

Journal of Visualized Experiments

Fluorescent Silver Staining of Proteins in Polyacrylamide Gels

--Manuscript Draft--

Article Type:	Invited Methods Article - JoVE Produced Video
Manuscript Number:	JoVE58669R1
Full Title:	Fluorescent Silver Staining of Proteins in Polyacrylamide Gels
Keywords:	Silver staining; Fluorescent gel staining; Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis (SDS-PAGE); Aggregation induced emission (AIE); Silver ion sensor; Fluorogenic detection; Proteomics
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Additional Information:	
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TITLE:**Fluorescent Silver Staining of Proteins in Polyacrylamide Gels****AUTHORS AND AFFILIATIONS:**

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KEYWORDS:

Silver staining, fluorescent gel staining, sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE), aggregation-induced emission (AIE), silver ion sensor, fluorogenic detection, proteomics

SUMMARY:

Here, we describe a detailed protocol outlining a new fluorescent staining technique for total protein detection in polyacrylamide gels. The protocol utilizes a silver ion-specific fluorescence turn-on probe, which detects Ag⁺-protein complexes, and eliminates certain limitations of traditional chromogenic silver stains.

ABSTRACT:

Silver staining is a colorimetric technique widely used to visualize protein bands in polyacrylamide gels following sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The classic silver stains have certain drawbacks, such as high background staining, poor protein recovery, low reproducibility, a narrow linear dynamic range for quantification, and limited compatibility with mass spectrometry (MS). Now, with the use of a fluorogenic Ag⁺ probe, **TPE-4TA**, we developed a fluorescent silver staining method for the total protein visualization in polyacrylamide gels. This new stain avoids the troublesome silver reduction step in traditional silver stains. Moreover, the fluorescent silver stain demonstrates good reproducibility, sensitivity, and linear quantification in protein detection, making it a useful and practical protein gel stain.

INTRODUCTION:

Many staining methods have been used to visualize proteins following gel electrophoresis, for example using colorimetric dyes such as Coomassie Brilliant Blue, silver stain, fluorescence, or radioactive labeling^{1,2,3,4}. Silver staining is considered to be one of the most sensitive techniques for protein detection requiring simple and cheap reagents. It can be categorized into two major families: the ammoniacal silver stain and the silver nitrate stain^{5,6}. In the alkaline ammoniacal silver method, the silver-diamine complex is produced with ammonia and sodium hydroxide and reduced to metallic silver during development using an acidic formaldehyde solution. The stain accommodates efficiently for basic proteins but shows a compromised performance for acidic and neutral proteins and is, furthermore, limited to classical glycine and taurine electrophoretic systems. In comparison, the silver nitrate stains exploit the high bio-affinity of silver ions to protein, primarily the sulfhydryl and carboxyl groups from the side chains, and tend to stain acidic proteins more efficiently⁷. After silver ion binding, a developing solution (typically made of a metal carbonate solution containing formaldehyde and sodium thiosulphate) is applied to reduce silver ions to metallic silver grains, which build up a brown-to-dark color to visualize the protein bands.

Although silver staining has been well known for its versatility and high sensitivity since its development in the 1970s⁸, the method is frequently regarded as tricky. Silver-staining methods have time-restricted steps and show low reproducibility. Since the color of silver stain is usually not uniform and dependent on the reduction step, which is hard to control, the silver stain is not a quantitative method and, thus, not recommended for gel comparison study and protein quantification⁹. Methods optimized in sensitivity may utilize aldehydes which can also provide a more uniform staining¹⁰. However, this is at the expense of further downstream analysis due to the crosslinking of proteins by aldehydes. Fast protocols mostly combine or shorten steps to reduce time, compromising the reproducibility and uniformity of the stain⁵. As a result, there are numerous silver staining variants within protein gel staining, each optimized to suit certain requirements; for example, simplicity, sensitivity, or peptide recovery rate for downstream analysis. These attributes may also have an impact on each other, and satisfying all requirements in one protocol can be difficult.

In this work, we introduce a new fluorescent silver staining method for protein detection in polyacrylamide gel. In this method, we use a fluorogenic probe for silver ions, **TPE-4TA (Figure 1)**, to visualize the silver-impregnated proteins¹¹. **TPE-4TA** is designed by the aggregation-induced emission (AIE) principle. It is non-emissive when dissolved in aqueous solution, but is highly emissive in the presence of silver ions. By replacing the chromogenic development in traditional silver stains with a fluorogenic developing step, the fluorescent silver method enables the robust staining of total proteins with a reduced background.

Furthermore, the fluorescent silver stain showed a good dynamic linear range for protein quantification, which is comparable with the widely-used SYPRO Ruby stain and not achievable with traditional silver stains. The gel can be imaged on commonly used gel documentation systems with an ultraviolet lamp (excitation wavelength: 302/365-nm channel; emission: ~490 - 530 nm) at many biological labs.

PROTOCOL:

1. Preparation of the Gel

Note: The demonstration follows a standard protocol to prepare the gel for staining shortly after SDS-PAGE¹². In brief, the following steps describe the preparation of the samples and gel electrophoresis.

1.1. Perform SDS-PAGE with 4% - 12% Bis-Tris protein gels (1 mm, 15-well) using a mini gel tank filled with 2-(*N*-morpholino)ethanesulfonic acid (MES) buffer.

1.2. Dilute the samples with a mixture of distilled water, lithium dodecyl sulfate (LDS) buffer, and a sample-reducing agent.

1.3. Load the first lane with double the amount of stock (10 μ L), followed by the normal stock amount (5 μ L) and a series of twofold dilutions of the stock thereafter (13 dilutions, from 2x to 8192x).

1.4. Run the gel at a constant voltage of 200 V for 30 min.

2. Fixation of the Gel

2.1. After electrophoresis, submerge the gels in a 100-mL solution of 40% ethanol/10% acetic acid on an orbital shaker at 50 rpm at room temperature 2x (each for 30 min), or overnight at 4 °C.

2.2. Wash the gels 3 x 10 min each in ultrapure water in a clean container. The washing step is critical. If the acid from the fixing solution is not removed properly in the fluorogenic developing step, the excessive **TPE-4TA** will be activated by the acid and lead to strong background fluorescence in the gel.

3. Preparation of the AgNO₃ Solution and Silver Impregnation of the Gel

3.1. To make the AgNO₃ solution (0.0001%) for the impregnation, first, dissolve 0.01 g of AgNO₃ in 10 mL of ultrapure water to prepare a 0.1% AgNO₃ stock solution. Next, add 100 μ L of the 0.1% AgNO₃ stock solution into 100 mL of ultrapure water to make the working solution.

Note: The AgNO₃ solution should be stored in the dark before use.

3.2. Impregnate the gel with 100 mL of silver working solution for 1 h on an orbital shaker at 50 rpm in a sealed glass container. It is critical to perform the silver impregnation under a fume hood, protected from light with aluminum foil.

3.3. Wash the gel with ultrapure water (about 100 mL) in a clean container for 2 x 60 s.

4. Fluorogenic Development of the Gel

4.1. To prepare the dye stock solution (0.1 mM), add 3.0 mg of the **TPE-4TA** dye in 50 mL of ultrapure water. Sonicate the solution for 3 min and add a few drops of a basic solution (for example, 1 mM NaOH) to help dissolve the dye.

4.2. Check the fluorescence of the solution under a 365-nm UV lamp to ensure that the dye molecule is fully dissolved. Only weak or nonemissive solutions indicate full dissolution.

Note: The dye **TPE-4TA** was synthesized following the protocol recently reported by Xie *et al.*¹⁰ The **TPE-4TA** solution is very stable and can be kept in the dark for months.

4.3. To prepare 100 mL of the fluorogenic developing solution (10 μ M), add 10 mL of the **TPE-4TA** stock solution into 90 mL of ultrapure water. Check the pH of the solution using a pH meter and tune it to 7 - 9 using a sodium hydroxide solution (1 μ M).

4.4. Transfer the gel to a clean and sealable container with 100 mL of the fluorogenic developing solution. Make sure the gel is totally immersed in the solution. Seal the container. Cover it from light and shake it overnight on an orbital shaker at 50 rpm at room temperature. Alternatively, the incubation step can also be shortened to about ~2 h by preheating the AIE developing solution to 80 °C for staining and then leaving it at room temperature.

5. Destaining and Imaging

5.1. Transfer the gel to a clean container and destain it in 100 mL of 10% ethanol for 30 min.

Note: Water alone can be used to wash the gel. However, it is more time-efficient to use a 10% ethanol solution for the destaining. This will help to reduce the destaining process from hours to 30 min.

5.2. Rinse the gel in ultrapure water for 5 min.

5.3. Image the gel at the 365-nm channel or 302-nm channel by a gel documentation machine.

REPRESENTATIVE RESULTS:

The protein bands stained by the fluorescent silver stain exhibit an intense green fluorescence under a 365-nm UV lamp. All 14 protein bands (10 - 200 kDa), from top to bottom, were clearly visible, correlating well with the 14 red-colored ones stained by the SYPRO Ruby dye (**Figure 2**)¹⁰.

Regarding quantitative protein detection, the gels were imaged by a gel imaging system using automated procedures, and the images were analyzed and compared using the commercial software. This fluorescent silver staining method appears to have a high resolution. For some protein bands, the sensitivity of the fluorescent silver stain is also slightly better than that of the

fluorescent SYPRO Ruby stain. In particular, the performance of the fluorescent silver stain was improved for the ~10- to 40-kDa protein bands, indicating that the new method is particularly useful for the detection of proteins with low molecule weights. The data also suggest that the fluorescent silver stain gave a good and uniform linearity for all 14 proteins over a relatively broad range for protein quantification (**Figure 3**)¹¹.

In contrast to the silver nitrate stain which gave a high level of background signal and distorted peaks, the fluorescent silver stain detected the bands with a good contrast and uniform intensity distribution comparable to the SYPRO Ruby stain across all 14 proteins (**Figure 3**).

Note that insufficient washing after the gel fixation will result in high background staining (**Figure 4**). The residual acetic acid will light up the **TPE-4PA** and lead to a strong background.

FIGURE AND TABLE LEGENDS:

Figure 1: Chemical structure of TPE-4TA and its sensing mechanism to Ag⁺. X = H or Na⁺. Adapted from previous work, copyright 2018 WILEY-VCH¹¹.

Figure 2: Fluorescent silver staining. A) Summary of the protocol. **B)** A comparison of gels stained by the fluorescent silver stain (left) and SYPRO Ruby stain (right) imaged parallel with a handheld UV lamp under 365 nm irradiation. The proteins (10–200 kDa) were loaded by twofold serial dilution starting from 200–500 ng/band at the most left lane. Adapted from previous work, copyright 2018 WILEY-VCH¹¹.

Figure 3: Fluorescent intensity of proteins against protein amount, representative gel images, and signal profiles (from the fifth lane) of gels with three different stains. (A) Silver nitrate stain, **(B)** fluorescent silver stain (365-nm excitation), and **(C)** SYPRO Ruby stain. The first column of the figure shows the intensity of the stain for each band of the 14 proteins (10–200 kDa) against the amount of protein normalized to the first lane (starting on the left). The second column shows the representative gel images of the corresponding stain. The amount of protein loaded into the gel is described in **Figure 2**. Adapted from previous work, copyright 2018 WILEY-VCH¹¹.

Figure 4: Stained gel from a suboptimal experiment. In this experiment, the washing was insufficient after the fixation step. The residual acetic acid induced the aggregation of **TPE-4TA** and caused a strong background.

DISCUSSION:

Presented here is a novel fluorescent silver staining method for proteins in polyacrylamide gels. This strategy integrates conventional silver stains and fluorescent stains. The staining exploits the selective binding of silver ion to proteins as in other silver stains but employs a highly sensitive fluorogenic silver probe **TPE-4TA** to light up the silver bound proteins. Since the fluorogenic probe **TPE-4TA** can sense silver ions at a fairly low concentration in the nanomolar range¹¹, it enables a highly sensitive detection without any further demand of a reductive visualization step as needed in traditional silver stains. This will minimize run-to-run variations. Meanwhile, it avoids the use

of harsh chemicals, including formaldehyde and glutaraldehyde, which frequently interfere with peptide identification in MS analysis. Moreover, as **TPE-4TA** is AIE-active and free of any self-quenching problems, the densely accumulated dyes at a protein band can emit collectively in response to the number of dye molecules. This contributes to a wide linear dynamic range (LDR) for protein detection and a much brighter staining when compared with the SYPRO Ruby stain.

Compared with traditional silver staining, a significantly less amount of silver nitrate is required for this new staining technique—specifically, 0.0001% from 0.1% reported in commonly used silver nitrate staining protocols⁴. It can be hypothesized that the success of the new stain is purely down to the combination of the incredibly bright and sensitive fluorescent probe against a high contrast background. In comparison, reduction during the developing step in traditional silver stains requires a higher quantity of silver to produce dark visible bands, particularly when it occurs against a high background. In this method, the fluorogenic development time is longer than the traditional chemical development; however, this can be debated as a limitation or as an advantage. The gel can be left in the developing solution for long periods of time with no consequences, resulting in lower variations that may derive from the operator. Not only are there fewer steps required, but also, more pauses can be taken, which makes this protocol quite convenient. There is no risk of overstaining unlike in the traditional developing step which can overdevelop the gel, resulting in a dark background. This fluorescent silver stain is also expected to have a smaller interprotein variation, albeit a further evaluation of additional protein samples is warranted (**Figure 3**).

It is critical to follow the suggested silver nitrate concentration from the protocol; using a higher concentration does not result in better performance and sensitivity but, instead, produces a strong background fluorescence which can engulf the fluorescence of the bands. In terms of troubleshooting, loss in sensitivity and brightness may result from over-washing in the wash step prior to fluorogenic development or from insufficient silver impregnation. For this method, the fluorescence signal of the protein band is dependent on the number of protein-bound silver ions, which is essential for the formation of the fluorescent **TPE-4TA-Ag⁺** complex. The protein should be saturated with silver ions to maximize the potential of this staining method.

Finally, as mentioned previously, the fluorogenic dye for silver ion used in this method, **TPE-4TA**, is also pH sensitive. A low pH can protonate the free **TPE-4TA** to fluoresce in the gel, resulting in a high background stain. Therefore, improper wash steps, which leave residual acetic acid from the fixation solution, will greatly affect the stain quality. It is important to keep the gel relatively neutral in pH in the fluorogenic development step (**Figure 4**). It could also be helpful to adjust the pH of the **TPE-4TA** solution before staining. It is expected that this dye will be commercialized in the near future.

In summary, we described a practical protocol of the novel fluorescent silver stain for the visualization of total proteins in SDS-PAGE gel. The staining is straightforward, easy-to-use, cost-efficient, and has a good staining performance. This method can greatly facilitate the identification of the protein bands in gels and provides a useful tool for protein analysis.

ACKNOWLEDGMENTS:

The authors would like to thank Patrick Chan at the Ming Wai Lau Centre for Reparative Medicine, Karolinska Institutet, for his technical support. S. X. is grateful for the support from the Swedish Research Council (Grant No. 2017-06344).

DISCLOSURES:

A patent application on this fluorescent silver staining has been filed.

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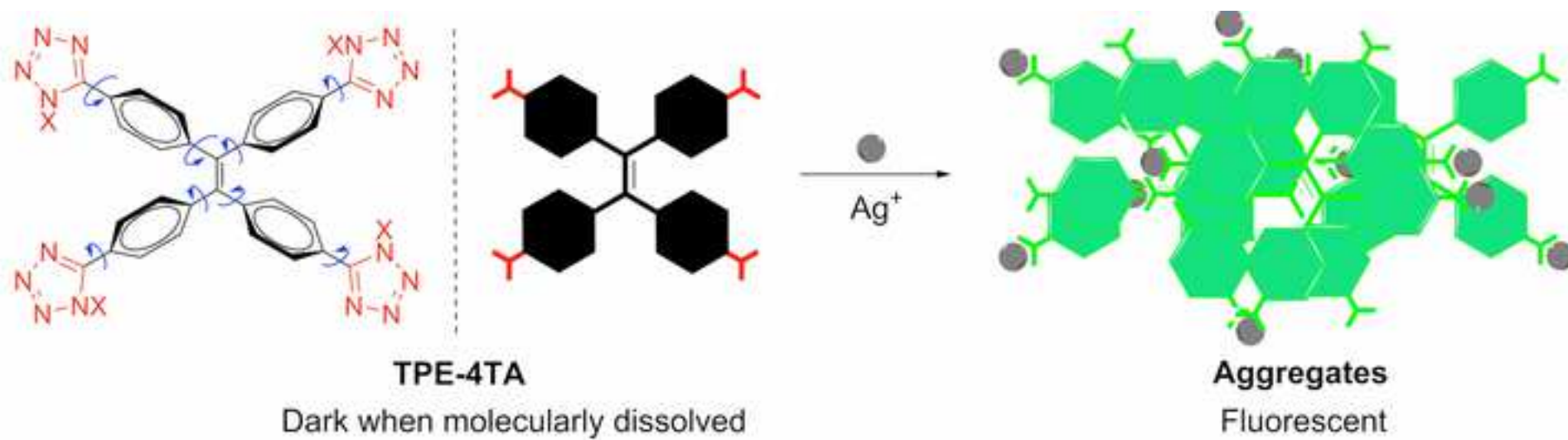


Figure 2

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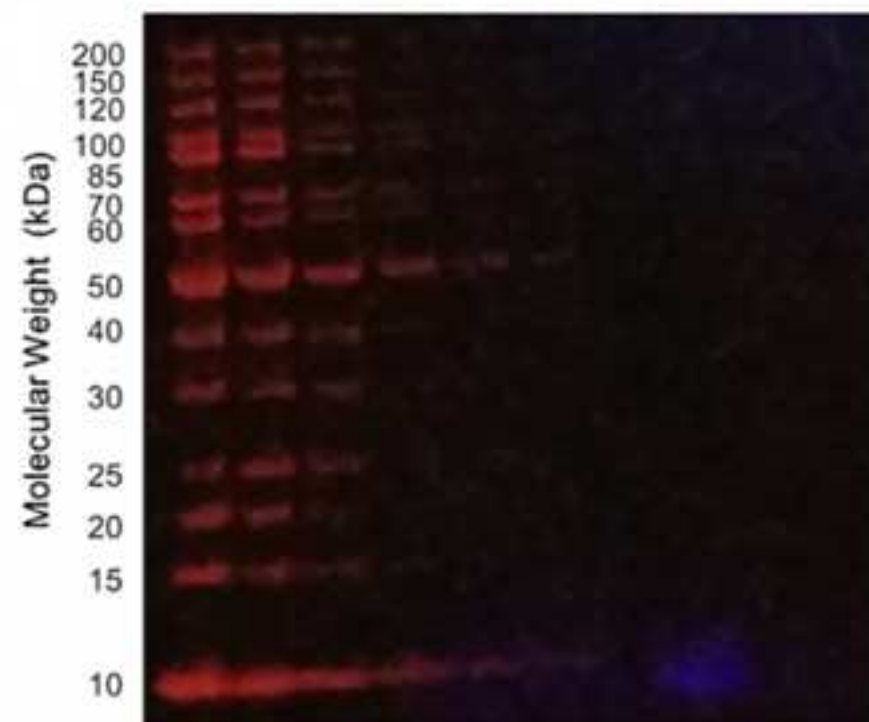
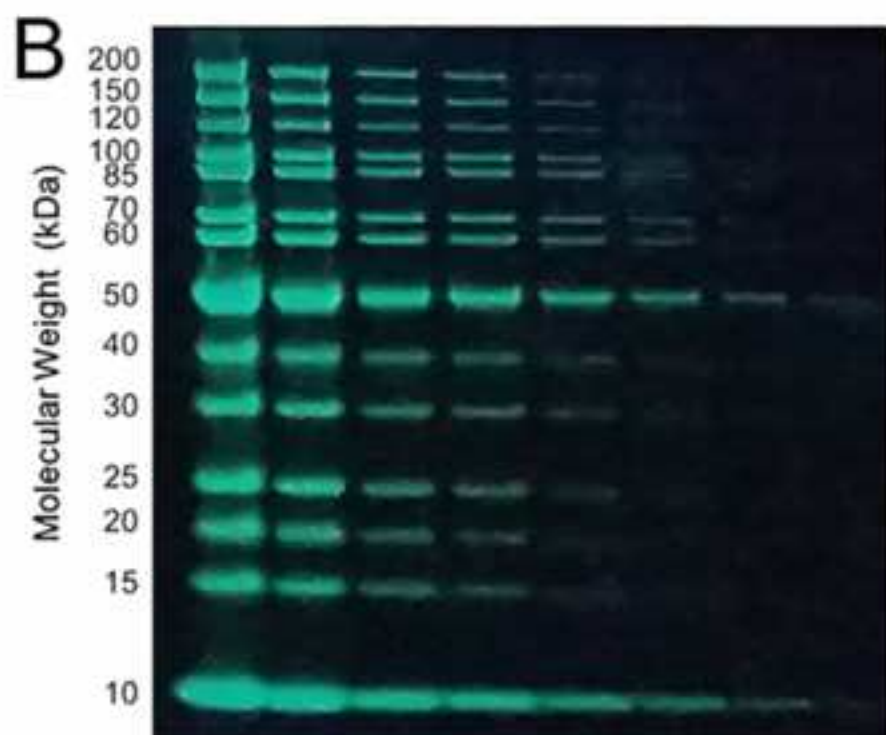


Figure 3

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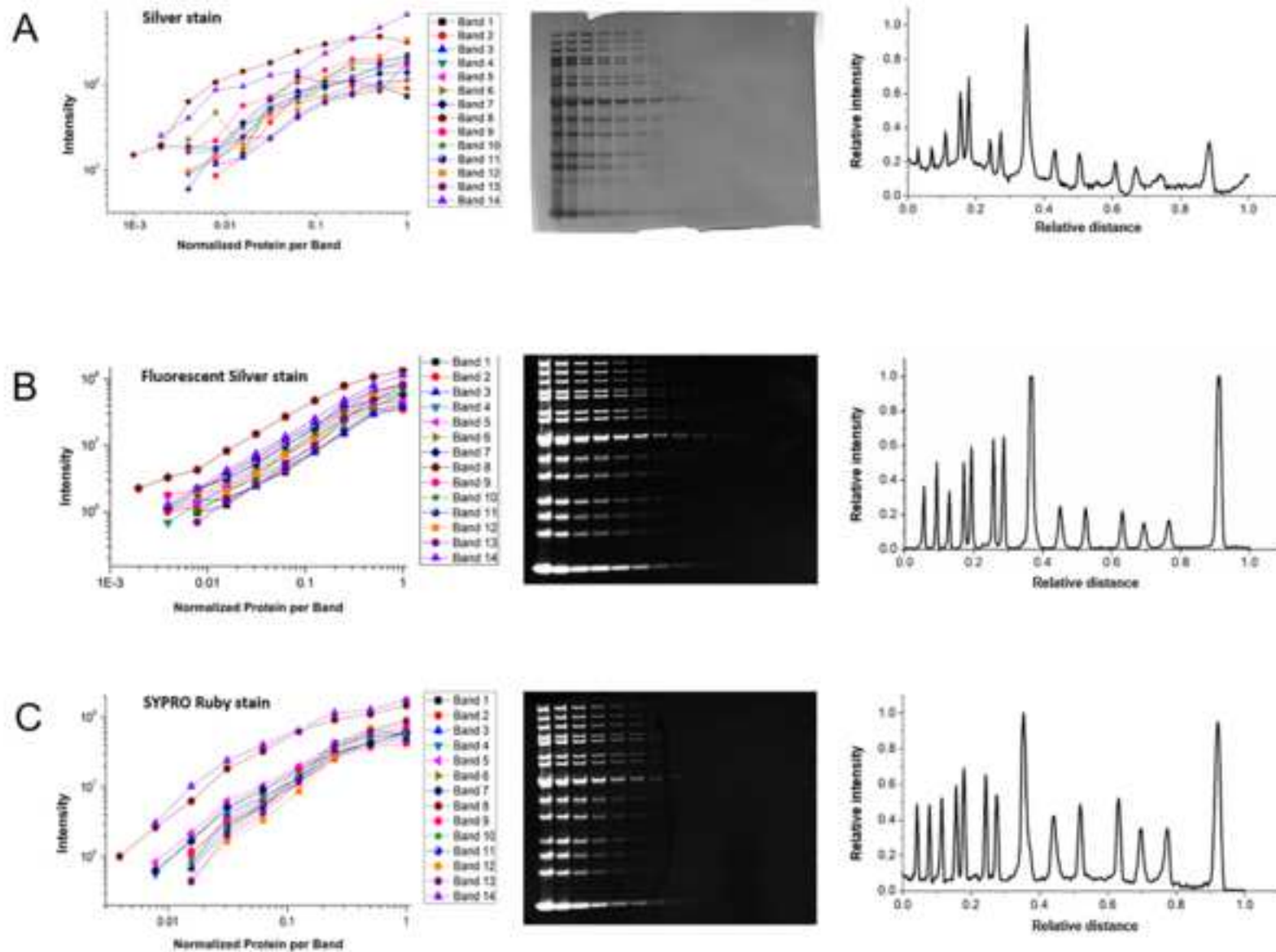


Figure 4

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Name of Material/ Equipment	Company	Catalog Number	Comments/Description
LDS Sample Buffer (4X)	Thermo Fisher Scientific	NP0007	Reagent
4-12% Bis-Tris Protein Gels, 1.0 mm, 15-well	Thermo Fisher Scientific	NP0323BOX	Precast gel
Sample Reducing Agent (10X)	Thermo Fisher Scientific	NP0004	Reagent
MES SDS Running Buffer (20X)	Thermo Fisher Scientific	NP0002	Reagent
Mini Gel Tank	Thermo Fisher Scientific	A25977	Equipment
300W Power Supply (230 VAC)	Thermo Fisher Scientific	PS0301	Equipment
Unstained Protein Ladder	Thermo Fisher Scientific	26614	Sample
Silver nitrate	Sigma-Aldrich	31630-25G-R	Reagent
Ethanol	Bragg and co.	42520J	Reagent
Acetic acid	J.T. Baker	103201A	Reagent
Milli-Q Synthesis A10	Merk	-	Provides 18.2 MΩ.cm water
gel documentation system (c600 model)	Azure biosystems	-	Equipment



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3rd Aug.2018

MS ID: **JoVE58669**
Subject: **Invited Video-Article**
MS Title: **Fluorescent Silver Staining of Proteins in Polyacrylamide Gels**
MS Authors: **Alex Wong, Sheng Xie, Ben Zhong Tang and Sijie Chen***

Dear Dr. Phillip Steindel,

Thank you very much for your email on 24 Jul. 2018 and for forwarding us the reviewers' comments and suggestions. We have revised our manuscript according to all the comments. All the modifications are tracked in the revised manuscript. Given below are our specific point-by-point responses to the reviewers' comments.

Response to your comments

1. *Please take this opportunity to thoroughly proofread the manuscript to ensure that there are no spelling or grammar issues.*

The manuscript has been proofread to correct any spelling or grammar issues. The spelling has been made consistent with standard American English.

2. *Please expand the Summary (10-50 words total) to briefly describe the applications of this protocol.*

We have expanded the summary to briefly describe the applications of this protocol as requested: "Here, we describe a detailed protocol which outlines a new fluorescent staining technique for total protein detection in polyacrylamide gels. The protocol utilizes a silver ion specific fluorescence turn-on probe, which detects the Ag⁺-protein complex. This technique makes use of the specific bio-affinity of silver ions to proteins, but eliminates certain limitations surrounding traditional silver stains."

3. *Please spell out each abbreviation the first time it is used.*

Correction have been made for abbreviations to be spelt out for its first use such as; Sodium Dodecyl Sulphate-Polyacrylamide-Gel Electrophoresis (SDS-PAGE), mass spectrometry (MS), Ultra-violet (UV) and Lithium Dodecyl Sulphate (LDS).

4. *JoVE cannot publish manuscripts containing commercial language. This includes trademark symbols (™), registered symbols (®), and company names before an instrument or reagent. Please remove all commercial language from your manuscript and use generic terms instead. All commercial products should be sufficiently referenced in the Table of Materials and Reagents. For example: Invitrogen, Thermo Fisher Scientific, Millipore, PowerEase, NUPAGE, Azurespot, etc.*

Thank you for the reminder, the trademark symbols and names have been removed from the manuscript and table of materials in the revision.

5. *Please revise the protocol text to avoid the use of any personal pronouns (e.g., "we", "you", "our" etc.).*

The personal pronouns have now been removed in the revised protocol. "The dye TPE-4TA was synthesized following the protocol that was recently reported.¹⁰"

6. *Please revise the protocol to contain only action items that direct the reader to do something (e.g., "Do this," "Ensure that," etc.). The actions should be described in the imperative tense in complete sentences wherever possible. Avoid usage of phrases such as "could be," "should be," and "would be" throughout the Protocol. Any text that cannot be written in the imperative tense may be added as a "Note." Please include all safety procedures and use of hoods, etc. Please move the discussion about the protocol to the Discussion.*

We have edited the words to the suggested action items in the revision and removed certain phrases as mentioned above.

7. *Lines 84-91: Please revise this paragraph to discrete numbered steps.*

We have revised the brief description of conditions for sample preparation and electrophoresis into discrete numbered steps.

8. *In the JoVE Protocol format, "Notes" should be concise and used sparingly. They should only be used to provide extraneous details, optional steps, or recommendations that are not critical to a step. Any text that provides details about how to perform a particular step should either be included in the step itself or added as a sub-step. Please consider moving some of the notes about the protocol to the discussion section.*

We have revised the 'Notes' as suggested.

9. 3.2: *Please specify how to adjust the pH of the solution.*

The revision now specifies how to check and adjust the pH of the solution: “To prepare 100 mL of the fluorogenic developing solution (10 μ M), add 10 mL of the **TPE-4TA** stock solution into 90 mL ultra-pure water. Check the pH of the solution using a pH meter and tune to 7-9 using acetic acid (1 mM) or sodium hydroxide (1 mM).”

10. 3.3: Please specify the incubation temperature.

The incubation temperature is at room temperature and is specified in the revision.

11. 4.2: What volume of water is used to rinse? How many times?

The gel is rinsed in 100 mL of water once, for a period of 5 min.

12. 4.3: Please add more details about how to image the gel.

The gel can be visualized with any UV lamp containing commercial gel documentation machine at the 302 nm and 365 nm channel. This has been specified more clearly in the revision.

13. Discussion: Please also discuss critical steps within the protocol and any limitations of the technique.

The critical step in the discussion includes the adjustment of pH and preventing the gel from acid exposure. Furthermore, an additional critical information within the protocol has been discussed, which is to follow the suggested silver nitrate concentration in the protocol as any higher may result in high background staining. Other limitations of the technique to be included in the revision is the fact that the dye is not widely commercially available and that the AIE development takes longer than a conventional chemical development.

14. References: Please do not abbreviate journal titles. Please include volume and issue numbers for all references.

The journal titles have now been corrected in the revision with the volume and issue numbers included.

15. Please follow the book citation example below to reformat book references: Kioh, L.G. et al. Physical Treatment in Psychiatry. Blackwell Scientific Pubs. Boston (1988).

Thank you for your suggestion, reference [6] now follows the example provided. Celis, J. E. et al. Cell Biology: A Laboratory Handbook. Elsevier Academic Press. Amsterdam (2006).

16. Please remove trademark (™) and registered (®) symbols from the Table of Equipment and Materials.

Thank you for the reminder, trademarks, symbols and commercial names have been removed from the Table of Equipment and Materials.

Response to the Comments and Suggestions of Reviewer 1

- 1) data suggest that apparently Sypro can stain proteins better than Silver staining, which is not normally true, of course it depends on the quality of silver staining, for example if silver stain is conducted by a nice protocol and properly or for example with Blue silver (see Electrophoresis. 2004 May;25(9):1327-33, that should be at least cited along with and its advantages/disadvantages compared to the method proposed) it is definitely more sensitive. The only real advantage of the proposed method, apart from its being fluorescent, is that it's fast and intrinsically displaying very low background.*

We fully agree with the comment regarding the SYPRO stain and silver stain. There are many silver staining protocols. In this study we used a simple silver nitrate stain, which has minimal steps and can be further modified for the silver impregnation step of the fluorescent silver stain. This serves as a control and benchmark for our fluorogenic silver stain to see how much the AIE dye can improve the performance in terms of sensitivity and contrast, when combined with the silver ions. This allows direct observation of how much the performance can improve with the AIE fluorophore without any extraneous influence from other reagents.

There are numerous silver staining protocols to compare. In particular when a sensitization step is applied, the detection of silver staining can be much improved. We also noticed the highly sensitive blue silver method, which is now cited in the revised version. Basically, we also prefer a silver nitrate staining protocol variant in the absence of an aldehyde sensitizer since it may lead to a stain compatible with mass spectroscopy.

- 2) Another drawback is the need for a particular device for the reading, anyway this should not be a problem.*

Yes, this is true for many fluorescence-based techniques. In our case, it is simple because many gel imagers have UV channels and thus can be used directly to read the gel.

Response to the Comments and Suggestions of Reviewer 2

- 1) The control method to which the new method has been compared, is not adequate. There are many other variants of silver staining that are one order of magnitude more sensitive than the one selected. It is as if someone describing a new staining method with a dye would compare it to old style Coomassie blue with alcohol-acid de-staining and not to modern colloidal Coomassie blue.*

Thank you for your comment. There are many silver staining protocols. In particular when a sensitization step is used, the quality can be improved highly. For the same protocol, the staining quality will also vary from operator to operator. There is no silver staining protocol

that can be claimed as the best for use as a standard silver staining method for comparison. Regarding the sensitivity, our method cannot exceed the detection limit of the best silver stain in this sample (the reported LOD is lower than many of the paper reporting highly sensitive silver staining method). That is why the SYPRO method is also used in parallel as a control group. SYPRO is a widely used fluorescent stain, and its performance has been fully compared with many silver stains.

In this work, we want to highlight the use of “fluorogenic development”, instead of the classic chemical development which reduces the silver ions. Taking this into account, the control method we have selected is fair to compare. Furthermore this allows us to understand the difference in the performance between our novel method and the basic fundamental silver staining method. We selected this protocol variant of the silver stain, specifically because it uses silver nitrate and avoids many harmful chemicals such as ammonia and glutaraldehyde without compromising MS compatibility.

Taking this into account, our selected silver protocol is a suitable control regarding the design of our staining strategy. Many other silver stains, in particular the sensitive ones, can be good alternatives to compare the detection limits in a fair way. Meanwhile, we adopt the standard SYPRO fluorescent stain as a suitable control to discuss about the sensitivity in the detection of proteins.

- 2) *The organic probe which is absolutely mandatory for the method is not widely available. Most of the end users do not have the chemical knowledge to prepare it or the money to buy a custom synthesis.*

At the time of writing, the AIE probe we have described is not yet widely available, however the probe is patented and will be commercialized very soon.

Please let me know if further information is required.

Yours sincerely,



Sijie Chen, Ph.D.
Assistant Professor
Karolinska Institutet

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Jun 25, 2018

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Estimated size of new article (pages)	6
Requestor Location	Ming Wai Lau Centre for Reparative Medicine, Karolinska Institutet, Hong Kong Units 608-613 Building 15 Science Park 15W Science Park W Ave Sha Tin, 000 Hong Kong

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