**TITLE:**

Evaluation of drug sorption to PVC- and non-PVC-based tubes in administration sets using a pump

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Drug, Sorption, Tubes, PVC, Non-PVC, Administration set

**SHORT ABSTRACT:**

We report a method for drug sorption evaluation using a pump and model drugs (diazepam and tacrolimus). After drug analysis using high-performance liquid chromatography, drug concentrations and sorption levels in the tubes of administration sets are calculated.

**LONG ABSTRACT:**

Administration sets are delivery tools for the direct application of drugs into the body and are composed of a spike, a drip chamber, tubes, Luer adapters (connectors), a needle cover for protection, and other accessories. Drug sorption to tubes of administration sets is a critical issue in terms of safety and efficacy. Although drug sorption is an important factor in the quality of an administration set, there are no standard evaluation methods for the regulation of drug sorption to the tubes. Here, we describe an evaluation protocol for drug sorption to tubes of administration sets. Tubes made of polyvinyl chloride (PVC)- and non-PVC-based polymeric materials were cut to 1 m in length. Diazepam and tacrolimus were used as model drugs. In the kinetic sorption study, we selected the drug concentration and flow rate based on the clinical usage of these drugs. After the dilution of each drug in a glass bottle, the diluted drug solution was delivered through tubes of administration sets using a pump. Samples were collected in amber vials at appropriate time points and the drugs were analyzed using high-performance liquid chromatography. Drug concentrations and sorption levels to tubes of the administration sets were calculated. Acceptable criteria to ensure the quality of administration sets are recommended.

**INTRODUCTION:**

Administration sets are composed of a spike, a drip chamber, tubes, Luer adapters (connectors), and a needle cover for protection. Other accessories, such as an airway check valve, a regulating clamp, an in-line filter, a Y-tube with cap (an injection port), and a needle, can also be attached to administration sets. Drug sorption to tubes is a critical issue in the delivery of injectable drugs1. Sorption describes the adsorption of a drug to the polymer surface and the absorption of a drug into the polymeric matrix2. Drug sorption to tubes in administration sets causes unpredictable drug loss and makes it difficult to control the delivered drug concentration. Drug sorption to polymeric tubes is therefore a major impediment to the precise transfer of injectable drugs into the body. However, there are no standard methods or regulatory guidelines for the evaluation of drug sorption to tubes in administration sets.

The levels of drug sorption to tubes in administration sets have been reported using various evaluation methods1-5. Test methods and model drugs are key factors in sorption evaluation. The pump method1, 3 and drip method4, 5 have been widely used for sorption tests. In general, the pump method should be used in the case of drugs with low concentrations and low flow rates as the infusion conditions. Using various evaluation methods, many studies of drug sorption have been reported for polyvinyl chloride (PVC)- and non-PVC-based tubes in administration sets1-3, 5. Many sorptive drugs can be selected to investigate whether the tubes of the administration sets have drug sorption potential or not. Diazepam (Figure 1a)1, 2, tacrolimus (Figure 1b)5, nitroglycerin2, and cyclosporin A3 are representative drugs with high sorption in PVC- and non-PVC-based tubes.

For the evaluation of drug sorption to the tubes, test conditions such as flow rate and drug concentration are based on the clinical use of the selected drugs1, 6, 7. In the case of diazepam, a high concentration of 100 μg/mL was used at a flow rate of 1 mL/min to mimic the initial dose for the treatment of status epilepticus1. For tacrolimus, a concentration of 10 μg/mL was delivered at a flow rate of 10 mL/h. Dextrose solution (5%) was used for the dilution of drug injections, and tube length was fixed at 1 m. Glass bottles and vials should be used to prevent additional sorption during the experiment and storage.

In this study, we conducted a kinetic sorption study with the model drugs, diazepam and tacrolimus, using a pump method. Specific details of this protocol, from tube preparation to sorption evaluation, were described previously. Methods for the evaluation of drug sorption have already been used to confirm drug properties for injections and to recommend the clinical use of injections with administration sets on a case-by-case basis1-7. This protocol may be used as a standard technique for the sorption evaluation of administration sets. International standards for the evaluation of drug sorption to tubes may be necessary to ensure the safety and efficacy of drug delivery.

**PROTOCOL:**

**1. Preparation of tubes in administration sets**

Note: Precisely perform the cutting step to eliminate the effect of differences in tube length on drug sorption.

1.1) Label the end of the tubes with the tube type (*e.g.,* PVC, polyurethane (PU), and polyolefin (PO)) using a marker.

1.2) Remove all detachable accessories, such as connectors and needle covers.

1.3) Using a sharp razor to ensure clean edges, cut the tubes to 1 m in length from the connection of the drip chamber.

**2. Dilution of drug injections**

Note: Use a glass bottle (1 L) as the container for the injected drug solution. Perform the dilution step precisely. Verify the composition of the marketed drug product and use the same batch number for a whole experimental set.

2.1) Label bottles with the drug names (*e.g.,* diazepam or tacrolimus).

2.2) Dilute the drug injections with 5% dextrose solution, from 1 mg/mL to 100 μg/mL for diazepam injections (10 mL of diazepam injection in 100 mL of 5% dextrose solution) and from 5 mg/mL to 10 μg/mL for tacrolimus injections (200 µL of tacrolimus injection in 100 mL of 5% dextrose solution).

Note: Set up the tested concentrations of drugs and solvents based on the clinical usage.

2.3) Gently mix the diluted solutions in the bottle by swirling to obtain homogenous drug solutions.

2.4) Collect diluted solutions (1-10 mL) in amber vials using a glass graduated cylinder.

2.5) Verify the concentration, as described in step 4.

Note: Use these concentrations of drugs as concentrations at the starting points.

**3. Kinetic sorption study using an infusion pump**

Note: Confirm the tube-dependent flow rate using a pump prior to the sorption test due to the hardness of tubes. Collect samples at precise time points and use glass bottles and vials to prevent additional drug sorption during storage. Perform the test as shown in Figure 2. Protect the drug solution against light if the drug has photosensitivity. Perform the experiments in triplicate.

3.1) Without creating air bubbles, preload a diluted solution of the drug into the tube using a syringe.

3.1.1) Connect one end of the tube to a syringe.

3.1.2) Put the other end of the tube into the bottled drug solution.

3.1.3) Pull back the syringe plunger until the tube is completely filled with the drug solution.

3.2) Install the tube into an infusion pump.

3.2.1) Open the door of the infusion pump and push the release lever.

3.2.2) Insert the preloaded tube into the infusion pump and keep it straight.

3.2.3) Remove the syringe at the end of the tube after installation.

3.2.4) Put the end of tube into a chemically resistant borosilicate glass graduated cylinder to collect the drug solution after it passes through the tube.

3.3) Set the flow rate based on the type of tube in the administration set (PVC, PU, or PO) and the drug (*e.g.,* 1 mL/min for diazepam and 10 mL/h for tacrolimus).

3.4) Collect samples into amber vials at various time points, at room temperature.

3.4.1) Collect 1-mL diazepam samples at 0.05, 0.30, 0.55, and 1.05 h.

3.4.2) Collect 10-mL tacrolimus samples at 1.05, 2.05, 3.05, and 4.05 h.

**4. Analysis of drug using high-performance liquid chromatography (HPLC)**

Note: Recommended HPLC methods for drug analysis are described in references 1, 8, 9. Use tandem mass spectrometry (MS/MS) and immunoassay after sample preparation as alternative methods10, 11. Perform the experiments in triplicate.

4.1) Weigh the drugs and dissolve them in organic solvents at a concentration of 1 mg/mL as stock solutions.

4.1.1) Use methanol as a solvent for diazepam stock solution due to the low solubility of diazepam in 5% dextrose.

4.1.2) Use acetonitrile as a solvent for tacrolimus stock solution due to the low solubility of tacrolimus in 5% dextrose.

4.2) Prepare standard solutions by diluting the stock solutions.

4.2.1) Dilute diazepam stock solutions with methanol to 0.3125, 0.625, 1.25, 2.5, 5.0, 10.0, and 20.0 µg/mL.

4.2.2) Dilute tacrolimus stock solutions with 5% dextrose to 2.5, 5.0, 10.0, 15.0, and 20.0 μg/mL.

4.3) Analyze standards using the HPLC method with UV detection1, 8, 9.

Note: Use an appropriate detection method (UV, fluorescence, etc.) for drug analysis.

4.3.1) Inject 10 µL of standards into an HPLC system with UV detection equipped with a C18 column (1.5 mm × 250 mm, 5 µm). See Table 1 for analysis conditions.

4.3.2) Confirm the specificity and linearity1, 8, 9.

4.3.2.1) For specificity, monitor the drug peak (*i.e.,* whether it was separated from other peaks in the chromatogram) to identify the drug.

4.3.2.2) Confirm linearity at the calibration range (*i.e.,* whether the peak area results are directly proportional to the concentrations).

4.3.3) Obtain calibration curves.

4.3.3.1) Create graphs based on concentration versus peak area value from chromatograms of the standards1, 8, 9.

4.3.3.2) Obtain linear regression equations with R2 for the calibration curves (*e.g.,* y = ax + b, x: concentration of drug, y: peak area)1, 8, 9.

4.4) Dilute the samples from the sorption study with methanol for diazepam or 5% dextrose for tacrolimus as appropriate so that they fall within the calibration range and directly inject 10 µL of diluted sample into the HPLC system.

**5. Calculation of drug concentration and sorption level**

5.1) Calculate the drug concentrations of the samples using calibration curves (unknown x and known y).

5.2) Calculate the sorption levels of the drugs using the following equation:

where S: Sorption level (%)

Co: Drug concentration after the dilution of the injections (μg/mL)

Cp: Drug concentration passed through the tubes (μg/mL)

**REPRESENTATIVE RESULTS:**

The sorption to tubes in the administration set was kinetically monitored using the model drugs, diazepam (Figure 1a) and tacrolimus (Figure 1b), and the pump method (Figure 2). Diluted drug (Figure 2a) was passed through PVC- and non-PVC-based tubes (Figure 2b) at a fixed flow rate using an infusion pump (Figure 2c). The glass bottle was opened slightly to allow the insertion of the administration set tube. After the drug was delivered through the tubes (Figure 2d), samples were collected into amber vials (Figure 2e). All samples, including samples at the starting points, were analyzed using an HPLC method with UV detection (Figure 3a). Analysis conditions are listed in Table 1. For the preparation of drug standards, diazepam and tacrolimus were dissolved in methanol and acetonitrile because of their insolubility in 5% dextrose. The drug concentrations at the starting points of the sorption study were calculated from the analysis of the samples after the drug dilution. Sorption levels in PVC- and non-PVC-based tubes of administration sets were determined by calculating the percentage of remaining drug content after passage through the tubes from the calibration curves (Figure 3b) and subtracting these values from 100% (Figure 3c). The recommended acceptable range of drug sorption percentages was less than 10%, based on the content of injections from pharmacopoeias12. In addition, the specific drugs (*e.g.,* anticancer drugs) should be confirmed with clinical guidelines. We determined whether drug sorption levels of the samples were appropriate (Figure 3d). Figure 4 and Figure 5 show representative chromatograms at low and high concentrations of drugs for standards and sample solutions, respectively. Retention time of each drug was 8.2 min for diazepam and 6.8 min for tacrolimus. There were no interfering peaks from the matrix. Specifically, in samples of the diazepam sorption study, the interfering peaks did not overlap with drug peak in the chromatogram, although a different solvent (5% dextrose) was used than in the standards. Table 2 shows calculations of representative sorption levels of diazepam and tacrolimus. The sorption level of each drug was the highest in PVC-based tubes, lower in PU-based tubes, and lowest in PO-based tubes at the initial phase of the kinetic sorption test.

**FIGURE LEGENDS:**

**Figure 1. Chemical structures of model drugs: (a) diazepam and (b) tacrolimus.** Diazepam is a benzodiazepine derivative and tacrolimus is a 23-membered macrolide lactone. This figure has been modified1.

**Figure 2. Test set of a kinetic sorption study using a pump.** (a) Drug diluted with 5% dextrose in a bottle, (b) tube of administration set (1 m in length), (c) infusion pump, (d) drug passed through the tube, and (e) amber vials for storage. To minimize additional drug sorption, drug solutions and samples were prepared and stored in a glass bottle and amber vials for injections, respectively. This figure has been modified1.

**Figure 3. Key steps for evaluating drug sorption level to tubes of administration sets.** (a) Analysis of drug using an HPLC method with UV detection, calculation of (b) drug concentration from the calibration curve and (c) sorption level (%), and (d) recommendation of acceptable criteria for drug sorption. Several steps in the sorption evaluation, from drug selection to the sorption test, such as the analysis of drug concentration and the calculation of sorption levels, are described.

**Figure 4. Representative chromatograms of diazepam.** (a) Blank (methanol), standards at (b) 0.3125 µg/mL and (c) 20 µg/mL, and (d) sample (5% dextrose). The peak of diazepam was detected at 8.2 min, and fluctuations in retention time occurred within 1 min in the chromatograms. The peaks from the solvent were presented before the diazepam peak in the blank and standard chromatograms.

**Figure 5. Representative chromatograms of tacrolimus.** (a) Blank (acetonitrile with 5% dextrose), standards at (b) 2.5 µg/mL and (c) 20 µg/mL, and (d) sample. The tacrolimus peak at 6.8 min was successfully separated in a chromatogram. There were no peaks that overlapped with the tacrolimus peak. The peak was detected at 2.5 µg/mL, with a signal-to-noise ratio greater than 10.

**Table 1. HPLC conditions.**

**Table 2. Representative sorption results for diazepam and tacrolimus in PVC- and non-PVC-based tubes (n = 3)**1**.**

**DISCUSSION:**

Drug sorption to administration sets is a cause of unexpected drug loss in intravenous drug delivery. During sorption, drugs are generally partitioned to polymeric materials of tubes at the early phase of infusion; after sorption equilibrium is reached, the delivered amount of drug is stabilized1. The sorption levels of drugs should be evaluated and minimized. Several evaluation methods for drug sorption have been studied, such as a pump method and a drip method. Compared to the drip method, the pump method can be easily manipulated without bias. Although administration sets with flow regulators (conventional form) are used in the drip method, desired flow rates less than 5 mL/h are hard to achieve. Therefore, we recommend the pump method for the sorption evaluation of tubes in administration sets.

When using the pump method, major factors that affect drug sorption to tubes of administration sets are drug properties (*e.g.,* hydrophobicity and charge), conditions of the sorption test (*e.g.,* drug concentration, flow rate, solvent compatibility, tube length, and temperature), analytical methods for drugs (*e.g.,* HPLC and MS), and the tube polymers in the administration sets (*e.g.,* PVC, PU, and PO)2-12. First, the selection of the model drugs is critical for obtaining precise and accurate experimental results. Even though diazepam management is tracked by the Psychotropic Drugs Control Act, we selected diazepam (Figure 1a) and tacrolimus (Figure 1b) as model drugs because of their high sorption levels to polymeric tubes of administration sets1 or containers13, 14. In this case, drugs at high concentrations showed less sorption than those at low concentrations in the early phase of infusion1, 2. These drugs have high log P values (diazepam: 2.8215, tacrolimus: 3.9616) and low solubility, as categorized by the Biopharmaceutical Classification System (BCS class 2). Because of their hydrophobicity, these drugs can interact with tubes of administration sets, leading to sorption. Other drugs showing high sorption levels (*e.g.,* nitroglycerin2 and cyclosporin A3) can be used as alternative model drugs for sorption evaluation. Furthermore, macromolecular drugs, such as biologics (antibody therapeutics, insulin, etc.), can be applied for the quality evaluation of administration sets regarding drug sorption12.

We set up a simple kinetic sorption study, using a pump to easily obtain precise results and to minimize artifacts. (Figure 2). In the pump method, drug solution (Figure 2a) was passed through a tube cut from the administration set (Figure 2b) after installation into an infusion pump (Figure 2c). Except for the tubes from administration sets, all devices (bottle, graduated cylinder, and sampling vials) were composed of chemically resistant borosilicate glass to prevent additional drug sorption to polymers. In this study, tubes without other accessories at a fixed length of 1 m were used to simplify the factors of drug sorption to tubes of administration sets. If a clinical condition requires it, a multiplication factor for the length of tube can be used. In the sorption test, the diluted drug solutions were used as the starting concentrations1,7. After delivery, the drug solution (Figure 2d) was collected into vials at various time points (Figure 2e). Drug solutions for sampling were passed completely through the tube at the preselected conditions of flow rate and sampling time points. Sorption generally occurs in the early phase of infusion, and the pattern is followed by a convection-interfacial resistance-diffusion model7. Diazepam sorption results are comparable to the double-lumen extension tube model17 when the initial drug concentration is considered before delivery. Therefore, sampling time points can be modified so that sorption evaluation takes less time. All factors of the test conditions were confirmed based on the clinical usage of drugs.

In this protocol, we chose the HPLC method for drug analysis based on previous reports1, 8, 9. Simple and reproducible HPLC methods have been developed. The HPLC conditions are listed in Table 1. Various other techniques, such as MS and immunoassay, have also been developed as alternative analysis methods of drug concentrations10, 11. MS/MS and immunoassay are highly sensitive for the detection of drugs and their metabolites. Specifically, an immunoassay can easily be performed without requiring large and expensive equipment for drug analysis.

Regarding the quality evaluation of administration sets, drug sorption to PVC- and non-PVC-based materials used in the tubes of administration sets has been studied. The evaluation of sorption to tubes in administration sets started with drug selection and ended with the consideration of acceptable criteria of sorption levels, as illustrated (Figure 3). PVC-based tubes showed high sorption levels for many drugs such as diazepam, tacrolimus (Table 2), nitroglycerin2, and cyclosporin A3. Among approaches to minimize drug sorption to the tubes of administration sets to less than 10%, alternative materials or polymeric combinations have been developed, such as PO-based materials and layer-by-layer designs2, 13. The PE/PB/PP blended PO-based tube of an administration set used in this study showed low sorption levels, as a non-PVC-based tube. On the other hand, PE-based tubes are not used for administration sets, but they are commercially used in the market as a syringe extension tube due to their hardness.

This protocol can be applied to the quality control of administration sets with respect to drug sorption. More drugs classified by sorption level (highest, lower, and lowest) should be used in sorption evaluations for the quality assurance of administration sets. This protocol can also be used in scientific research for the development of new alternative polymeric materials or new designs for tubes in administration sets that do not result in drug sorption1, 13.

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**DISCLOSURES:**

The authors have nothing to disclose.

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