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Functionalization and dispersion of carbon nanomaterials using an environmentally friendly ultrasonicated-ozonolysis process --Manuscript Draft--

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Corresponding Author:	Andrew Rider Defence Science and Technology Group Fishermans Bend, VIC AUSTRALIA
Corresponding Author Secondary Information:	
Corresponding Author E-Mail:	andrew.rider@dsto.defence.gov.au;eudora.yeo@dsto.defence.gov.au
Corresponding Author's Institution:	Defence Science and Technology Group
Corresponding Author's Secondary Institution:	
First Author:	Eudora Yeo
First Author Secondary Information:	
Other Authors:	Eudora Yeo
	Gary Mathys
	Narelle Brack
	Erik Thostenson
Order of Authors Secondary Information:	
Abstract:	Functionalization of carbon nanomaterials is often a critical step that facilitates their integration into larger material systems and devices. In the as-received form, carbon nanomaterials, such as carbon nanotubes (CNTs) or graphene nanoplatelets (GNPs), may contain large agglomerates. Both agglomerates and impurities will diminish the benefits of the unique electrical and mechanical properties offered when CNTs or GNPs are incorporated into polymers or composite material systems. Whilst a variety of methods exist to functionalize carbon nanomaterials and create stable dispersions, a lot of the processes use harsh chemicals, organic solvents or surfactants, which are environmentally unfriendly and may increase the processing burden when isolating the nanomaterials for subsequent use. The current research details the use of an alternative, environmentally friendly technique for functionalizing CNTs and GNPs that produces stable aqueous dispersions free of harmful chemicals. Both CNTs and GNPs can be added to water at concentrations up to 5 g/L and recirculated through a high powered ultrasonic cell. Simultaneous injection of ozone into the cell progressively oxidizes the carbon nanomaterials and the combined ultrasonication breaks down agglomerates and immediately exposes fresh material for functionalization. The prepared dispersions are ideally suited for deposition of thin films onto solid substrates using electrophoretic deposition (EPD). CNTs and GNPs from the aqueous dispersions can be readily used to coat carbon and glass reinforcing fibers using EPD for the preparation of hierarchical composite materials.
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**Australian Government****Department of Defence**
Science and TechnologyDefence Science and Technology Group
506 Lorimer Street
Fishermans Bend VIC 3207
Australia
+61 3 9626 7172, fax +61 3 9626 7174
eudora.yeo@defence.gov.au

File Reference - Objective ID: AV12752438

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1 Alewife Centre #200
Cambridge, MA 02140
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1. In most technical publications, the experimental method is only briefly mentioned. It can be difficult for other researchers to understand how to undertake novel processes. This work is ideally suited to JoVE's multimedia format as it will demonstrate to the viewer exactly how to replicate our experiments.
2. All the authors were involved in various aspects of the experimental or analytical process, with the manuscript written by Eudora Yeo and Andrew Rider.
3. We were first invited to publish with JoVE by Alison Hamlin. Later, Neelima Chandrayan took over as our editorial contact, and most recently Benjamin Werth has assisted with submission enquiries.
4. Suggested peer reviewers:
 - Dr Peter Kappen, Scientist – XAS, Australian Synchrotron,
peter.kappen@synchrotron.org.au
 - Dr Michelle Spencer, Senior Lecturer, RMIT University,
michelle.spencer@rmit.edu.au
 - Dr Mathew Joosten, Lecturer, Deakin University,
mathew.joosten@deakin.edu.au
 - Sandeep Tamraker, Center for Composite Materials (UD-CCM), Department of Civil & Environmental Engineering, University of Delaware,
tamrakar@udel.edu
 - Associate Professor Paul Pigram, Director, Centre for Materials and Surface Science, Department of Chemistry and Physics, La Trobe University,
p.pigram@latrobe.edu.au
 - Dr Grant McAdam, Senior Research Scientist, Defence Science and Technology Group, grant.mcadam@dsto.defence.gov.au

A handwritten signature in black ink, appearing to read 'Eudora Yeo'.

Eudora Sia Ying Yeo
Materials Engineer

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TITLE:

Functionalization and dispersion of carbon nanomaterials using an environmentally friendly ultrasonicated ozonolysis process

AUTHORS:

Yeo, Eudora S Y
Aerospace Division
Defence Science and Technology Group
Melbourne, Victoria 3207, Australia
eudora.yeo@dsto.defence.gov.au

Mathys, Gary I
Maritime Division
Defence Science and Technology Group
Melbourne, Victoria 3207, Australia
gary.mathys@dsto.defence.gov.au

Brack, Narelle
Department of Chemistry and Physics,
La Trobe Institute for Molecular Science, La Trobe University
Melbourne, Victoria 3086, Australia
n.brack@latrobe.edu.au

Thostenson, Erik T
Department of Mechanical Engineering, Department of Materials Science and Engineering, and
Center for Composite Materials,
University of Delaware,
Newark, DE 19716, USA
thosten@udel.edu

Rider, Andrew N
Aerospace Division
Defence Science and Technology Group
Melbourne, Victoria 3207, Australia
andrew.rider@dsto.defence.gov.au
pH: 613 9626 7393

CORRESPONDING AUTHOR:

Andrew N. Rider (andrew.rider@dsto.defence.gov.au)

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SHORT ABSTRACT:

Here, a novel method for the functionalization and stable dispersion of carbon nanomaterials in aqueous environments is described. Ozone is injected directly into an aqueous dispersion of carbon nanomaterial that is continuously recirculated through a high-powered ultrasonic cell.

LONG ABSTRACT:

Functionalization of carbon nanomaterials is often a critical step that facilitates their integration into larger material systems and devices. In the as-received form, carbon nanomaterials, such as carbon nanotubes (CNTs) or graphene nanoplatelets (GNPs), may contain large agglomerates. Both agglomerates and impurities will diminish the benefits of the unique electrical and mechanical properties offered when CNTs or GNPs are incorporated into polymers or composite material systems. Whilst a variety of methods exist to functionalize carbon nanomaterials and to create stable dispersions, many the processes use harsh chemicals, organic solvents, or surfactants, which are environmentally unfriendly and may increase the processing burden when isolating the nanomaterials for subsequent use. The current research details the use of an alternative, environmentally friendly technique for functionalizing CNTs and GNPs. It produces stable, aqueous dispersions free of harmful chemicals. Both CNTs and GNPs can be added to water at concentrations up to 5 g/L and can be recirculated through a high-powered ultrasonic cell. The simultaneous injection of ozone into the cell progressively oxidizes the carbon nanomaterials, and the combined ultrasonication breaks down agglomerates and immediately exposes fresh material for functionalization. The prepared dispersions are ideally suited for the deposition of thin films onto solid substrates using electrophoretic deposition (EPD). CNTs and GNPs from the aqueous dispersions can be readily used to coat carbon- and glass-reinforcing fibers using EPD for the preparation of hierarchical composite materials.

INTRODUCTION:

The use of carbon nanomaterials to modify polymeric and composite systems has seen intensive research interest over the past 20 years. Recent reviews on both the use of carbon nanotubes¹ (CNTs) and graphene nanoplatelets² (GNPs) provide an indication of the breadth of research. The high specific stiffness and strength of CNTs and GNPs, as well as their high intrinsic electrical conductivity, make the materials ideally suited for incorporation into polymeric systems to enhance both the mechanical and electrical performance of the nanocomposite materials. CNTs and GNPs have also been used for the development of hierarchical composite structures by using the carbon nanomaterials to modify both fiber interfacial adhesion and matrix stiffness^{3,4}.

The homogeneous dispersion of carbon nanomaterials into polymeric systems often requires processing steps, which chemically alter the nanomaterials to improve the chemical compatibility with the polymer matrix, remove impurities, and reduce or remove agglomerates from the as-received materials. A variety of methods to chemically modify carbon nanomaterials are available and may include wet chemical oxidation using strong acids^{5,6}, modification with surfactants⁷, electrochemical intercalation and exfoliation⁸, or dry chemical processing using plasma-based processes⁹.

The use of strong acids in the oxidation step of CNTs introduces oxygen functional groups and

removes impurities. However, it has the disadvantage of significantly reducing the CNT length, introducing damage to the CNT outer walls and using dangerous chemicals, which need to be isolated from the treated material for further processing¹⁰. The use of surfactants combined with ultrasonication offers a less aggressive method to prepare stable dispersions, but the surfactant is often difficult to remove from the treated material and may not be compatible with the polymer being used to prepare the nanocomposite materials^{1,11}. The strength of the chemical interaction between the surfactant molecule and CNT or GNP may also be insufficient for mechanical applications. Dry plasma treatment processes conducted under atmospheric conditions may be suitable for functionalizing arrays of CNTs, present on fiber or planar surfaces, used to prepare hierarchical composites⁹. However, the atmospheric plasma is more difficult to apply to dry powders and does not address the problems with agglomerates present in as-manufactured raw carbon nanomaterials.

In the present work, we introduce a detailed description of the ultrasonicated-ozonolysis (USO) method that we have previously applied to carbon nanomaterials¹²⁻¹⁴. The USO process is used to prepare stable, aqueous dispersions that are suitable for electrophoretically depositing (EPD) both CNTs and GNPs onto carbon and glass fibers. Examples of EPD using USO-functionalized CNTs to deposit thin, uniform films onto stainless steel and carbon fabric substrates will be provided. Methods and typical results used to chemically characterize the functionalized CNTs and GNPs will also be provided, using both X-ray photoelectron spectroscopy (XPS) and Raman spectroscopy. A brief discussion of the characterization results in comparison with other functionalization techniques will be provided.

Work Health and Safety Notice

The effects of exposure to nanoparticles such as CNTs, on human health are not well understood. It is recommended that special measures be taken to minimize exposure to and avoid environmental contamination with CNT powders. Suggested hazard isolation measures include working within a HEPA filter-equipped fume cupboard and/or glove box. Occupational hygiene measures include wearing protective clothing and two layers of gloves and performing regular cleaning of surfaces using damp paper towels or a vacuum cleaner with a HEPA filter to remove stray CNT powders. Contaminated articles should be bagged for hazardous waste disposal.

Exposure to ozone can irritate the eyes, lungs, and respiratory system, and at higher concentrations may cause lung damage. It is recommended that measures be taken to minimize personal and environmental exposure to generated ozone gas. Isolation measures include working within a fume cupboard. As the return air stream will contain unused ozone, it should be passed through an ozone destruct unit before being releasing into the atmosphere. Dispersions that have had ozone bubbled through them will contain some dissolved ozone. After ozonolysis operations, allow the dispersions to sit for 1 h before undertaking further processing so that the ozone can undergo natural decomposition.

PROTOCOL:

1. Functionalization of CNTs and GNPs by ultrasonic ozonolysis

1.1) Weigh the nanomaterials in a glove box inside a HEPA filter-equipped fume cupboard. Weigh the desired quantity of nanomaterials into a beaker. Transfer to a bottle and add ultrapure water to make a concentration of 1 g/L.

1.2) Seal the bottle with a lid. Ultrasonicate in a standard ultrasonic bath (see the table of materials; frequency: $\sim 43 \pm 2$ kHz; power: 60 W) to disperse the CNTs or GNPs.

Note: CAUTION. See the Work Health and Safety Notice above.

1.3) Carefully pour the nanomaterial suspension into a reactor flask containing a magnetic stirrer bar. Place this on a stirring plate and turn on the magnetic stirrer by adjusting the dial to prevent the CNTs from settling out of suspension.

1.4) Assemble the ultrasonic horn into the ultrasonic cell and check that the horn tip is in good condition. See Figure 1.

[Insert Figure 1 here]

1.5) Connect the silicone tubing to the reactor, the ultrasonic cell, and the peristaltic pump. Refer to Figure 1.

1.6) Insert the ozone bubbler and connect it to the ozone generator.

Note: CAUTION. See the Work Health and Safety Notice above.

1.7) Flip the switch to turn on the cooling water recirculation unit to cool the ozone reactor and ultrasonic cell. Ensure that the cooling water recirculation system is flowing through the ultrasonic cell and carbon nanomaterial dispersion bath.

1.7.1) Use a thermocouple connected to the ultrasonic horn control unit to measure the reactor temperature. Wait until the system has cooled to 5 °C.

1.8) Place the reactor flask with the CNTs in position and insert the ozone tube and the ultrasonic horn. Place the ozone diffuser on the bottom of the reactor flask, with the ultrasonic tip submerged at least 25 mm below the surface of the dispersion.

1.8.1) Connect the outlet tube from the reactor to the ozone destruct unit using the same silicone tubing as in the peristaltic pumping operation.

1.9) Turn on the oxygen supply to the ozone generator and adjust the valve to achieve a 0.5-L/min flow rate through the ozone generator.

1.10) Turn on the ozone generator by flipping the switch. Allow it to run for 30-60 min prior to

starting the pumping or the ultrasonication process.

1.11) Turn on the peristaltic pump, adjust it to 0.67 Hz by turning the pump dial to a setting of approximately 5 or 6, and ensure that the CNT dispersion flows evenly through the cell.

1.11.1) Turn on the ultrasonic horn and adjust the power to 60 W by entering this value into the control module under the manual operation menu.

1.11.2) At initial startup, ramp up the amplitude setting slowly to achieve the desired power output of 60 W.

Note: The ultrasonic horn operates at 20 kHz. The amplitude setting and power setting are usually of similar values, but upon initial startup, the amplitude setting may need to be slowly increased to achieve the desired power output of 60 W.

1.12) Observe the sonication process for at least 30 min to ensure stable pumping and ultrasonic horn operation.

1.13) Monitor the flexible tubing in direct contact with the peristaltic pump mechanism for wear. Adjust the tube position at least every 2 h to ensure that the tubing maintains integrity. Replace the tubing for each separate carbon nanomaterial processing run.

1.14) When the desired USO processing time has elapsed, press the switches to turn off the ozone generator, sonicator, and pump. Maintain stirring until the carbon nanomaterial dispersion is ready to be transferred to a sealed container for later use in electrophoretic deposition.

1.14.1) Wait around 1 h before transferring the dispersion to allow the ozone in solution to decompose.

1.15) If electrophoresis is not performed within 24 h of USO processing, sonicate the nanomaterial dispersion again to minimize the number of particles settling out of dispersion. Insert the sonic horn into the bottle of the carbon nanomaterial dispersion and sonicate (as above) for 30 min prior to use.

2. Electrophoresis

2.1) Prepare three stainless-steel electrodes, 60 mm (L) x 25 mm (W), from 0.2-mm sheet material.

2.1.1) Abrade the electrodes using P1000 aluminium oxide sandpaper with ultrapure water as a lubricant. Following the abrasion, place the electrodes in the ultrasonic cleaning bath and clean them for 10 min in ultrapure water.

2.2) Place the electrode to be used for anodic deposition in a clean oven at 100 °C and dry it for 10 min. Move the electrode to a desiccator and allow it to cool. Weigh the electrode on a four-figure analytical balance.

2.3) Record the weight of the electrode to four decimal places as the uncoated weight in grams. Dry the remaining two electrodes and store them in the desiccator for use as cathodes for deposition.

2.4) Using a suitable clamping arrangement (see Figure 2), such as a small vice and 10-mm non-conductive plastic spacers, assemble the electrodes, with the outer electrodes being used as the cathodes and the inner electrode as the anode.

[Place Figure 2 here]

2.5) Transfer 35 mL of carbon nanomaterial dispersion to a 50-mL beaker.

2.6) Place the stainless-steel electrode fixture onto a retort stand and slowly lower the electrodes to the full depth of beaker. Attach the positive terminal of the DC power supply to the central stainless-steel anode. Attach the negative terminal of the DC power supply to the outer cathode. Connect the two outer cathodes using a lead with alligator clips.

2.7) If accurate current measurements are required, connect a multimeter in series with the power supply unit and electrodes to enable monitoring during deposition.

2.8) Manually adjust the current setting on the DC power supply to the maximum current (nominally, 2 A) and then the voltage to the required value for the studies, ensuring that the power supply output is turned off. Prepare a stopwatch and switch on the power supply for the required coating duration.

2.9) For a typical coating experiment, adjust the voltage to 10 V, with the coating times ranging from 1 to 15 min. Switch off the DC power supply and then slowly raise the electrodes out of the dispersion, ensuring that the film is not disturbed.

2.10) Disconnect the terminals from the electrodes and slowly invert the electrodes to a horizontal orientation to allow the film on the anode to dry evenly. After the moisture has evaporated from the film at room temperature, disassemble the electrode fixture and place the anode in the oven at 100 °C to dry for 1 h.

2.10.1) Clamp and hang the anode in the oven to ensure that the film does not come into direct contact with any surfaces.

2.11) Place the film-coated anode in the desiccator and allow it to cool, then weigh it on a four-figure analytical balance. Record the weight to four decimal places. Subtract the weight of the uncoated anode to determine the mass of the deposited film.

2.12) Photograph the coated anode. Use a suitable image processing package to accurately measure the area of the deposited film. Use the mass and area to record the areal density of the film in mg/cm^2 .

2.13) Repeat the coating and weighing steps for each deposition time required to determine the rate of deposition of the carbon nanomaterial onto the stainless steel for the specific field strengths being used.

3. Chemical characterization – X-ray photoelectron spectroscopy (XPS¹⁵)

3.1) To prepare the samples for XPS analysis, drop-cast the aqueous CNT dispersions from the original 1 g/L dispersion.

3.1.1) Using a pipette, deposit a droplet of the dispersion on a stainless-steel disk and it allow to dry at room temperature. Repeat the process until a uniformly thick CNT coating of approximately 2 μm in thickness is observed on the disk. Mount the stainless-steel disk onto sample stubs using double-sided conductive tape.

3.2) Insert the samples into the sample entry chamber and pump down to a vacuum of 5×10^{-7} Torr prior to transferring it to the main analysis chamber. Wait for the main chamber pressure to reach 5×10^{-9} Torr before positioning the sample and undertaking the analysis.

3.3) To follow the typical operating conditions for the XPS measurements, use a monochromatic Al $K_{\alpha,1}$ X-ray source operating at 150 W to illuminate an area of 700 μm x 300 μm . Detect photoelectrons at a 90° take-off angle with respect to the sample surface.

3.3.1) Calibrate the spectrometer energy scale using the Au 4f_{7/2} photoelectron peak at a binding energy of 83.98 eV.

Note: The energy scale is adjusted in the instrument control software during the initial commissioning of the instrument. It is regularly checked following any maintenance operations by the spectroscopist in charge of the equipment and should not be altered by users.

3.4) Monitor the initial sample charge by positioning a 30-eV window over the C 1s region. Perform charge compensation by adjusting the electron flood gun parameters systematically to position the C 1s peak at 284.6 eV and to minimize the peak width at half the maximum peak intensity (FWHM).

3.5) Conduct survey scans at 160 eV pass energy and 0.5 eV steps, with a dwell time of 0.1 s, in fixed analyzer transmission mode to identify the surface elemental species. Acquire region spectra at a 20-eV pass energy using 0.05-eV steps and a 0.2-s dwell. Use between 4 and 8 sweeps to improve the signal-to-noise ratio.

3.5.1) To quantify the spectra, use a Shirley background fit and sensitivity factors from a suitable elemental library¹⁶.

3.6) Deconvolve C 1s high-resolution X-ray photoelectron spectra to determine the different carbon-oxygen species present with USO treatment time¹⁵.

3.6.1) To accurately deconvolute the C 1s peak, perform an initial peak fit to the shape of graphitic carbon to ensure that the peak shape due to the sp^2 bonding and the contributions from the low-energy π -type shake-up satellites are maintained¹⁷. For all peak fitting, constrain the component binding energies and FWHM by ± 0.1 eV and ± 0.2 eV, respectively, and use a Gaussian/Lorentzian ratio of 30.

3.6.2) Fit the graphite peak shape using six separate peaks. Maintain at fixed ratios the areas of each peak relative to the main graphite peak at 284.6 eV to maintain a consistent shape due to the graphitic component in the processed nanomaterial^{15,9,14}.

4. Structural characterization – Raman spectroscopy¹⁸

4.1) Prepare the nanomaterials for Raman spectroscopic characterization by diluting the original dispersion with deionized water to achieve a 0.1 g/L concentration. Pipette 0.05 mL of dispersion onto a suitable polished-gold substrate. Allow the droplet to evaporate at room temperature to produce a thin carbon film on which to focus the Raman laser spot.

Note: The size of the gold substrate is incidental to the result when a large substrate is used, multiple specimens can be deposited on a single slide.

4.2) Undertake Raman measurements with a 50x magnification objective lens using a dispersive confocal microscope and a 532-nm laser. Acquire the spectra between 3,500 and 50 cm^{-1} at a resolution of 2 cm^{-1} . Use a 50- μm pinhole aperture with a laser energy between 1 and 10 mW, using the maximum energy possible such that no sample degradation occurs during collection.

4.2.1) Collect the spectra by accumulating between 10 and 50 scans and using an exposure time between 2 and 5 s for each individual scan to achieve a reasonable signal-to-noise ratio.

5. Film morphology – Scanning electron microscopy (SEM)

5.1) For the SEM analysis of functionalized versus unfunctionalized CNT/GNPs, dilute a droplet of each dispersion to 0.1 g/L using deionized water. Deposit it onto an aluminium SEM stub and allow to dry in air overnight.

5.2) For EPD films deposited onto the stainless-steel electrodes, cut the electrodes into 1 cm x 1 cm squares and mount them onto sample stubs using double-sided conductive tape. Paint a small line of silver dag (*i.e.*, conductive adhesive/paint) from the sample stub onto the top surface of the specimen to improve the conductive path. Dry the sample under a heater lamp for at least

15 min.

5.3) Place the mounted specimen in the sputter coater and give it a conductive iridium coating approximately 1.3-1.5 nm thick¹⁹.

5.3.1) On the thickness measurement unit, set the material density to the appropriate level, which is 22.56 g/cm³ for iridium. Adjust the target thickness setting to 1.0 nm to give a thickness of approximately 1.3-1.5 nm when accounting for thermal drift during the coating process.

5.3.2) Set the coater unit to allow the sputter coater operation to be controlled by the thickness measurement unit. Select the desired power output, which should be 80 mA for a high-resolution coating¹⁹.

5.3.3) Zero the coating thickness gauge on the thickness control unit and press the “Cycle” button to commence the automated chamber evacuation, argon bleed/flush and coating sequence. When the power has reached the desired level, move aside the target shield to allow the coating to commence. Rotate and tilt the stage during the coating to facilitate an even coating on all sides.

5.4) Examine the nanomaterials using a field-emission SEM with an in-lens detector. Use a working distance of approximately 3 mm and an accelerating voltage of 3.0 kV.

REPRESENTATIVE RESULTS:

Figure 3 shows the XPS wide-scan characterization of CNTs that had undergone USO treatment. CNTs that had not undergone USO show almost no oxygen content. As the USO time increases, the surface oxygen level increases. Figure 4 charts the oxygen-to-carbon ratio increases as a function of USO time. Table 1 shows the deconvoluted carbon species atomic concentrations of GNP treated with USO. The peak fitting used a combination of constrained peaks, represented by Gr1 to Gr6, to represent the inherent peak shape due to graphite and the associated energy loss features¹⁷. The oxygen-containing species were then added, and a cross-correlation with the C 1s peak fitting data and the elemental C and O percentages was made to ensure that sensible peak fitting results were achieved. The C-O, C=O, and COO species are at insignificant levels without USO treatment but increased markedly with 12 h of USO.

Raman spectra of CNTs treated by USO are shown in Figure 5. The intensity of the D band at 1,346 cm⁻¹ denotes the presence of defects in the CNTs²⁰ and indicates that defects already existed in the CNTs prior to the USO treatment. With increasing USO treatment time, there is a shift in the G band, from 1,576 cm⁻¹ to 1,582 cm⁻¹, with a second component becoming more pronounced at 1,618 cm⁻¹. This corresponds to an increase in the oxidation level of the CNTs. There is also a noticeable decrease in intensity of the 2D band at 2,698 cm⁻¹ and a small increase in the D+G band at 2,941 cm⁻¹ with USO treatment time, indicating an increased level of oxidation in the CNTs²⁰.

The intensity ratio of the D to G bands, I_D/I_G , was measured at 1.18 with no USO, increased to

1.37 after 155 min of USO treatment, and continued to increase at a much slower rate with greater USO treatment times. This may indicate that oxidation is occurring primarily at existing defects, and once the existing defect sites are saturated, the oxidizing effect of further USO treatment time occurs at a slower rate. This suggests that there is minimal damage to the as-received CNT structure from USO.

The SEM images in Figure 6 show USO-treated CNTs compared with untreated CNTs. The untreated CNTs are highly clumped, while the USO-treated CNTs are much more evenly dispersed across the surface, showing that the USO has helped to reduce agglomerates present in the as-received material.

The USO-treated CNTs were then electrophoretically deposited using a variety of conditions. Thicker films were produced as the electrophoresis time increased; thicker films were also produced by using higher CNT concentrations, as shown in Figure 7. Similar results are achieved when GNPs are used. Figure 8 shows the rate of electrophoretic deposition at various CNT concentrations, using deposition rates calculated from curve-fitting the data in Figure 7. It appears that the rate of EPD of USO-treated CNTs can be predicted reasonably well if the initial CNT concentration is known. The linear relationship in deposition rate and dispersion concentration is predicted from the Hamaker Law²¹, but the logarithmic rate in deposition shows that changes in the deposition parameters are occurring with time. Our previous work, in which CNTs were deposited onto carbon¹² and glass¹³ fabrics, also clearly showed that the deposition rate with time is non-linear. Under constant voltage deposition, the resistance of the CNT film increases, leading to a reduction in the deposition rate.

FIGURE LEGENDS:

Figure 1: Ultrasonic Ozonolysis System. This schematic diagram illustrates how to connect the various elements of the ultrasonic ozonolysis system.

Figure 2: Electrophoretic Deposition Cell. This schematic diagram illustrates the configuration of the electrophoretic deposition cell.

Figure 3: Chemical Characterization – XPS. CNTs that had been subjected to USO for varying times were analyzed using XPS. This image shows the wide scan changes in oxygen (532 eV) and carbon (284.6 eV) intensity, with the oxygen intensity increasing at higher USO times.

Figure 4: Atomic Ratio – XPS. This graph shows the change in the O:C atomic ratio of CNTs with increasing USO times, calculated from the scans shown in Figure 3. The ozonolysis mechanism is very complex, and the polynomial fit is provided, as a guide to the reader, to show the trend in oxidation rate, but not to explain anything about the ozonolysis mechanism.

Figure 5: Structural Characterization – Raman. All spectra showed strong D bands ($\sim 1,346\text{ cm}^{-1}$), G bands ($\sim 1,576\text{ cm}^{-1}$), 2D bands ($\sim 2,698\text{ cm}^{-1}$), and D+G bands ($\sim 2,941\text{ cm}^{-1}$). The D-band increase indicates some increase in the graphitic defects.

Figure 6: Film Morphology – SEM. These images show CNTs that were (a) not chemically treated and (b) USO-treated for 16 h.

Figure 7: Electrophoresis Measurements. This graph shows the density of the EPD films versus the deposition time using three different concentrations of CNTs dispersed in water. Blue diamonds = 2 g/L, red squares = 1.5 g/L, and green triangles = 1 g/L. A 14 V/cm DC electric field was used for these samples.

Figure 8: Electrophoretic Deposition Rate. Plot of the EPD rate versus the CNT concentrations, using deposition rates calculated from the curve fit of the Figure 7 plot of film density versus deposition time.

Table 1: Chemical Characterization – XPS. This table shows the peak fitting parameters used to deconvolute the C 1s spectrum for the GNPs processed using the USO method for 0-h and 12-h treatment times. It also shows the resulting atomic concentrations of each species¹⁴. Gr 1, Gr 2, etc. represent the various graphite peaks used in the fit.

DISCUSSION:

When working with nanoparticles of high hardness, such as CNTs, the potential erosion effect on containers and tubing should not be overlooked. Step 1.14 in the protocol was inserted after the tubing became worn at a bend due to CNTs impinging on the tube side wall, causing a system leak.

Also, note that the CNTs are in suspension, not solution, and that they must be stirred before each use if a homogeneous suspension is desired. For example, this would be necessary to maintain the desired concentrations if smaller volumes are decanted from a large batch.

It is very difficult to obtain a homogeneous dispersion of CNTs without the chemical modification of their surfaces. SEM imaging shows how untreated CNTs in a dispersion deposited on a surface remain highly agglomerated, while USO-treated CNTs are well dispersed across the surface. The EPD of untreated CNTs was also problematic, with untreated CNTs unable to be deposited by electrophoresis due to the absence of surface charge, which led to the zero electrophoretic mobility of the CNTs.

XPS showed that CNTs and GNPs treated with USO had high levels of surface oxygen, compared to insignificant oxygen levels in untreated CNTs or GNPs. The presence of the oxygen functional groups clearly provided the surface charge on the carbon nanomaterials, which facilitated stable dispersion and electrophoretic mobility under the applied field used during EPD. Likewise, Raman spectroscopy demonstrated that oxidation of the CNTs increased with USO treatment time. Raman spectroscopy also suggested that USO caused minimal damage to the graphitic structure of the CNTs.

The USO method was found to be highly effective at improving the compatibility of CNT and GNP

in polymeric composite systems. Processes where reinforcing fibers were coated with USO-treated CNTs prior to resin infusion were shown to improve the in-plane shear strength of composite laminates^{12,13} and the adhesion strength of glass-resin bonds²². CNTs and GNPs that were treated by USO produce stable dispersions that enable EPD on both conducting and non-conducting substrates at a controlled rate.

Compared to other techniques for the chemical modification of carbon nanomaterials, ozonolysis has relatively low workplace health and safety risks. No harsh chemicals are used, and the water in which the carbon nanomaterials are processed can be easily removed, providing a functionalized material that can be readily used in a range of practical applications.

While the USO method has several advantages, such as allowing for the production of functionalized carbon nanomaterials without the use of harsh chemicals, the process does take up to 16 h to produce a stable dispersion suitable for EPD. Clearly, future work will need to examine how the process can be accelerated to enable greater throughput.

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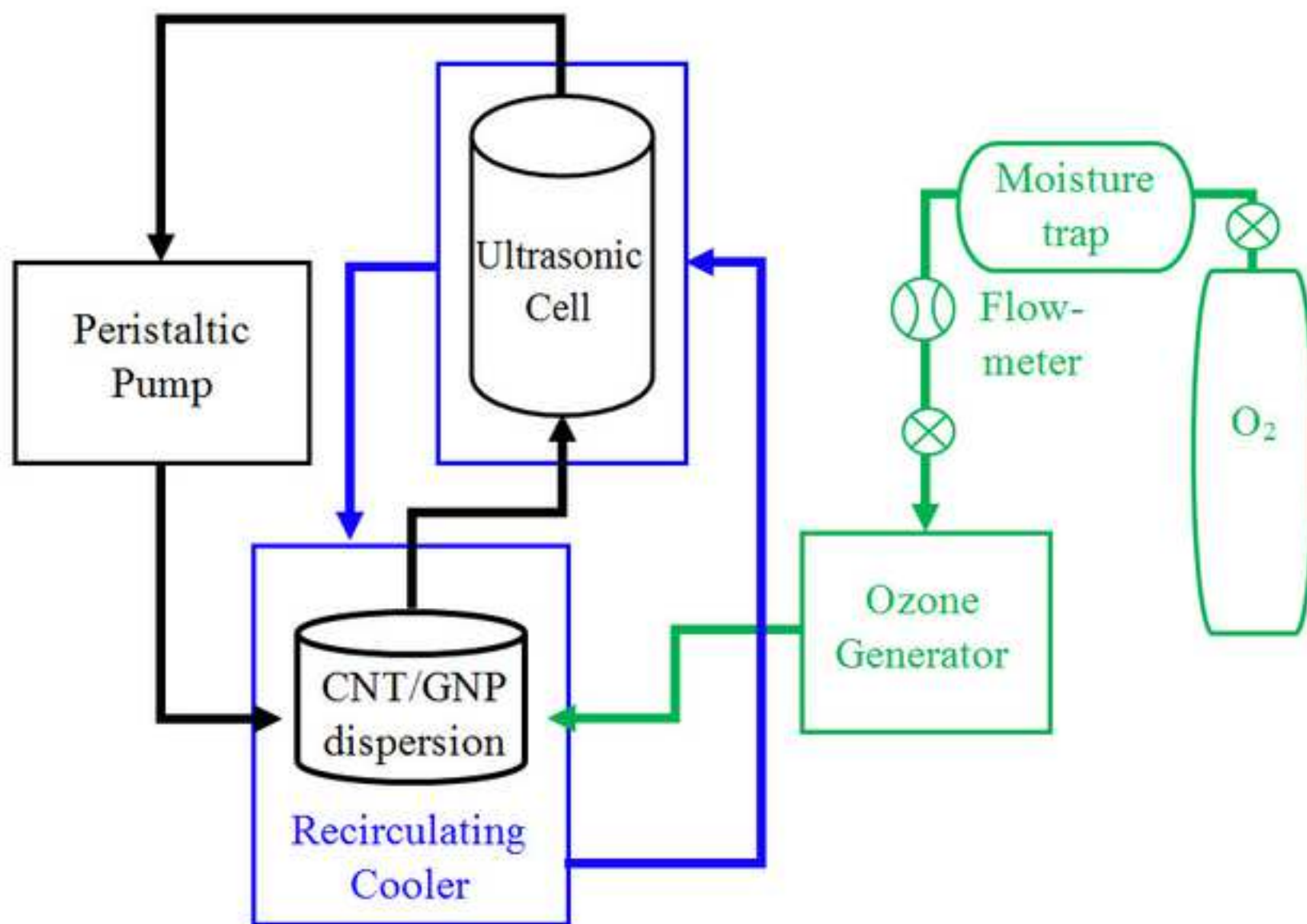
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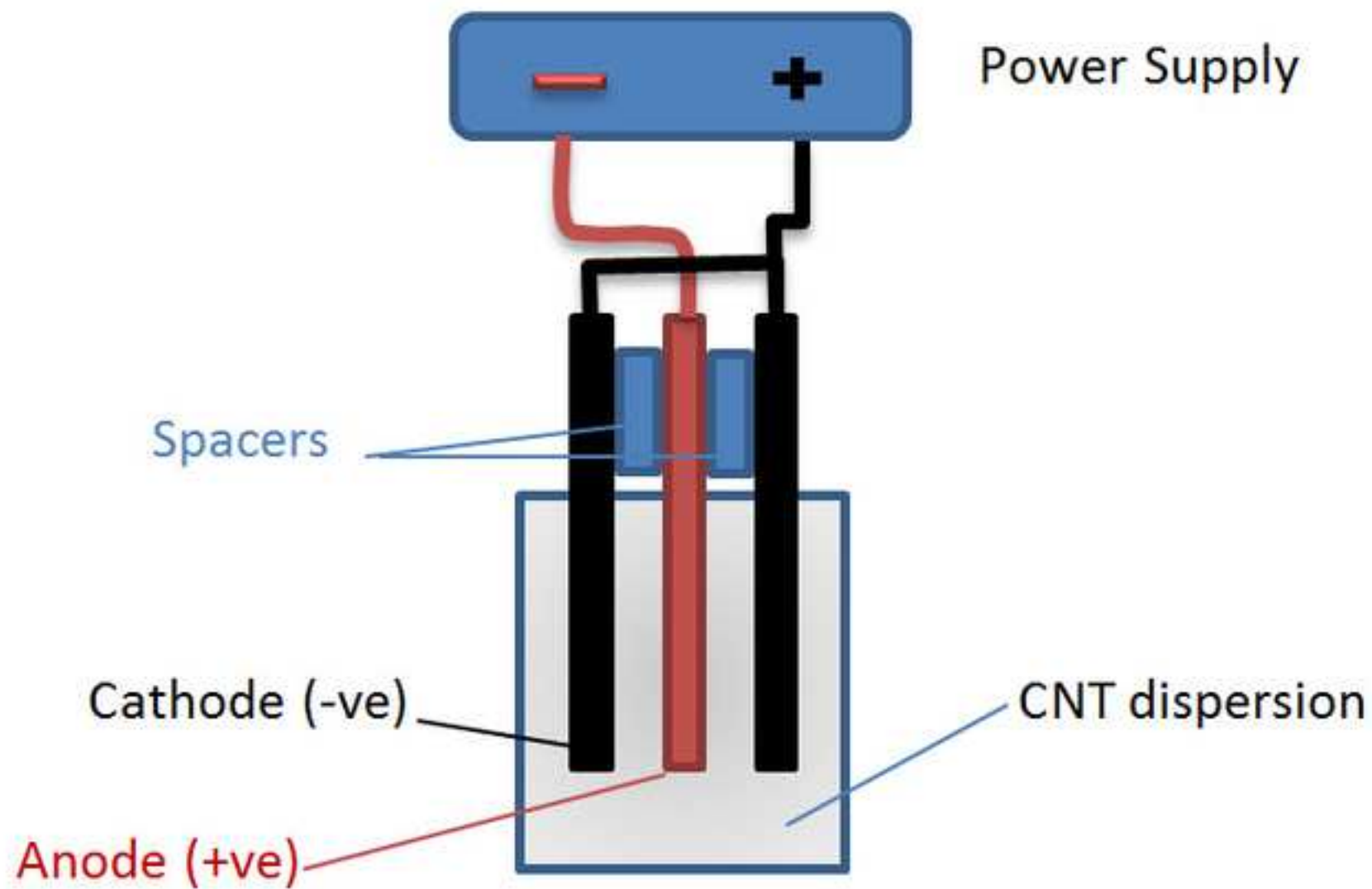
The authors have nothing to disclose.

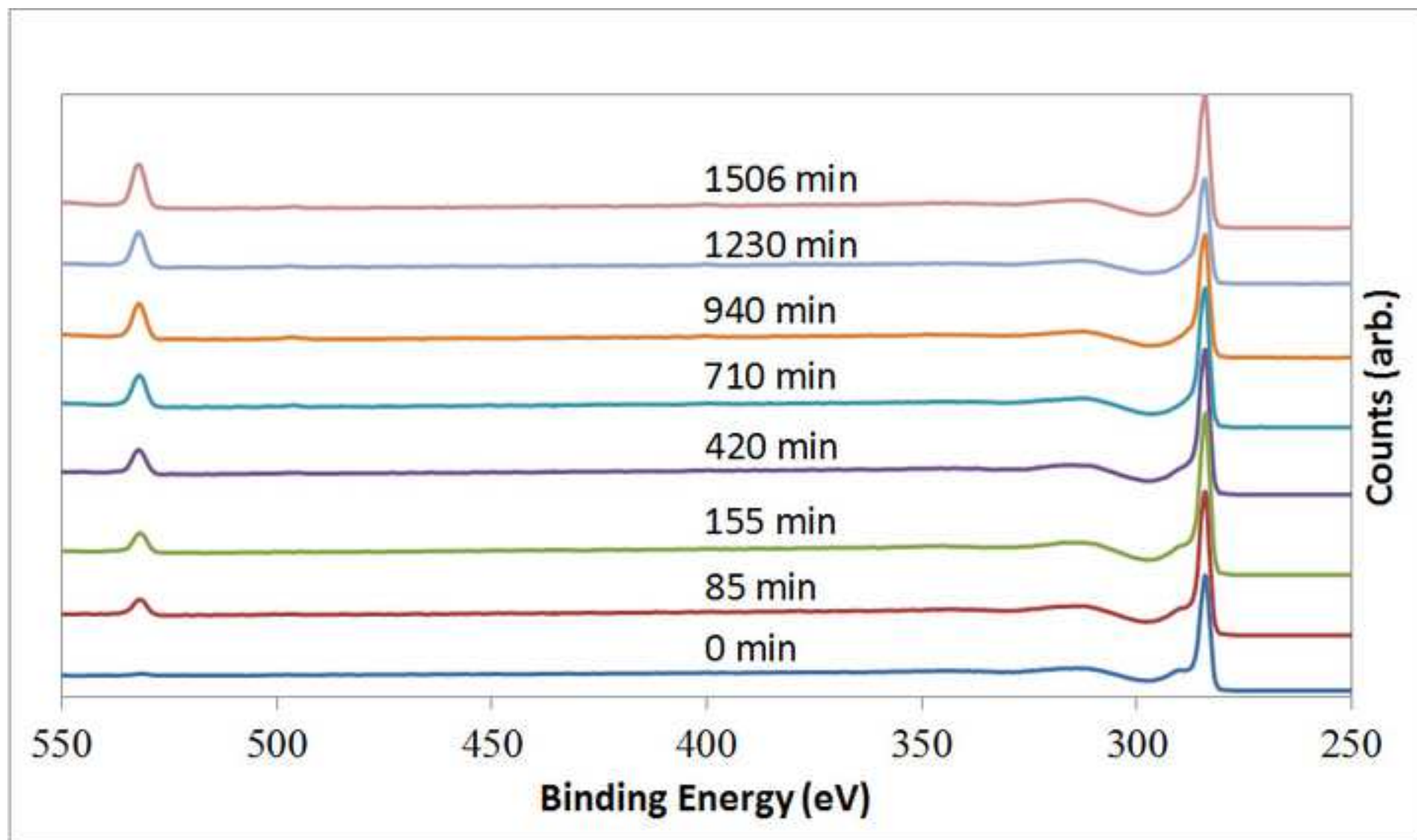
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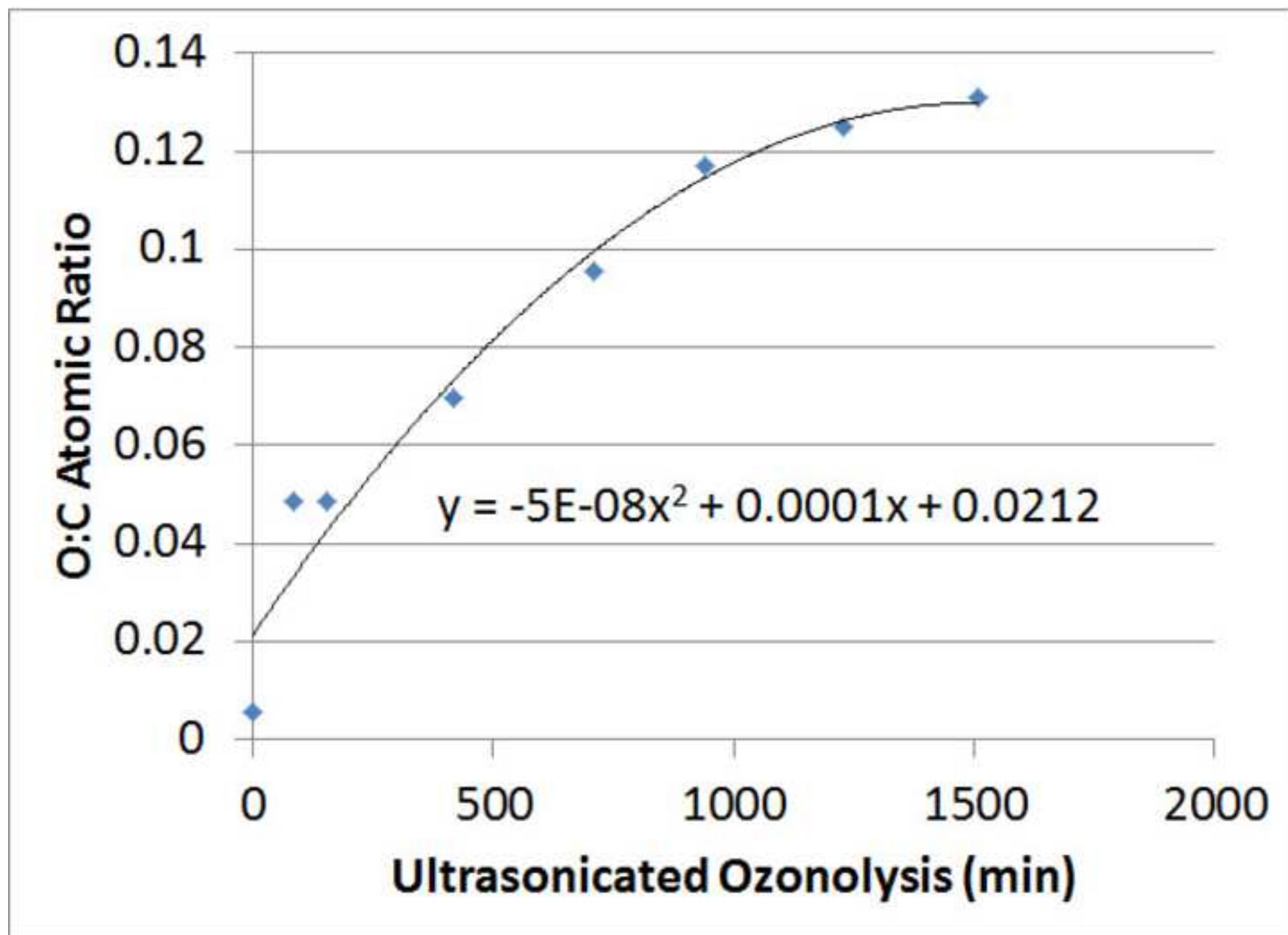
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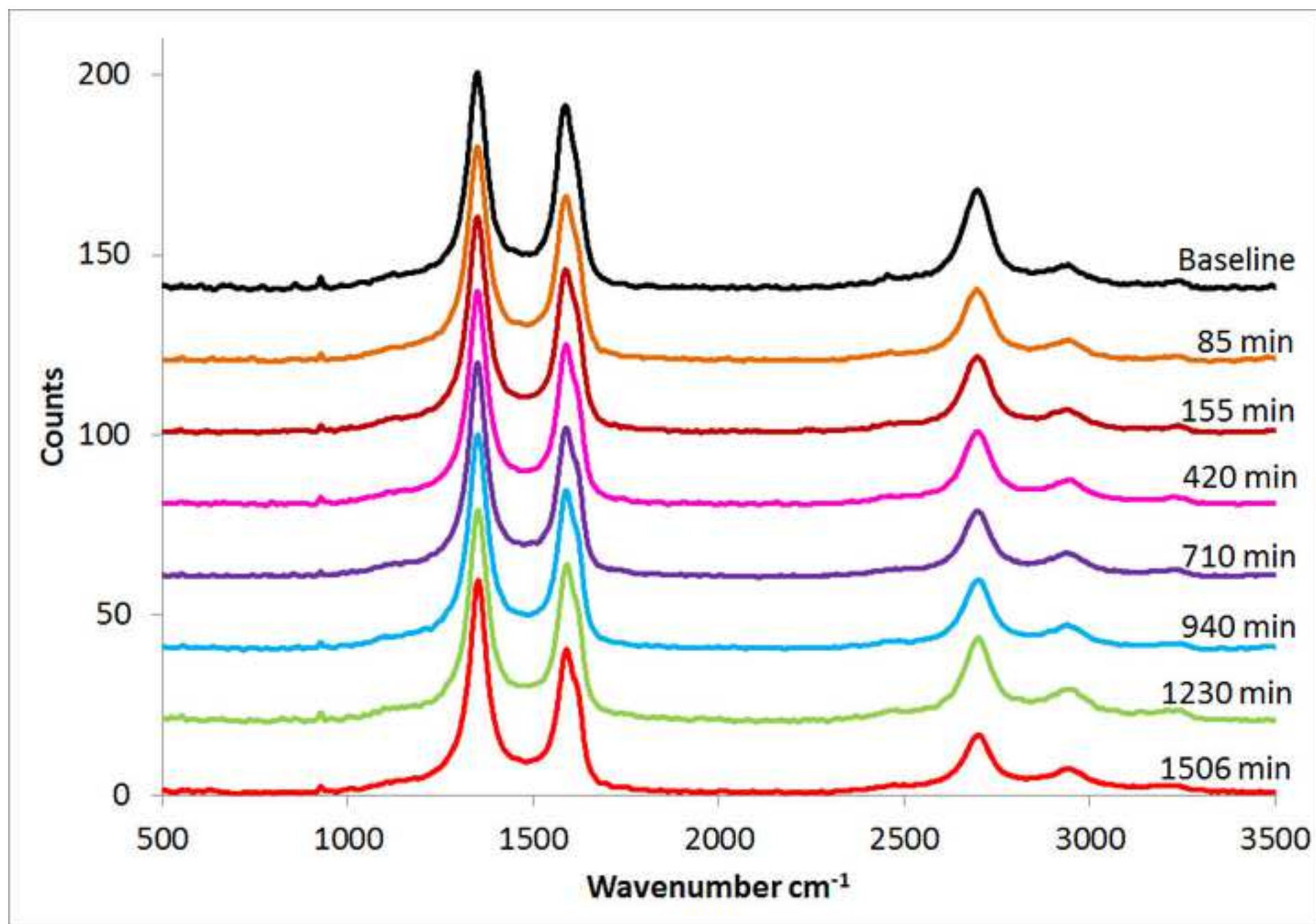
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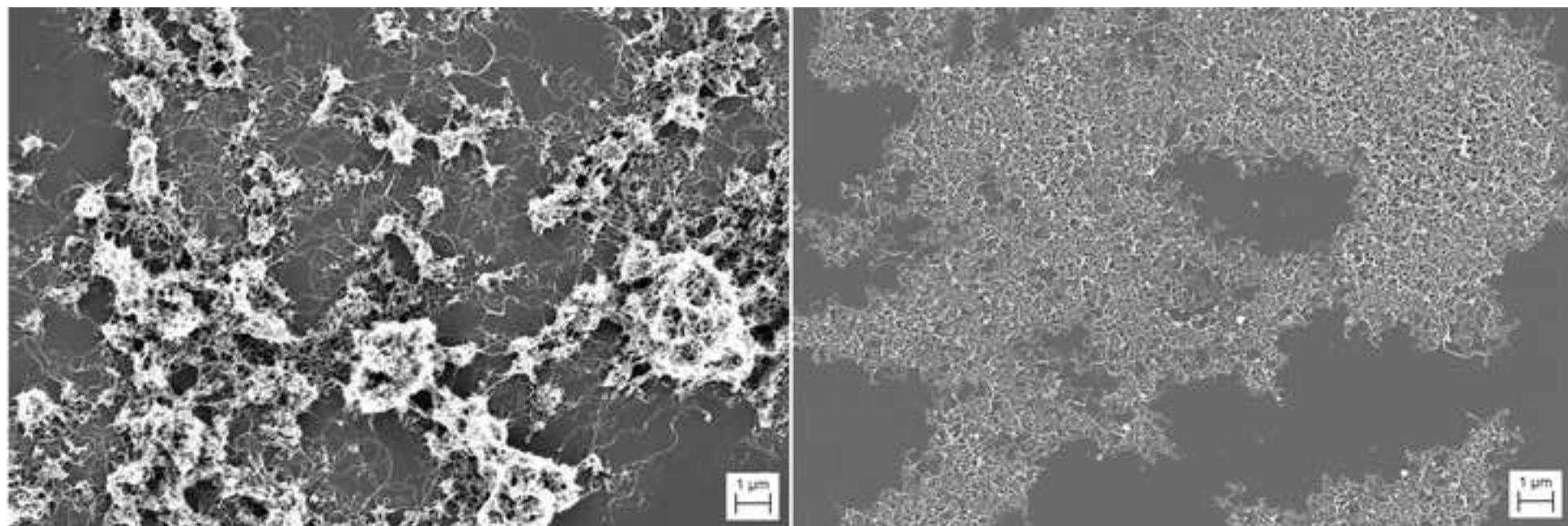


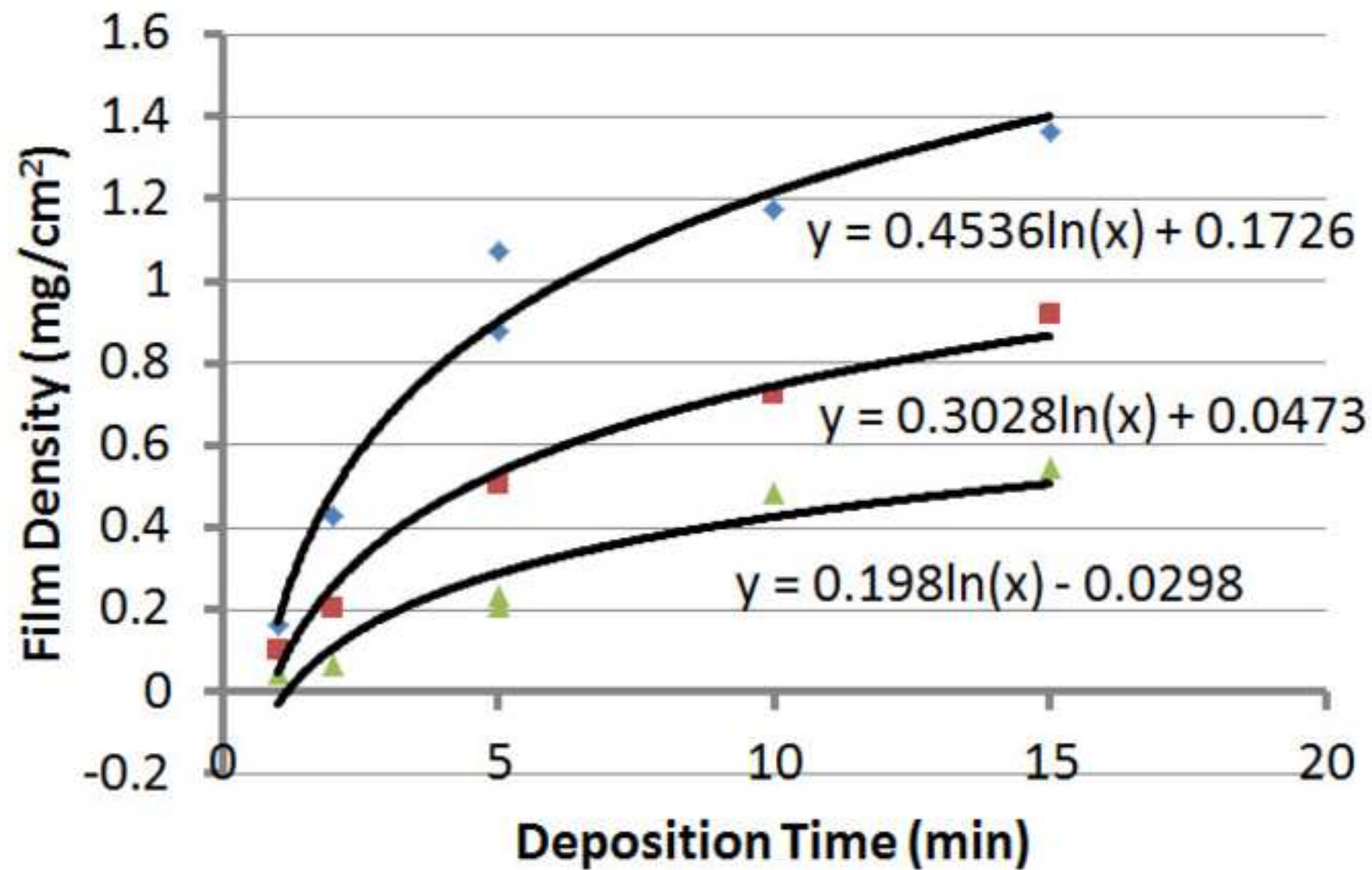


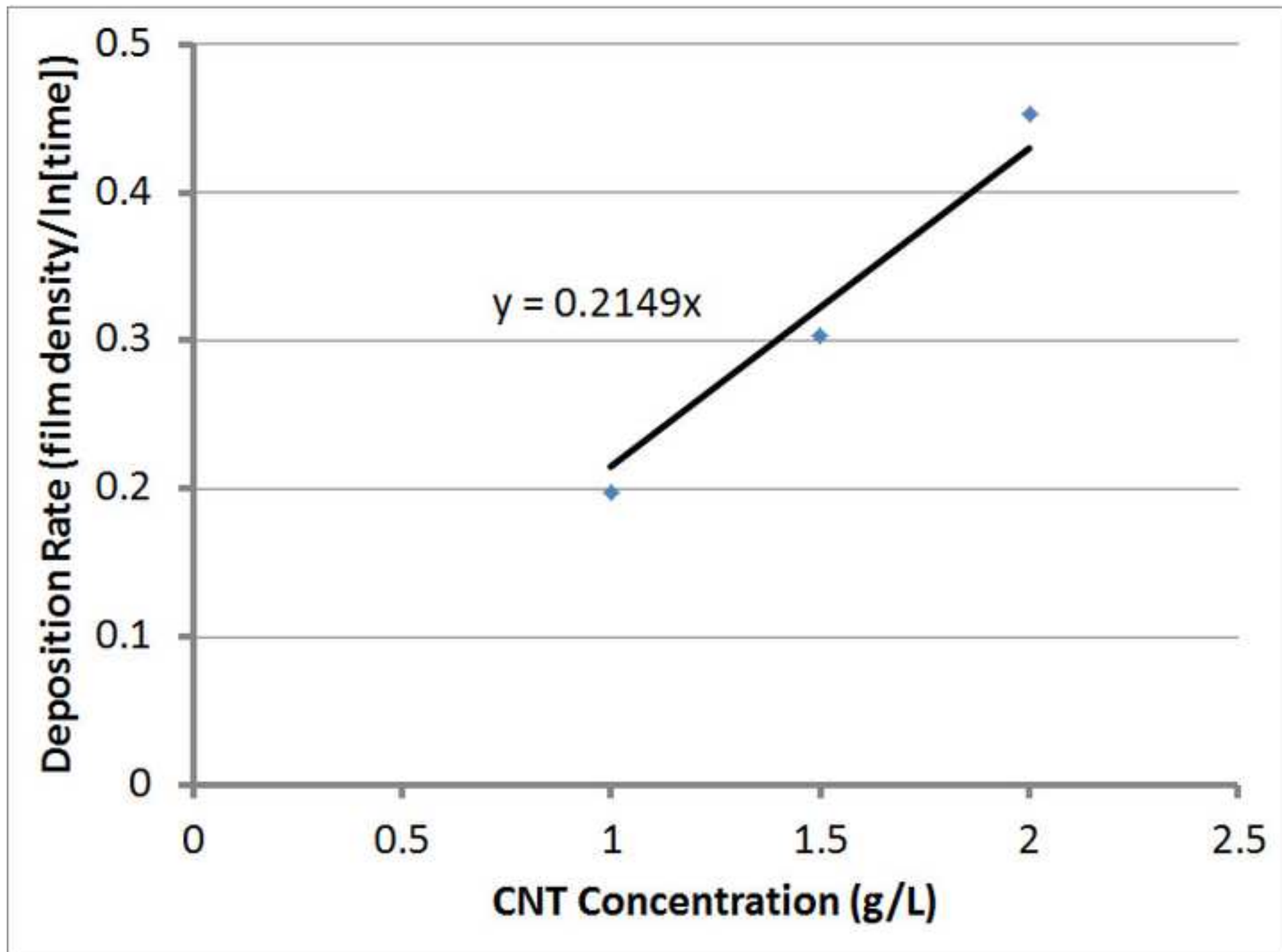












Name	Position	Constraint	FWHM	Constraint	Area Constraint	% Atomic Concentration	
						0 h	12 h
Gr 1	284.6	±0.1	0.7	±0.2	Best fit	29.4	17.6
Gr 2	284.8		1.2		Gr1*1.3	36.9	22.2
Gr 3	286.1		1.3		Gr1*0.4	12.5	7.5
Gr 4	287.9		1.3		Gr1*0.2	5.9	3.5
Gr 5	289.5		1.6		Gr1*0.1	3.3	1.8
Gr 6	291.3		2.4		Gr1*0.2	6.6	1.7
C-C	285		1.2		Best fit	2.1	13.7
C-O	287		1.2		Best fit	1.5	14.3
C=O	288.6		1.2		Best fit	0.9	12.4
COO	289.4		1.2		Best fit	0.9	5.3

Name of Material/ Equipment	Company	Catalog Number	Comments/Description
Ultrasonic bath	Soniclean	80TD	Daintree Scientific
Ultrasonic horn	Misonix	S-4000-010 with	
Ozone generator	Ozone	CL5 converter	
Recirculating liquid cooler	Solutions	TG-20	
Multi-mode power supply unit	Thermoline	TRC2-571-T	
High resolution computing multimeter	TTi	EX752M	
X-ray photoelectron spectroscopy	TTi	1906	
	Kratos		
	Analytical	Axis Nova	
XPS analysis software	Casa		
Kratos elemental library for use with Casa XPS	Software	Casa XPS	www.casaxps.com
Raman dispersive confocal microscope	Casa	Download Kratos	
Field emission scanning electron microscope	Software	Related Files	http://www.casaxps.com/kratos/
	Thermo	DXR	
	Leo	1530 VP	
Sputter coater with iridium target	Cressington	208 HR	
Thickness measurement unit	Cressington	mtm 20	
Magnetic stirrer	Stuart	CD162	
Analytical balance	Kern	ALS 220-4N	
	Mettler	NewClassic MF MS	
Analytical balance	Toledo	2045	
Laboratory balance	Shimadzu	ELB 3000	
Electrodes from 316 stainless steel sheet	RS Components	559-199	
Sanding sheets, P1000 grade	Norton	No-Fil A275	

Multi-walled carbon nanotubes	Hanwha	CM-95	http://hcc.hanwha.co.kr/eng/business/bus_table/nano_02.jsp
Graphene nanoplatelets	XG Sciences	XGNP Grade C	http://xgsciences.com/products/graphene-nanoplatelets/grade-c/

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Aerospace Division
506 Lorimer Street
Fishermans Bend, VIC 3207
AUSTRALIA

Tel: +61 3 9626 8703
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A handwritten signature in dark ink, appearing to read 'Brian Hanlon'.

Dr Brian Hanlon
A/Chief Aerospace Division
For and on behalf of the Commonwealth of Australia

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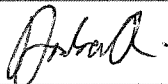
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Replies to Editorial Comments

Please note: There seems to be a problem with Track Changes and Comments. Some of the changes do not appear to be highlighted, and my comments look like they were written by you.

- Protocol Detail: Please see the in-text comments made in Word. These need to be addressed.

- Thank you for your guidance, checked changes and addressed the comments as appropriate.

- Results:

1) Apologies, I have checked the figure call outs and hopefully they are correct now.

2) & 3) The direct correlation between the concentration and rate is well known and expected from the Hamaker law. The plot is provided to show that our results are in agreement with theory, rather than to substantiate a new theory. The rate reflects the reduction in deposition rate with time that results from the increase in resistivity that occurs as the film builds up due to the low conductivity of the CNTs compared to the electrodes. This comment has now been added from line 407 to explain the logarithmic relationship. The non-linear trend is shown in our previous two EPD papers and is now referenced.

4) Inserted commentary at lines 375-380.

- Discussion: Inserted additional discussion at lines 490-193.

- Figures:

1) Am I allowed to submit differently formatted figures for the print version of the article and the video version? If so, I would like to submit additional figures. However due to other work commitments, this might take a few weeks before I have time to reformat them. Also, Benjamin Werth said I would be asked to check the quality of the video before it is made available online. He suggested that I could revise the figures if we decided they were not optimal for video format.

2) Figure 3: Scientific protocol dictates that an axis be provided to indicate it represents the number of counts measured during spectrum recording. The actual numbers are not important as this plot shows qualitative changes. This is a standard method to present XPS data. If you wish, I can move the x-axis to the left side instead of the right.

3) Figure 4: The ozonolysis mechanism is very complex and the line fit is provided to guide the viewer to show the trend in oxidation rate but not to explain anything about the ozonolysis mechanism. (This comment could be added to the caption if required.)

4) Table 1: Inserted missing label.

- Figure/Table Legends:

1) Fig 1: Edited figure and removed reference to MWCNT.