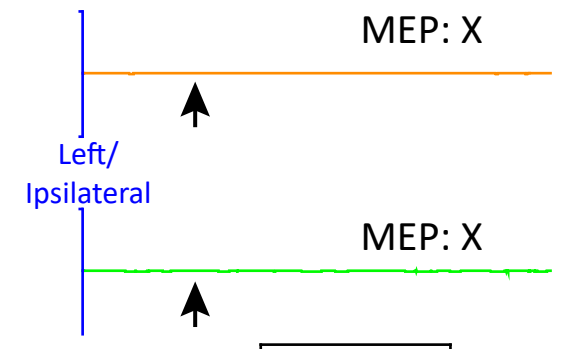
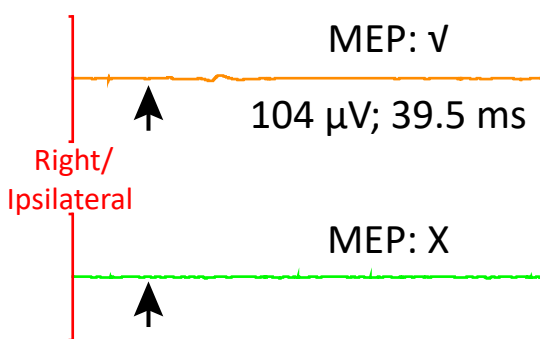
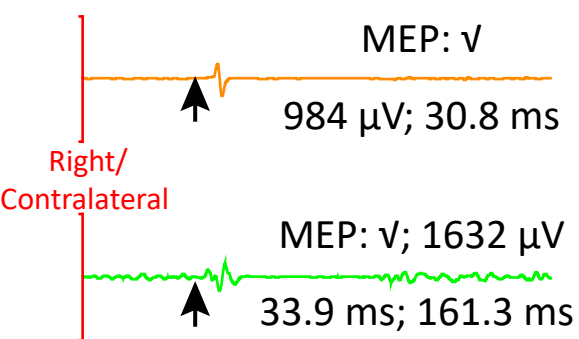
A. Right/Contralateral TA Hot Spot (43% MSO)

C. Left/Contralateral TA Hot Spot (48% MSO)



B. Right/Contralateral SOL Hot Spot (43% MSO)

D. Left/Contralateral SOL Hot Spot (49% MSO)



Tibialis Anterior
Soleus

↑ TMS Onset

Name of Material/ Equipment	Company	Catalog Number	Comments/Description
2 Magstim stimulators (Bistim module)	The Magstim Company Limited; Whitland, UK		Used to elicit bilateral motor evoked potentials in tibialis anterior and soleus muscles.
Adaptive parameter estimation by sequential testing (PEST) for TMS	http://www.clinicalresearcher.org/software.htm		Used to determine motor thresholds.
Amplifier	Motion Lab Systems; Baton Rouge, LN, USA	MA-300	Used to amplify EMG data.
Data Acquisition Unit	Motion Lab Systems; Baton Rouge, LN, USA	Micro 1401	Used to acquire EMG data.

Double cone coil	The Magstim Company Limited; Whitland, UK	PN: 9902AP	Used to elicit bilateral motor evoked potentials in tibialis anterior and soleus muscles.
Polaris	Northen Digital Inc.; Waterloo, Ontario, Canada		Used to track the reflective markers located on subject tracker and coil tracker.
Signal	Cambridge Electronics Design Limited; Cambridge, UK	version 6	Used to collect motor evoked potentials during rest and TVA.
Single double differential surface EMG electrodes	Motion Lab Systems; Baton Rouge, LN, USA	MA-411	Used to record EMG signals.
TMS Frameless Stereotaxy Neuronavigation Sytem	Brainsight 3, Rouge Research, Montreal, Canada		Used to navigate coil position during TMS assessment.

Walker boot

Mountainside Medical
Equipment, Marcy, NY

Used to stabilize ankle joint.

ARTICLE AND VIDEO LICENSE AGREEMENT

Title of Article: **Bilateral assessment of the corticospinal pathways of the ankle muscles using frameless Stereotaxic transcranial magnetic stimulation.**

Author(s): **Charalambos C. Charalambous**

Item 1: The Author elects to have the Materials be made available (as described at <http://www.jove.com/publish>) via:

☒ Standard Access

☐ Open Access

Item 2: Please select one of the following items:

☒ The Author is **NOT** a United States government employee.

☐ The Author is a United States government employee and the Materials were prepared in the course of his or her duties as a United States government employee.

☐ The Author is a United States government employee but the Materials were NOT prepared in the course of his or her duties as a United States government employee.

ARTICLE AND VIDEO LICENSE AGREEMENT

1. **Defined Terms.** As used in this Article and Video License Agreement, the following terms shall have the following meanings: "Agreement" means this Article and Video License Agreement; "Article" means the article specified on the last page of this Agreement, including any associated materials such as texts, figures, tables, artwork, abstracts, or summaries contained therein; "Author" means the author who is a signatory to this Agreement; "Collective Work" means a work, such as a periodical issue, anthology or encyclopedia, in which the Materials in their entirety in unmodified form, along with a number of other contributions, constituting separate and independent works in themselves, are assembled into a collective whole; "CRC License" means the Creative Commons Attribution-Non Commercial-No Derivs 3.0 Unported Agreement, the terms and conditions of which can be found at: <http://creativecommons.org/licenses/by-nc-nd/3.0/legalcode>; "Derivative Work" means a work based upon the Materials or upon the Materials and other pre-existing works, such as a translation, musical arrangement, dramatization, fictionalization, motion picture version, sound recording, art reproduction, abridgment, condensation, or any other form in which the Materials may be recast, transformed, or adapted; "Institution" means the institution, listed on the last page of this Agreement, by which the Author was employed at the time of the creation of the Materials; "JoVE" means MyJoVE Corporation, a Massachusetts corporation and the publisher of The Journal of Visualized Experiments; "Materials" means the Article and / or the Video; "Parties" means the Author and JoVE; "Video" means any video(s) made by the Author, alone or in conjunction with any other parties, or by JoVE or its affiliates or agents, individually or in collaboration with the Author or any other parties, incorporating all or any portion

of the Article, and in which the Author may or may not appear.

2. **Background.** The Author, who is the author of the Article, in order to ensure the dissemination and protection of the Article, desires to have the JoVE publish the Article and create and transmit videos based on the Article. In furtherance of such goals, the Parties desire to memorialize in this Agreement the respective rights of each Party in and to the Article and the Video.

3. **Grant of Rights in Article.** In consideration of JoVE agreeing to publish the Article, the Author hereby grants to JoVE, subject to Sections 4 and 7 below, the exclusive, royalty-free, perpetual (for the full term of copyright in the Article, including any extensions thereto) license (a) to publish, reproduce, distribute, display and store the Article in all forms, formats and media whether now known or hereafter developed (including without limitation in print, digital and electronic form) throughout the world, (b) to translate the Article into other languages, create adaptations, summaries or extracts of the Article or other Derivative Works (including, without limitation, the Video) or Collective Works based on all or any portion of the Article and exercise all of the rights set forth in (a) above in such translations, adaptations, summaries, extracts, Derivative Works or Collective Works and (c) to license others to do any or all of the above. The foregoing rights may be exercised in all media and formats, whether now known or hereafter devised, and include the right to make such modifications as are technically necessary to exercise the rights in other media and formats. If the "Open Access" box has been checked in Item 1 above, JoVE and the Author hereby grant to the public all such rights in the Article as provided in, but subject to all limitations and requirements set forth in, the CRC License.

ARTICLE AND VIDEO LICENSE AGREEMENT

4. **Retention of Rights in Article.** Notwithstanding the exclusive license granted to JoVE in Section 3 above, the Author shall, with respect to the Article, retain the non-exclusive right to use all or part of the Article for the non-commercial purpose of giving lectures, presentations or teaching classes, and to post a copy of the Article on the Institution's website or the Author's personal website, in each case provided that a link to the Article on the JoVE website is provided and notice of JoVE's copyright in the Article is included. All non-copyright intellectual property rights in and to the Article, such as patent rights, shall remain with the Author.

5. **Grant of Rights in Video – Standard Access.** This Section 5 applies if the "Standard Access" box has been checked in Item 1 above or if no box has been checked in Item 1 above. In consideration of JoVE agreeing to produce, display or otherwise assist with the Video, the Author hereby acknowledges and agrees that, Subject to Section 7 below, JoVE is and shall be the sole and exclusive owner of all rights of any nature, including, without limitation, all copyrights, in and to the Video. To the extent that, by law, the Author is deemed, now or at any time in the future, to have any rights of any nature in or to the Video, the Author hereby disclaims all such rights and transfers all such rights to JoVE.

6. **Grant of Rights in Video – Open Access.** This Section 6 applies only if the "Open Access" box has been checked in Item 1 above. In consideration of JoVE agreeing to produce, display or otherwise assist with the Video, the Author hereby grants to JoVE, subject to Section 7 below, the exclusive, royalty-free, perpetual (for the full term of copyright in the Article, including any extensions thereto) license (a) to publish, reproduce, distribute, display and store the Video in all forms, formats and media whether now known or hereafter developed (including without limitation in print, digital and electronic form) throughout the world, (b) to translate the Video into other languages, create adaptations, summaries or extracts of the Video or other Derivative Works or Collective Works based on all or any portion of the Video and exercise all of the rights set forth in (a) above in such translations, adaptations, summaries, extracts, Derivative Works or Collective Works and (c) to license others to do any or all of the above. The foregoing rights may be exercised in all media and formats, whether now known or hereafter devised, and include the right to make such modifications as are technically necessary to exercise the rights in other media and formats. For any Video to which this Section 6 is applicable, JoVE and the Author hereby grant to the public all such rights in the Video as provided in, but subject to all limitations and requirements set forth in, the CRC License.

7. **Government Employees.** If the Author is a United States government employee and the Article was prepared in the course of his or her duties as a United States government employee, as indicated in Item 2 above, and any of the licenses or grants granted by the Author hereunder exceed the scope of the 17 U.S.C. 403, then the rights granted hereunder shall be limited to the maximum

rights permitted under such statute. In such case, all provisions contained herein that are not in conflict with such statute shall remain in full force and effect, and all provisions contained herein that do so conflict shall be deemed to be amended so as to provide to JoVE the maximum rights permissible within such statute.

8. **Protection of the Work.** The Author(s) authorize JoVE to take steps in the Author(s) name and on their behalf if JoVE believes some third party could be infringing or might infringe the copyright of either the Author's Article and/or Video.

9. **Likeness, Privacy, Personality.** The Author hereby grants JoVE the right to use the Author's name, voice, likeness, picture, photograph, image, biography and performance in any way, commercial or otherwise, in connection with the Materials and the sale, promotion and distribution thereof. The Author hereby waives any and all rights he or she may have, relating to his or her appearance in the Video or otherwise relating to the Materials, under all applicable privacy, likeness, personality or similar laws.

10. **Author Warranties.** The Author represents and warrants that the Article is original, that it has not been published, that the copyright interest is owned by the Author (or, if more than one author is listed at the beginning of this Agreement, by such authors collectively) and has not been assigned, licensed, or otherwise transferred to any other party. The Author represents and warrants that the author(s) listed at the top of this Agreement are the only authors of the Materials. If more than one author is listed at the top of this Agreement and if any such author has not entered into a separate Article and Video License Agreement with JoVE relating to the Materials, the Author represents and warrants that the Author has been authorized by each of the other such authors to execute this Agreement on his or her behalf and to bind him or her with respect to the terms of this Agreement as if each of them had been a party hereto as an Author. The Author warrants that the use, reproduction, distribution, public or private performance or display, and/or modification of all or any portion of the Materials does not and will not violate, infringe and/or misappropriate the patent, trademark, intellectual property or other rights of any third party. The Author represents and warrants that it has and will continue to comply with all government, institutional and other regulations, including, without limitation all institutional, laboratory, hospital, ethical, human and animal treatment, privacy, and all other rules, regulations, laws, procedures or guidelines, applicable to the Materials, and that all research involving human and animal subjects has been approved by the Author's relevant institutional review board.

11. **JoVE Discretion.** If the Author requests the assistance of JoVE in producing the Video in the Author's facility, the Author shall ensure that the presence of JoVE employees, agents or independent contractors is in accordance with the relevant regulations of the Author's institution. If more than one author is listed at the beginning of this Agreement, JoVE may, in its sole

ARTICLE AND VIDEO LICENSE AGREEMENT

discretion, elect not take any action with respect to the Article until such time as it has received complete, executed Article and Video License Agreements from each such author. JoVE reserves the right, in its absolute and sole discretion and without giving any reason therefore, to accept or decline any work submitted to JoVE. JoVE and its employees, agents and independent contractors shall have full, unfettered access to the facilities of the Author or of the Author's institution as necessary to make the Video, whether actually published or not. JoVE has sole discretion as to the method of making and publishing the Materials, including, without limitation, to all decisions regarding editing, lighting, filming, timing of publication, if any, length, quality, content and the like.

12. **Indemnification.** The Author agrees to indemnify JoVE and/or its successors and assigns from and against any and all claims, costs, and expenses, including attorney's fees, arising out of any breach of any warranty or other representations contained herein. The Author further agrees to indemnify and hold harmless JoVE from and against any and all claims, costs, and expenses, including attorney's fees, resulting from the breach by the Author of any representation or warranty contained herein or from allegations or instances of violation of intellectual property rights, damage to the Author's or the Author's institution's facilities, fraud, libel, defamation, research, equipment, experiments, property damage, personal injury, violations of institutional, laboratory, hospital, ethical, human and animal treatment, privacy or other rules, regulations, laws, procedures or guidelines, liabilities and other losses or damages related in any way to the submission of work to JoVE, making of videos by JoVE, or publication in JoVE or elsewhere by JoVE. The Author shall be responsible for, and shall hold JoVE harmless from, damages caused by lack of sterilization, lack of cleanliness or by contamination due to

the making of a video by JoVE its employees, agents or independent contractors. All sterilization, cleanliness or decontamination procedures shall be solely the responsibility of the Author and shall be undertaken at the Author's expense. All indemnifications provided herein shall include JoVE's attorney's fees and costs related to said losses or damages. Such indemnification and holding harmless shall include such losses or damages incurred by, or in connection with, acts or omissions of JoVE, its employees, agents or independent contractors.

13. **Fees.** To cover the cost incurred for publication, JoVE must receive payment before production and publication the Materials. Payment is due in 21 days of invoice. Should the Materials not be published due to an editorial or production decision, these funds will be returned to the Author. Withdrawal by the Author of any submitted Materials after final peer review approval will result in a US\$1,200 fee to cover pre-production expenses incurred by JoVE. If payment is not received by the completion of filming, production and publication of the Materials will be suspended until payment is received.

14. **Transfer, Governing Law.** This Agreement may be assigned by JoVE and shall inure to the benefits of any of JoVE's successors and assignees. This Agreement shall be governed and construed by the internal laws of the Commonwealth of Massachusetts without giving effect to any conflict of law provision thereunder. This Agreement may be executed in counterparts, each of which shall be deemed an original, but all of which together shall be deemed to be one and the same agreement. A signed copy of this Agreement delivered by facsimile, e-mail or other means of electronic transmission shall be deemed to have the same legal effect as delivery of an original signed copy of this Agreement.

A signed copy of this document must be sent with all new submissions. Only one Agreement is required per submission.

CORRESPONDING AUTHOR

Name:

Name:	Charalambos C. Charalambous
Department:	Department of Health Sciences & Research
Institution:	Medical University South Carolina
Title:	

Signature:



Date:

8/14/18

Please submit a **signed** and **dated** copy of this license by one of the following three methods:

1. Upload an electronic version on the JoVE submission site
2. Fax the document to +1.866.381.2236
3. Mail the document to JoVE / Attn: JoVE Editorial / 1 Alewife Center #200 / Cambridge, MA 02140

JoVE58944 "Bilateral assessment of the corticospinal pathways of the ankle muscles using frameless stereotaxic transcranial magnetic stimulation,"

We thank the editor and the reviewers for their constructive and thoughtful comments, which we believe have resulted in a significantly improved manuscript. We have responded to editor's and each reviewer's comments below. Where appropriate, we refer to sections in the revised manuscript. All changes in the manuscript are indicated using track changes. We have highlighted the sections that will be recorded.

Response to Editor and Reviewers:

Editor:

1. Please take this opportunity to thoroughly proofread the manuscript to ensure that there are no spelling or grammar issues.

Response: We thank the editor for this reminder.

Action: We have thoroughly proofread the manuscript to ensure that there are no spelling or grammar issues.

2. Please adjust the numbering of the Protocol to follow the JoVE Instructions for Authors. For example, 1 should be followed by 1.1 and then 1.1.1 and 1.1.2 if necessary. Please refrain from using bullets, dashes, or indentations.

Response: We thank the editor for this instruction.

Action: We have numbered correctly all sections.

3. Please add more details to your protocol steps. There should be enough detail in each step to supplement the actions seen in the video so that viewers can easily replicate the protocol. Please ensure you answer the "how" question, i.e., how is the step performed? Alternatively, add references to published material specifying how to perform the protocol action. See examples below:

In the JoVE Protocol format, "Notes" should be concise and used sparingly. They should only be used to provide extraneous details, optional steps, or recommendations that are not critical to a step. Any text that provides details (e.g., lines 103-109, 138-145, 204-207, 216-220, 225-231, 234-248, 254-262, 291-297, 309-318, 330-368, etc.) about how to perform a particular step should either be included in the step itself or added as a sub-step. Please consider moving some of the notes about the protocol to the discussion section.

Response: We apologize for any confusion.

Action: We have converted all the NOTES into steps.

4. Lines 135: This step does not have sufficient details to replicate. Please provide more details.

Response: We apologize for any missing information.

Action: We have elaborated on this step.

“2.2.1 Reconstruct the skin and full curvilinear brain model by adjusting the bounding box around the skull and brain tissue, respectively. Identify four anatomical landmarks (tip of the nose, nasion - bridge of the nose, and supratragic notch of the right and left ear) using the skin model (see Figure 1A).”

5. Line 211: It is unclear how this is done.

Response: We apologize for the insufficient clarity of this step.

Action: We have added additional information to improve the clarity of this step.

“4.2 Calculate the maximum muscle activity value during each MVIC (i.e., the average within a 100 ms window centered around the maximum rectified and smoothed EMG) of the last three trials, the average of the three values, and the 15 % and 5 % of each muscle’s average MVIC.”

6. Lines 251-262: Please break up into sub-steps.

Response: Thank you for the suggestion.

Action: Per editor’s recommendation, we broke up this section into sub-steps.

“6.2.1 Find the suprathreshold intensity, which will be used during the hot spot hunting, by applying a single stimulus over the centered spot next to the interhemispheric fissure (see blue and red squares in Figure 1B). Use this spot because it is located at the locus of the leg motor area^{36,42}.

6.2.2 Start at low intensity (e.g., 30 % maximum stimulator output; MSO) and gradually increase the TMS intensity by 5 % increments, until reaching the intensity that elicits a motor evoked potential (MEP) with a peak-to-peak amplitude greater than 50 μ V in all contralateral examined muscles for 3 consecutive stimuli.

6.2.3 Determine immediately after each stimulus whether a MEP has been elicited based on both the raw waveforms and peak-to-peak amplitudes (search window: 20-60 ms post-TMS onset) of all examined muscles.”

7. Please combine some of the shorter Protocol steps so that individual steps contain 2-3 actions and maximum of 4 sentences per step.

Response: We thank the editor for this suggestion.

Action: We have revised the manuscript to combine short Protocol steps where feasible and appropriate.

8. Please include single-line spaces between all paragraphs, headings, steps, etc.

Response: We thank the editor for this instruction.

Action: We have included single-line spaces between all paragraphs, headings, steps, etc.

9. After you have made all the recommended changes to your protocol (listed above), please highlight 2.75 pages or less of the Protocol (including headings and spacing) that identifies the essential steps of the protocol for the video, i.e., the steps that should be visualized to tell the most cohesive story of the Protocol. Please note that calculations are not appropriate for filming.

Response: We thank editor for this instruction

Action: We have highlighted the appropriate sections.

10. Please highlight complete sentences (not parts of sentences). Please ensure that the highlighted part of the step includes at least one action that is written in imperative tense.

Response: We thank editor for this instruction

Action: We highlighted complete sentences.

11. Please include all relevant details that are required to perform the step in the highlighting. For example: If step 2.5 is highlighted for filming and the details of how to perform the step are given in steps 2.5.1 and 2.5.2, then the sub-steps where the details are provided must be highlighted.

Response: We thank editor for this instruction

Action: We highlighted sections per editor's instructions.

12. Figures 3 and 4: Please include a space between all numbers and their corresponding units (260 μ V, 34.6 ms, etc.).

Response: We apologize for any confusion.

Action: We have added space between values and units.

13. References: Please do not abbreviate journal titles.

Response: We apologize for any confusion.

Action: We have corrected all journal titles.

Reviewer #1:

1. The strong recommendation to use a neuronavigation system is not well-supported, given the authors' comment that no study has yet determined whether neuronavigation improves accuracy of lower extremity TMS measures (line 507). It seems that the "true" hotspot could be identified without neuronavigation by selecting the location that produces MEPs with the shortest latency using the lowest stimulus intensity identified by PEST - reflecting direct activation of the hotspot, rather than oligosynaptic ipsilateral MEPs (which would produce longer latency) or current spread (which would require higher intensity). Given the costs of MRI and neuronavigation, it would be a pity to suggest that these are an absolute necessity, as this would discourage research in an area that already receives relatively little attention.

Response: The reviewer raises an important point, and we apologize for the misunderstanding. It was not our intention to strongly recommend that neuronavigation is required for the assessment of the lower extremity TMS measures. In the present manuscript, our goal was to present the experimental procedures of a protocol that a neuronavigation system can be used to improve the accuracy and precision of the stimulation of TA and SOL optimal spots. Numerous studies have examined the leg muscles CMR without using navigated TMS. Yet, using this protocol the CMR of both SOL and TA can be more accurately assessed.

Action: We have revised the last section of the fourth paragraph in the discussion to acknowledge the potential burden of using navigated TMS.

“Third, use of structural MRI of each subject with the neuronavigation system may not be feasible in all settings due to high cost of obtaining MRI and the neuronavigation system. However, certain neuronavigation systems including the one used in this protocol, can be used without subject’s MRI; but an average MRI is used. In this case, coil can be still precisely positioned over the stimulated site.”

2. Line 106: By "acknowledge the consent process", do you mean provide written informed consent?

Response: Yes, we meant to say written informed consent.

Action: We have revised this line to improve clarity.

“After answering any questions or concerns that subjects may have, ask subjects to acknowledge the consent process and sign the informed consent form.”

3. Line 160: It's easy to locate TA by palpation during dorsiflexion. By "tip of the fibula" do you mean head of the fibula?. And do you mean 1/3 from the top or the bottom of the imaginary line? It might be easier to recommend locating the muscle by palpation, immediately lateral to the tibial crest.

Response: We thank reviewer for pointing this out.

Action: We have revised this step to improve clarity.

“3.1.2 Attach electrodes bilaterally on TA. While in the standing position, ask subjects to lift their toes upwards and then place the electrode at the upper third of the line between the head of the fibula and medial malleolus (i.e., muscle belly immediately lateral to the tibial crest).”

4. How far apart should the recording electrodes be placed on the skin?

Response: We are using a single differential electrodes that the two discs housed in plastic case have distance of 20mm from each other.

5. Line 185: What is "underwrap"?

Response: We apologize for the confusion. It is a light foam wrap.

Action: We have revised this sentence to improve clarity.

“3.3 Secure all electrodes using light foam pre-wrap tape. Periodically throughout the experiment, check to ensure that electrodes are securely attached and that the signal has good quality.”

6. The criterion of 50 microvolts peak to peak seems fairly generous for 'rest' EMG. It's common elsewhere to use a criterion of < 0.01mV RMS calculated over at least 100 ms.

Response: We thank reviewer for this excellent methodological suggestion.

Action: We have added this suggestion as an alternative approach at the end of the third paragraph in the discussion.

“Lastly, the current protocol used the criterion of less than 50 μ V peak-to-peak amplitude to assess for baseline noise and for the “true” resting state. Discarding any EMG signal greater than 10 μ V (root mean square calculated over 100 ms) is an alternative approach.”

7. Line 190: Is 5 degrees from vertical noticeably reclined? This will seem essentially upright for many participants.

Response: The reviewer is correct; it is 5 degrees from vertical position. 5 degrees takes some pressure off of the hamstring and helps those with balance limitations. Much easier to maintain a neutral pelvis in this position. We have used this position in both neurologically intact (see Charalambous et al J Electromyography Kines 2018) and impaired adults (ongoing studies), and we had no complains. However, we do agree with the reviewer that this position might be upright and uncomfortable for some participants.

Action: To avoid any confusion, we have deleted the instruction about the chair adjustment.

“3.4 Seat the subject in a chair. To ensure consistent feet placement across subjects, secure both feet in walking boots (i.e., ankle foot orthosis) that allow the ankle ROM to be adjusted to a specific position and provide resistance during TVA testing. Adjust both hip and knee angles to avoid subject discomfort. Instruct the subject to keep still throughout the experiment. Use a forehead rest attached to the chair to keep subjects still during TMS application, if available.”

8. Line 204: Is the AFO expected to provide resistance against a maximal isometric dorsiflexion (or plantarflexion)? Is this sufficient and stable?

Response: We thank the reviewer for this excellent question. Yes, the specific AFO used in this study can provide resistance during both motions with adequate stability. During dorsiflexion, subject pull the foot against two straps whereas for plantarflexion subject push against the bottom of the AFO which was placed either against the wall or the floor. Yes, the AFO we used was able to provide sufficient stability.

Action: To improve clarity on this matter, we have revised the corresponding section.

“3.4 Seat the subject in a chair. To ensure consistent feet placement across subjects, secure both feet in walking boots (i.e., ankle foot orthosis) that allow the ankle ROM to be adjusted to a specific position and provide resistance during TVA testing. Adjust both hip and knee angles to avoid subject discomfort. Instruct the subject to keep still throughout the experiment. Use a forehead rest attached to the chair to keep subjects still during TMS application, if available.”

Reviewer #2:

1. Major problem is that authors claim that proposing protocol can be used for neurophysiological evaluation of the descending motor pathways in neurologically intact and IMPAIRED subjects. However, results are presented for non-injured subjects only. They have not examined this protocol in the impaired subjects. In the neurologically impaired, for example subjects with spinal cord injury, it may not be feasible to use the similar protocol because of the diminished conduction through cortico-spinal tract and thus transmission to corresponding motoneurons and then to leg muscles. So, authors must be careful and restrict wording to healthy subjects, as they are reporting.

Response: We thank reviewer for pointing this out. Though we present results from a neurologically intact subject and the protocol as written may imply that it can be used only in this population, we have used this protocol in individuals with neurologic impairments with success (between two dissertation projects and two funded NIH studies, totaling of approximately 240 TMS lower extremity sessions have been completed in individuals post-stroke). The manuscripts for these projects are currently work in progress.

2. Minor problems - it must be clearly indicated parameters of stimulation protocol, such as TMS frequency, coil used, the range of % of max intensity for inducing threshold MEP, etc.

Response: We thank the reviewer for these methodological questions. The TMS frequency was 0.25 Hz, the coil used was Magstim double cone coil (as listed in the material table), and the range of RMT was: SOL: left - 53 ± 13 , right - 52 ± 11 ; TA: left - 52 ± 12 ; 48 ± 11 % MSO. This information and data are included in our recent paper (Charalambous et al J Electromyography Kines 2018).

Reviewer #3:

1. The methods section states that "apply one TMS pulse on each spot of the grid (total 36 stimuli)". Given the known inter-trial variability of TMS-evoked MEP amplitudes, as well as additional variability caused by slight variations in coil orientation or location, the delivery of only 1 stimulus pulse at each grid site can be considered a disadvantage. Please provide your rationale for not delivering 3-5 pulses at each site. If this was to save experimental time, it should be clearly stated, with a caveat added regarding the limitation related to not delivering multiple pulses at each site (to enable the use of an averaged response for each site).

Response: This is an excellent issue that the reviewer has pointed out. We agree with the reviewer that more stimuli per spot would be better, yet we chose a single stimulus per spot for two reasons. First, the scope of this protocol was not to map the representations of TA and SOL, which would otherwise be ideal to employ the multiple stimuli per spot approach and using the average response per site. Second, this protocol was developed to assess SOL and TA CMR before and after either biomechanical assessment or a single day/session intervention. Therefore, we intended to administer the TMS assessment within an optimized time duration.

Action: We have acknowledged the methodological consideration in the third paragraph of the discussion section.

"Similarly, more than one stimulus per spot can be applied during the hot spot hunting (e.g., 2-5 stimuli/spot) compared to a single stimulus per spot used in this protocol. By applying more than one stimulus per spot, hot spot of each muscle might be more reliably determined. Recent study suggested that as few as two stimuli per spot might be sufficient for hot spot determination⁶⁷."

2. The methodological approach used here has several strengths. For example, the authors identify the hot spot of contralateral SOL and TA as the location in the grid with the largest amplitude and the shortest latency. The readers could benefit from inclusion of statements about whether other / previous studies have used both latency and amplitude to determine hotspot. If not, the innovation, pros, and cons of this methodological approach need to be emphasized.

Response: We appreciate the reviewer's comment. Though the majority of the studies define hot spot as the cortical location with the largest response (i.e., largest peak-to-peak amplitude), Rossini 1999 defines hot spot the "scalp position where the stimulus elicits the MEP of largest amplitude and minimal latency". Furthermore, recent paper (Kalliomeni 2015) suggested that the latency should be also considered for motor mapping.

Action: We have corrected the references cited. We also acknowledged this methodological consideration in the fifth paragraph of the discussion.

“Furthermore, rather than using the same hot spot for assessing a single muscle’s CMR, each muscle’s hot spot is determined using a standardized grid, which was laid over the leg cortical representation, and is defined as the spot with the largest amplitude and shortest latency⁴³.”

3. In contrast to the number of pulses delivered at each site (1), the adaptive algorithm for determining the motor threshold was run twice for each muscle. This adds significantly to data-collection time. Please provide justification for this aspect of the approach.

Response: We thank the reviewer for pointing this out. As others have previously reported (Silbert et al Clin Neurophys 2013), adaptive method is more efficient than the traditional relative frequency method even though both methods shared same accuracy. Therefore, we chose the adaptive method because it required fewer stimuli than the relative frequency method. Given that less than 20 stimuli required to assess the TA and SOL RMT (see Table 1 in Charalambous et al J Electromyography Kines 2018), we suggested running the RMT protocol twice to ensure accurate determination of the RMT. Each RMT determination takes less than 5 minutes. Also, pilot work demonstrated greater variance in lower extremity RMT, so we decided to perform the RMT determination procedure twice to reduce the effect of this variability.

4. In the methods, authors state that "in case any contralateral examined muscles are active before or after TMS, discard that stimulus and apply an extra single pulse again". Please clarify if the trials were discarded in case the targeted contralateral muscle (e.g. TA for trials involving delivery of TMS to the TA hotspot) was active or ANY contralateral muscle. As currently stated, the sentence is confusing. This is important because this will help readers ascertain whether the authors controlled for activation of the agonist or target muscle or the antagonist muscle as well.

Response: We thank the reviewer for pointing this out. For the rest condition, we expected all 4 muscles to be at rest, whereas for the TVA condition we expected only the target muscle to be activated. Therefore, if there was any activation during rest or TVA (any muscle other than the target muscle) that trial was discarded.

Action: We have revised the corresponding steps, the sections now read as follows:

“6.4.2 Prior to each stimulus, instruct the subject to stay still and relax the examined muscles bilaterally and monitor the activity of all muscles using a real time visual feedback displaying on a computer screen. In case that any muscle is active before or after TMS, discard that trial and apply an additional single pulse. Repeat until 10 waveforms for each contralateral examined muscle at rest have been collected.”

“6.5.4 Monitor the muscle activity of the active examined muscle and the remaining resting muscles using a real time visual feedback display on a computer screen. Discard that stimulus and apply an additional single pulse again in case that either the examined muscle’s activity is

either below or above the predetermined range or any other muscle is activated. Collect 10 trials while the examined muscle is activated at the predetermined range.”

5. Please provide the rationale for using 10 waveforms instead of a greater number (some studies have recommended 15 to 30) for the collection of suprathreshold MEPs for each condition.

Response: This is a great comment that the reviewer has pointed out. We reported 10 stimuli instead of higher number for two reasons. 1) 10 stimuli was number of stimuli used at the time that this protocol was developed (4-5 years ago), as well as those commonly reported previously. 2) The two papers that have showed that greater number of stimuli required for more accurate CMR assessment were published in 2016 and 2017. However we have acknowledged in the discussion section that more than ten stimuli should be considered. *“To assess the CMR of each muscle, 10 stimuli are applied on each hot spot during rest and TVA, yet recent reports have suggested that more than 10 stimuli should be used to assess reliably the CMR of a muscle^{65,66}.”*

6. Similar to detailed methods provided for other aspects of the protocol, additional details regarding methods and anatomical landmarks used to determine the MRI site used for positioning the center of the rectangular grid would be valuable to readers for future replication. Also, how did the authors handle any variability in sulcus anatomy when locating the grid?

Response: We thank the reviewer for this comment. The neuronavigation file of each subject was created by the same person who has extensive experience in cortical neuroanatomy. To ensure accurate grid placement across subjects the grid was placed based on two criteria. The middle row of the grid was placed perpendicular to the leg motor area while the medial column was placed parallel and adjacent to the medial wall of the hemisphere tested.

Action: We elaborated on this matter at section 2.5.

“2.2.2 Place a rectangular grid over leg motor cortical area at each hemisphere (see Figure 1B). Position the centered row of the grid at the center and over the gyrus of the leg motor cortical area where the corticospinal tracts that innervate leg motor pools originate³⁶. Position the medial column of the grid parallel and adjacent to the medial wall of the ipsilateral hemisphere.”

7. How was the window for determining peak to peak MEP (20 to 60 ms after TMS onset) determined? Is there a possibility that the window may need to be widened in patient populations (e.g. stroke, multiple sclerosis) beyond the 60-ms timeframe?

Response: This is an excellent question. Based on a work done in our lab in both neurologically intact and stroke-impaired subjects with varying height, we found that the latency of these two muscles was never less than 24 ms or greater than 50 ms. Therefore, we suggest the search window for MEP to be between 20 and 60 ms. Nevertheless, we agree

with the reviewer that neurological populations other than that we have considered, may have longer latencies outside of this window. We have incorporated this into the text.

Action: We have added a CAUTION line in 7.2 section.

“CAUTION: Though the MEP search window of 20-60 ms may work for neurologically intact subjects and people post-stroke, wider MEP search windows (e.g., 20-75 ms) might be required for other neurological populations (e.g., multiple sclerosis).”

8. The following finding is somewhat confusing and merits more detailed discussion and presentation in the paper: "As in rest, TMS over right TA and left SOL hot spots also elicited ipsilateral responses; those responses were present only in the ipsilateral TA; conversely, TMS over the right SOL and left TA hot spots elicited only contralateral MEPs."

The results and figures do not provide data to support this finding at a group level (only individual subject data are shown related to this point) or using statistics. Was this phenomena or observation consistent across all participants? Is there a potential methodological explanation for this finding? Does this relate to limb dominance? Please comment or clarify. If the finding is indeed variable, perhaps the emphasis on it can be reduced pending further confirmation using a larger sample study and support with statistical analysis?

Response: This is an excellent observation. In the results section we present data from a neurologically intact subject to show the feasibility of the protocol. We recently published a paper which used this protocol and presents bilateral TA and SOL data from 21 healthy subjects. We are currently working on manuscripts that used the same protocol in stroke patients. We did observe similar results in other subjects as well. The main rationale for this ipsilateral response is due to the location of the stimulated site. Given the size of the double cone coil and the location of the leg motor area, it is highly expected that ipsilateral responses might be elicited. Based on our data, it is unclear whether this finding is related to leg dominance.

9. The discussion/interpretation regarding ipsilateral MEPS (iMEPs) being ascribed to an oligo-synaptic pathway needs more clarification and justification with references. Could iMEPs be caused by activation of uncrossed corticospinal tract fibers too instead of brain-stem-mediated pathways?

Response: We thank the reviewer for pointing this out. It is possible that the uncrossed monosynaptic motor pathways might be stimulated. However, the ipsilateral latency should not be delayed and the amplitude should be similar to the contralateral MEP. Given the location of the motor leg area and the size of the double cone coil, any ipsilateral response in the lower extremity might be most likely due to the stimulation of the opposite hemisphere (i.e., stimulation of the crossed motor pathways of the opposite hemisphere). On the other hand, if the ipsilateral response is delayed and has much smaller amplitude than the contraletarel MEP, then this response might be due to oligosynaptic pathways. Definitely, future studies should further investigate this matter using the present protocol.

Action: To improve the clarity on this matter, we have revised the last section of the discussion's second paragraph.

"The second critical step is the bilateral assessment of each muscle. In contrast to upper extremity motor areas, the two leg motor areas are adjacent to each other, and when a pulse is applied over one area the opposite area might be stimulated due to current spread. Therefore, any ipsilateral response in either muscle may indicate either the presence of an iMEP (a potential proxy of cortico-reticulo-spinal pathway)⁵⁰ or just a direct stimulation of the opposite leg motor area. In the past, ipsilateral TA responses were reported, yet the stimulated site was based on anatomical landmark (10 and 15 mm posterior and lateral to vertex)⁶². Using this protocol, the hot spot of each muscle can be determined separately, and depending on the hot spot's location either contralateral or bilateral responses can be elicited (see Figures 3 and 4). Whether the bilateral response is a result of multiple descending pathways or just stimulation of a single pathway requires further investigation."

10. Additional data, summary descriptive statistics, and figures would be beneficial in the results section. For instance, was the hotspot location determined using MEP latency and MEP amplitude identical or was there a disparity in the location based on these 2 criteria for certain participants?

Response: We thank reviewer for this comment. Group data (descriptive statistics, reliability, etc.) have been presented in our recent paper (Charalambous et al J Electromyography Kines 2018). In this manuscript we chose to present representative data from a neurologically intact subject to demonstrate the pros and cons of this protocol. In the event that the largest amplitude and shortest latency is not at the same spot, then the spot with the largest amplitude should be defined as hot spot.

Action: We have added a CAUTION line in section 6.2.4.

"CAUTION: In an occasion that the largest amplitude and shortest latency are not at the same spot, define hot spot using the largest amplitude."

11. Participant characteristics and demographics would be useful.

Response: Thank you for the suggestion. Information on participant characteristics and demographics has been reported in our recent paper – Charalambous et al J Electromyography Kines 2018.

12. The average values of MEP amplitudes, latencies, and silent period for TA and Soleus are not provided in the results or figures. While the paper focuses on methodology, these data would serve as a useful reference for future studies using similar methodology in the same or other populations. Citations or references of these results from other papers from the authors' lab would also be useful.

Response: Thank you for the suggestion. This data can be found in our recent paper – Charalambous et al J Electromyography Kines 2018.