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Engineering the Corona Phase Molecular Recognition of Single Walled Carbon Nanotubes

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TITLE:

Engineering the Corona Phase Molecular Recognition of Single Walled Carbon Nanotubes

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molecule microscopy; neurotransmitters

SHORT ABSTRACT:

We present a protocol for engineering the corona phase of near infrared fluorescent single walled carbon nanotubes (SWNTs) using amphiphilic polymers and DNA to develop sensors for molecular targets without known recognition elements.

LONG ABSTRACT:

Semiconducting single-wall carbon nanotubes (SWNTs) are a class of optically active nanomaterial that fluoresce in the near infrared, coinciding with the optical window where biological samples are most transparent. Here, we outline techniques to adsorb amphiphilic polymers and polynucleic acids onto the surface of SWNTs to engineer their corona phases and create novel molecular sensors for small molecules and proteins. These functionalized SWNT sensors are both biocompatible and stable. Polymers are adsorbed onto the nanotube surface either by direct sonication of SWNTs and polymer or by suspending SWNTs using a surfactant followed by dialysis with polymer. The fluorescence emission, stability, and response of these sensors to target analytes are confirmed using absorbance and near-infrared fluorescence spectroscopy. Furthermore, we demonstrate surface immobilization of the sensors onto glass slides to enable single-molecule fluorescence microscopy to characterize polymer adsorption and analyte binding kinetics.

INTRODUCTION:

Single-walled carbon nanotubes (SWNTs) are atomically thin layers of carbon atoms rolled into long, thin cylinders that exhibit unique electronic and optical properties.¹ Such properties include a band-gap producing near infrared (nIR) fluorescence emission via exciton recombination that is highly sensitive to its local environment. The nIR emission of SWNTs falls within the near infrared window in which the penetration depth of light is maximal for biological tissue.^{2,3} Additionally, SWNTs exhibit several unique features atypical in contrast to organic fluorophores: SWNT exhibit a large Stokes shift, do not photobleach, and do not blink.⁴ Recently, exploiting these characteristics has led to the development of an assortment of novel molecular sensors with applications to biology.^{5,6} Unmodified, however, SWNTs are insoluble in water, and obtaining suspensions of individual SWNTs can be a challenge.^{7,8} Bundling and aggregation of SWNTs in solution can obfuscate their band-gap fluorescence,² rendering them unsuitable for sensing applications.

Dispersing individual carbon nanotubes in aqueous solution requires modifying their surface to prevent hydrophobicity-driven aggregation.⁹ While covalent modification can render SWNTs water-soluble,¹⁰ as well as impart specific binding chemistry, defect sites in the SWNT lattice reduce or abate their fluorescence emission. Instead, SWNT functionalization can be accomplished by using surfactants, lipids, polymers and DNA^{9,11–13} that adsorb to the nanotube surface through hydrophobic and pi-pi stacking interactions. The resulting chemical environment surrounding surface-functionalized SWNTs is referred to as its corona phase. Perturbations to the corona phase can have a large impact on excitons traveling on the nanotube surface, causing modulations to SWNT fluorescence emission. It is this sensitive relationship between the corona phase and SWNT fluorescence that can be exploited to develop new molecular sensors by incorporating specific binding modalities onto the large surface area of SWNT. Perturbations to

the SWNT corona phase upon binding analyte can lead to changes in the local dielectric environment, charge transfer, or introduce lattice defects, all of which can modulate the fluorescence emission of the SWNTs to serve as a signal transduction mechanism.¹⁴ This approach is used in the development of novel fluorescent sensors for the detection of many different classes of molecules including DNA^{15,16}, glucose¹⁷ and small molecules such as ATP,¹⁸ reactive oxygen species¹⁹ and nitric oxide.^{20,21} However, these approaches are limited in that they rely on the existence of a known binding modality for the target analyte.

Recently, a more generic approach to designing fluorescent sensors was developed using SWNTs non-covalently functionalized with amphiphilic heteropolymers, phospholipids, and polynucleic acids. These molecules adsorb to carbon nanotube surfaces to produce highly stable suspensions of individual SWNTs^{22–25} with unique corona phases that can specifically bind proteins^{26,27} or small molecules including the neurotransmitter dopamine.^{28–30} Engineering the corona phase to disperse SWNTs and specifically bind target analytes is referred to as corona phase molecular recognition (CoPhMoRe).²⁸ The small size, low toxicity, high stability and unbleaching nIR fluorescence of CoPhMoRe SWNT sensors make them excellent candidates for *in vivo* sensing for extended time-resolved measurements.⁶ Recent work has shown their applications in plant tissues for optical detection of reactive nitrogen and oxygen species.³¹ A particularly exciting application for CoPhMoRe SWNT sensors is the potential for label free detection of neurotransmitters such as dopamine *in vivo*, where other techniques, such as electrochemical sensing or immunohistochemistry, suffer from a lack of spatial resolution, temporal resolution, and specificity.

Designing and discovering CoPhMoRe SWNT sensors has so far been restrained by the size and chemical diversity of the dispersant library, limiting the likelihood of finding a sensor for a particular target. To date, researchers have only scratched the surface of available conjugated, co-block, biological and biomimetic polymers that could serve as functionally active dispersants for SWNT sensors. Here, we present different methods for both dispersing SWNTs and characterizing their fluorescence for high throughput screening and for single SWNT sensor analysis. Specifically, we outline the procedure for coating SWNTs with polynucleic acid oligomers using direct sonication as well as how to functionalize SWNT with amphiphilic polymers through surfactant exchange by dialysis. We use (GT)₁₅-DNA and polyethylene glycol functionalized with rhodamine isothiocyanate (RITC-PEG-RITC) as examples. We demonstrate the use of (GT)₁₅-DNA SWNTs as a CoPhMoRe sensor for the detection of dopamine. Lastly, we outline procedures for performing single molecule sensor measurements, which can be used for characterization or single molecule sensing.

PROTOCOL:

Caution: Please consult all relevant material safety data sheets (SDS) before use. Nanomaterials may have additional hazards compared to their bulk material counterpart. Use all appropriate safety practices including engineering controls (fume hood, noise enclosure) and personal protective equipment (safety glasses, goggles, lab coat, full length pants, closed-toe shoes).

1. Preparation of Buffer, Surfactant and Polymer Solutions

1.1) Preparation of 100 mM NaCl solution

1.1.1) Dissolve 584 mg of NaCl in 80 mL of deionized water. Add deionized water to bring total volume to 100 mL.

1.2) Preparation of 3% sodium dodecyl sulfate (SDS) solution

1.2.1) Dissolve 3 g of SDS in 80 mL of deionized water. Add deionized water to bring total volume to 100 mL.

1.3) Preparation of 2% sodium cholate (SC) solution

1.3.1) Dissolve 2 g of sodium cholate hydrate in 80 mL of deionized water. Add deionized water to bring total volume to 100 mL.

1.4) Preparation of imaging buffer (1X Tris: 20 mM Tris, 100 mM NaCl)

1.4.1) Dissolve 22.23 g of Tris base and 58.44 g of NaCl in 500 mL of deionized water using a magnetic stir bar and plate.

1.4.2) Carefully add concentrated HCl until a pH of 8.1 is reached.

1.4.3) Add deionized water to reach a final volume of 1 L.

1.5) Synthesis of RITC-PEG-RITC polymer

1.5.1) Dissolve amine difunctionalized polyethylene glycol (PEG) (5 kDa or 20 kDa, 0.1 mol/L) and rhodamine isothiocyanate (RITC, 0.2 mol/L) in 1 mL of a 1:1 mixture of dichloromethane and dimethylformamide (DMF).

1.5.2) Add 0.2 mol/L of N,N-diisopropylethylamine (DIEA).

1.5.3) After three hours, precipitate with 10X volume of diethyl ether followed by vacuum filtration.

1.5.4) Redissolve in DMF and repeat ether precipitation followed by vacuum filtration.

Note: Other isothiocyanate modified molecules (e.g. fluorescein isothiocyanate, FITC) can be attached to PEG or other amine modified polymers using a similar method.

1.6) Preparation of pegylated-DNA (PEG-DNA)

1.6.1) Combine 100 μ L of Tris(2-carboxyethyl)phosphine hydrochloride (TCEP) solution (0.5 M, pH 7.0) with 44.9 μ L of 5'-thiol-modified DNA (1 mg/10 μ L in 0.1 M NaCl) and add to 4.855 mL of deionized water.

1.6.2) Stir for 1 hour.

1.6.3) Dissolve 500 mg of methoxypolyethylene glycol maleimide in 5 mL of phosphate buffered saline.

1.6.4) Combine DNA and PEG solutions (10 mL total) and stir for 24 hours.

2. Preparation of Single Walled Carbon Nanotube (SWNTs) Suspensions

2.1) Wash of SWNTs to remove catalyst and impurities.

2.1.1) Add 200-300 mg of unwashed SWNTs into a plastic centrifuge tube containing 45 mL of deionized water.

2.1.2) Vortex the solution for 2 minutes and sonicate using a bath sonicator for 5 minutes. Note that sonicator settings vary from instrument to instrument, so check that the optical density of the SWNT solution increases (i.e. turns black) to ensure that the SWNTs are being dispersed.

2.1.3) Centrifuge the solution for 20 min at $16100 \times g$ and discard supernatant.

2.1.4) Add approximately 45 mL of fresh deionized water.

2.1.5) Repeat steps 2.1.2-2.1.4 up to 8 times.

2.1.6) Remove as much water as possible being careful not to disturb pelleted SWNTs and allow SWNT pellet to air dry.

2.2) Nucleic acid suspensions of SWNTs

2.2.1) Dissolve nucleic acids (NA) in 0.1 M NaCl to a concentration of 100 mg/mL.

2.2.2) Remove static electricity from disposable spatula, microcentrifuge tubes and SWNT stock using an anti-static gun. In a fume hood, add 20 μL of the NA solution to 980 μL of 0.1 M NaCl followed by the addition of 1 mg SWNTs.

2.2.3) Using an ultrasonicator with 3 mm diameter tip, sonicate the solution for 10 minutes at 40% amplitude in an ice bath.

2.2.4) Centrifuge the DNA-SWNT solution TWICE for 90 minutes at $16100 \times g$. If using PEG-DNA, purify out excess and unreacted DNA and PEG using a 100 kDa spin-filter. Add enough PBS to fill the spin-filter and spin at $9300 \times g$ for 1.5 min, Repeat this wash step 3 times.

2.2.5) Collect and keep the supernatant, being careful not to disturb the pellet containing CNT bundles and aggregates. Discard the pellet in accordance with institutional hazardous waste procedures appropriate for nanomaterials.

2.2.6) Measure the solution absorbance at 632 nm using a UV/Vis spectrophotometer to determine approximate concentration of suspended SWNTs using $\epsilon=0.036 \text{ L/cm}\cdot\text{mg}$ in accordance with Lambert-Beer's law and the appropriate dilution factor.

2.3) Amphiphilic polymer suspensions of SWNTs

2.3.1) Remove static electricity from disposable spatula, micro centrifuge tubes, and SWNT stock. In a fume hood, add 5 mg SWNTs to 5 mL of 2% SC solution (alternatively, SDS solution can be used).

2.3.2) Using an ultrasonicator with 6 mm diameter tip, sonicate the solution for 1 hour at 40%

amplitude in an ice bath.

2.3.3) Centrifuge sample at 150,000 x g for 4 hours using an ultracentrifuge and carefully collect supernatant.

2.3.4) Dissolve 1 wt % of amphiphilic polymer (e.g. RITC-PEG-RITC) in SC-SWNT solution.

2.3.5) Dialyze the polymer-SC-SWNT solution using a 3.5 kDa dialysis membrane against 1 L of deionized water or buffer for 5 days. Change out the water or buffer after hour 2 and hour 4. A larger dialysis membrane can be used, so long as it allows removal of the surfactant of choice, but retention of both the SWNT and amphiphilic polymer.

2.3.6) Measure the solution absorbance at 632 nm using a UV/Vis spectrophotometer to determine approximate concentration of suspended SWNTs using $\epsilon=0.036 \text{ L/cm}\cdot\text{mg}$ in accordance with Lambert-Beer's law and the appropriate dilution factor.

3. Preparation of Surface Immobilized CoPhMoRe SWNTs

3.1) Preparation of BSA-biotin and Neutravidin stock solutions

3.1.1) Dissolve 10 mg lyophilized BSA-biotin in 1 mL deionized water to make a 10 mg/mL stock solution and store at 4° C.

3.1.2) Dissolve 10 mg NeutrAvidin protein (NAV, deglycosylated avidin protein) in 2 mL of deionized water to make a 5 mg/mL stock solution. Store aliquots at -20° C. Thawed aliquots can be kept for several days at 4° C.

3.2) Prepare BSA-Biotin coated microscope slides

3.2.1) Clean a microscope slide and 0.17 mm cover glass (or as appropriate for the microscope objective) with deionized water, followed by methanol, acetone and a final rinse of deionized water.

3.2.2) Create channels by placing several pieces of double-sided tape approximately 5mm apart on the clean microscope slide. Seal the channels by taking a long glass coverslip and pressing it onto the top of the double sided tape. Be sure it is centered on the microscope slide so the edges of the cover slip and slide are not flush.

3.2.3) Add 100 μL of BSA-Biotin stock solution to 900 μL of 1X Tris buffer to a final concentration of 1 mg/mL.

3.2.4) Flow 50 μL of the BSA-biotin solution into the channel by pipetting the solution into one end and wicking away solution at the other using a tissue. Incubate for 5 minutes followed by 3-5 flushes with 50 μL of 0.1 M NaCl.

3.2.5) Dilute 40 μL of 5 mg/mL NAV stock in 960 μL 1X Tris buffer to a final concentration of 0.2 mg/mL.

3.2.6) Flow 50 μL of the NAV solution into the channel by pipetting the solution into one end and

wicking away solution at the other using a tissue. Incubate for 2-5 minutes followed by 3-5 flushes with 50 μ L of 0.1 M NaCl.

3.2.7) Dilute stock solutions of suspended SWNTs in imaging buffer to a concentration of 1 mg/mL and flow 50 μ L of the solution into the channel and incubate for 5 minutes.

3.2.8) Gently rinse away excess SWNTs using 50 μ L of imaging buffer.

3.3) Preparation of APTES silanized microscope slides

Note: APTES silanized slides allow a way to immobilize negatively charged DNA-wrapped SWNTs to the surface of a glass substrate.

3.3.1) Prepare a 10% solution of (3-Aminopropyl)triethoxysilane (APTES) in ethanol.

3.3.2) Using channels made using the steps outlined in 3.2.2, flow in 100 μ L of 1X Tris buffer.

3.3.3) Flush the channel with APTES solution and incubate for 5 min. Wash with 1X Tris buffer.

3.3.4) Dilute stock solutions of suspended DNA-SWNTs in imaging buffer to a concentration of 1 mg/mL and flow 50 μ L of the solution into the channel and incubate for 5 minutes.

3.3.5) Gently rinse away excess SWNTs using 50 μ L of imaging buffer.

4. Fluorescence Spectroscopy and Microscopy of SWNTs

4.1) nIR Fluorescence Microscopy

4.1.1) Perform imaging of surface immobilized SWNTs using laser excitation and an inverted microscope outfitted with an InGaAs sensor array for imaging.

4.1.2) Direct the laser beam to enter the back illumination port of an inverted microscope using mirrors on adjustable kinematic mounts and a pair of post-mounted irises set to the port height. Ensure the beam is level and straight by confirming it passes through both irises when placed between subsequent mirrors and before it enters the illumination port. If necessary, adjust the beam height using a periscope assembly. Remove any short-pass heat filters on the illumination port that would attenuate the beam.

4.1.3) Insert an appropriate filter cube into the microscope to reflect the excitation light into the objective and collect emission light. This typically consists of a dichroic long pass filter with a cutoff above the excitation wavelength, (e.g. 750 LP) and an emission long pass filter (e.g. 850 LP) to further minimize scattered excitation light from hitting the sensor. Additionally, the emission filter can be chosen to be selective for the emission of SWNTs of a particular chirality.

4.1.4) Perform fine adjustments to the beam alignment by inserting an appropriate alignment cube replacing the objective with two offset, frosted discs containing 1 mm pinholes. Align the beam to the center of both the lower and upper alignment discs.

4.1.5) Attach a 2D InGaAs sensor array to the side imaging port of the microscope using an

appropriate adapter and a 0.5X lens if necessary to accommodate the sensor size.

4.1.6) Using a 100X oil immersion objective (1.4 NA), apply fluorescence-free immersion oil and place the immobilized SWNT sample onto the microscope stage.

4.1.7) Raise the objective until the oil contacts the bottom of the cover glass (#1.5, 170 μm thickness). Make adjustments to the objective collar if necessary for imaging conditions, e.g. temperature, glass thickness etc.

4.1.8) Slowly raise the objective with the excitation source on and monitor the fluorescence signal of the InGaAs camera. Fluorescence intensity should gradually increase as the focal plane approaches the surface and the immobilized sensors come into focus.

4.2) nIR Fluorescence Spectroscopy

Note: Fluorescence spectroscopy can be performed using the same microscope setup but by directing the collected light out of the microscope body and into a spectrometer and InGaAs linear array detector.

4.2.1) Using kinematic mounts and mirrors rated for nIR, direct the light path towards the entrance slit of the spectrometer. Focus the light down to a point onto the entrance slit using a focusing lens (e.g. plano-convex, $f=150\text{mm}$). This alignment is accomplished by attenuating the laser power to $<1\text{mW}$ and replacing the microscope objective with a 1" mirror and using a 50/50 beam-splitting filter cube. The excitation light will then leave the microscope body through the exit port and can be used to adjust the mirrors and lens to focus the beam onto the entrance slit.

4.2.2) After replacing the microscope objective and long pass filter, place a wellplate onto the stage and record the spectra of an in-focus sample using the spectrometer and InGaAs array.

4.3) Measuring the reversible response of (GT)₁₅ DNA-SWNTs to dopamine

4.3.1) Mount sensor coated channels onto the microscope stage and bring into focus using a 100X oil objective and InGaAs camera. Add 50 μL of 100 μM dopamine solution in phosphate buffered saline (PBS) to the flow channel and record the change in fluorescence intensity.

4.3.2) Wash out the dopamine solution using phosphate buffered saline and observe the change in fluorescence of the individual SWNT sensors.

4.4) Well-plate screening for analyte response of CoPhMoRe SWNTs

Note: Screening of different analytes can be performed using a transparent well plate controlled using a motorized stage. An ideal well plate is transparent to visible and IR and has black-sidewalls to minimize crosstalk between the wells.

4.4.1) Pipette equal volumes of suspended SWNT sensor (e.g. 5 mg/L concentration) into each well, enough to cover the bottom of the well uniformly, typically $>100\text{ }\mu\text{L}$ for a 96-well plate and $>30\text{ }\mu\text{L}$ for a 384-well plate.

4.4.2) Pipette analytes (e.g. 2 μL , 100 μM final volume) from screening library into the wellplates.

Prepare each analyte in triplicate to account for potential well-to-well variation, fluctuations of excitation intensity, or temperature.

4.4.3) Raise the objective (e.g. 20X achroplan, 0.45 NA) while monitoring the emission spectra using the spectrometer and InGaAs array. Optimal position of the objective will put the focal plane approximately in the middle of the sample volume in the well, which should correspond to a maximum in measured intensity.

4.4.4) For each sample well, record a 1-10 s exposure to collect a spectrum.

4.4.5) Compare the emission spectra for each well to a control containing just SWNTs without additional analyte to quantify the fluorescence response.

REPRESENTATIVE RESULTS:

SWNTs were suspended in aqueous solution using both surfactants and amphiphilic polymers by direct sonication and by dialysis exchange. Figure 1 shows SWNTs, grown using the iron carbonyl catalyzed method, suspended via direct sonication using SDS and C₂₆-DNA. The optical density of a SWNTs with SDS (or polymer) increases dramatically after sonication and decreases upon removal of aggregates and contaminants through purification by centrifugation (Figure 1). Measurements of absorbance at 632 nm quantified the concentration of suspended SWNT.²⁸

The photophysical properties of the SWNT suspensions are characterized using absorbance and fluorescence spectroscopy. Figure 2 shows the absorbance and fluorescence emission spectra of SWNTs suspended using C₂₆-DNA and (GT)₁₅-DNA respectively. The absorbance spectra are a superposition of the individual absorbance peaks for each distinct chirality of nanotube present in the sample. Similarly, each chirality exhibits its unique fluorescence emission peak. Differences in relative emission peak intensity are a result of differences in the population distribution of chiralities as well as differences in excitation efficiency using the 721 nm laser.

The fluorescence response of (GT)₁₅DNA-SWNTs (where I_0 is the initial SWNT fluorescence intensity and I is the SWNT intensity after dopamine addition) in the presence of different concentrations of dopamine is measured by monitoring the fluorescence of a sample using a spectrophotometer and InGaAs linear array (Figure 3). The fluorescence response is a function of the emission peak (Figure 3a-d), indicating that the response may be chirality specific. The fluorescence of the 1044 nm and 1078 nm peaks increase in intensity 2-fold as dopamine concentration approaches 2 μ M. Figure 3e shows the intensity of the entire emission spectra of PEG-(GT)₁₅ DNA-SWNT increase in response to the addition of dopamine.

Individual SWNTs coated using an alternative DNA sequence, C₂₆-DNA (prepared using the same methods at (GT)₁₅), tethered to the surface of a microscope slide are measured under laser illumination using an InGaAs camera and 100X oil immersion objective (Figure 4). Monitoring single emitters tethered to a surface can be used to verify the reversibility of sensor response by washing target molecules away using buffer solution. Total internal reflection fluorescence (TIRF) microscopy can also be used to image dye-conjugated DNA adsorbed on the SWNTs to quantify the number of DNA molecules attached to each tube through photobleaching

experiments. Figure 4 shows 3 distinct bleaching events of Cy3-labeled DNA inferred from the quantized steps of the fluorescence intensity trace of a single emitter. These results indicate that three DNA molecules are attached to the SWNT.

Figure 1: Polymer and Surfactant Suspended SWNTs.

(a) Photograph of RITC-PEG20-RITC SWNTs suspended using 2% SC at various points of preparation. Left: SWNTs added directly to SC solution prior to sonication. Center: After 10 min of bath sonication followed by 10 min probe tip sonication at 90% amplitude followed by centrifugation. The optical density of the solution increases as SWNTs bundles are dispersed (~100 mg/L SWNT concentration). Right: After dialysis with RITC-PEG20-RITC polymer, the final concentration of RITC-PEG-RITC suspended SWNTs is ~20 mg/L. (b) SWNT suspension yield can vary depending on the polymer used to suspend the SWNT. The optical density of the solution provides a good estimate of solution-phase SWNT concentration. Pictured are different concentrations of (GT)₁₅-DNA suspended tubes at different concentrations. From left to right: 100 mg/L, 10 mg/L, 1 mg/L, 0 mg/L.

Figure 2: Absorption and fluorescence emission spectra of surfactant and polymer suspended SWNTs.

(a) Representative absorbance spectra of SWNTs suspended using (GT)₁₅-DNA by direct sonication. The concentration of SWNT is 10 mg/L. Inset: The UV region of the absorbance spectra shows the characteristic DNA absorbance peak at 260 nm. (b) Absorbance spectra of RITC-PEG20-RITC SWNTs after exchange of SC by dialysis. Inset: Absorbance of a 10X diluted sample of RITC-PEG-RITC SWNTs shows the characteristic absorbance of rhodamine. (c) Representative nIR emission spectra of SWNTs suspended using (GT)₁₅-DNA by direct sonication (785 nm excitation).

Figure 3: Fluorescence detection of dopamine using (GT)₁₅-DNA wrapped SWNTs.

(a) Fluorescence response of (GT)₁₅-DNA wrapped SWNTs to the addition of dopamine. Samples of sensors at a concentration of 5 mg/L were excited using a 500 mW, 721 nm CW laser. The integrated fluorescence of sensor emission from 900-1350 nm increases with added dopamine concentration 1 μ M to 250 μ M. (b) Fluorescence emission spectra of PEG-(GT)₁₅-DNA wrapped SWNTs before and after addition of dopamine. The concentration of sensor is 10 mg/mL to which dopamine was added to a final concentration of 100 μ M. The samples were excited using a 500 mW, 721 nm CW laser. The two highest intensity peaks approximately double in intensity after addition of dopamine.

Figure 4: Fluorescence imaging of single surface-immobilized SWNTs.

(a) Fluorescence emission of individual C₂₆-DNA-SWNT (red arrows) immobilized onto a silica cover slip (#1.5) using an APTES silanization procedure and imaged using a 2D InGaAs sensor array, inverted microscope with a 100X oil immersion objective (plan apochromat, 1.4 NA), and a 500 mW, 721 nm CW laser. (b) Fluorescence bleaching experiment of C₂₆-Cy3 DNA-SWNTs tethered to a surface using APTES. The DNA strands are 3' terminally-labeled with Cy3 prior to tube suspension. Tracking the incremental step-wise photobleaching (red fitted trace) of individual sensors is used to determine the number of DNA molecules adsorbed onto the surface of the SWNTs. Images were acquired using an inverted microscope in TIRF mode with a 100X

oil immersion objective (plan apochromat, 1.4 NA), and 561 nm laser excitation. Scale bar: 10 μm .

DISCUSSION:

SWNTs are readily suspended in aqueous solution via direct sonication with SDS or ssDNA, as indicated by an increase in optical density provided by the colloidal dispersion of the resulting SWNT-polymer hybrid. SDS and ssDNA disperses and solubilizes bundles of SWNTs by adsorbing onto the SWNT surface through hydrophobic or π - π interactions. Additionally, other polymers, such as genomic DNA, amphiphilic polymers, conjugated polymers and lipids, can be adsorbed onto the surface of SWNTs by dialysis of samples suspended using SC or SDS. Hydrophilic polymers, such as PEG, can be end-modified with hydrophobic “anchors” such as RITC or FITC to enable surface adsorption of the block copolymer. For polymers that are susceptible to shearing or degradation upon exposure to the high powers of probe-tip sonication, or for polymers with low binding affinities to SWNT, dialysis is the best method to produce a stable SWNT-polymer suspension. Following polymer encapsulation, centrifugation removes large SWNT bundles, amorphous carbon, residual metal catalyst, and other insoluble contaminants to leave a uniformly dispersed sample. Typical final concentrations of dispersed SWNTs after centrifugation range between 10-100 mg/L.

Sonication power and duration can be adjusted and optimized for the particular choice of dispersant. This is a critical step in the procedure for coating SWNTs because too little or weak sonication can result in poor dispersal, while too much or too powerful sonication can lead to poor fluorescence. Typically, lower powers are necessary to minimize damage when using polymers susceptible to shearing such as DNA. Longer sonication durations or higher intensities can lead to reduction in the length of SWNTs, where SWNTs below the ~ 100 nm exciton recombination length become non-fluorescent. The size of the sonicator probe tip should also match the sample volume to avoid splashing and foaming for optimal results (typically provided by manufacturer). Avoid touching the probe tip to the sides of the container and place the solution on an ice block to minimize heating of the solution. Once dispersed, solutions of SWNTs are stable at room temperature indefinitely.

Absorbance and emission spectra of solutions of dispersed generated SWNTs contain multiple peaks, indicating the presence of a mixture of dispersed SWNTs of different chiralities. Alternative methods of SWNT generation or purification methods can change the chirality distribution, leading to different peak excitation and emission fluorescence spectra. Additionally, different synthesis methods can yield samples of SWNTs with different chirality population distributions. For example: cobalt-molybdenum catalyst grown SWNTs are rich in (6,5) chirality, while high pressure with iron carbonyl catalyst grown SWNTs are rich in (7,6) chirality, leading to differences in the absorption and photoluminescence spectra.

Certain adsorbed polymers enable the specific detection of analytes by modulating the fluorescence emission of the SWNT by changing the local environment at the tube surface. This approach offers the distinct advantage over covalent attachment of binding moieties by not permanently disrupting the SWNT lattice, which can reduce fluorescence emission intensity.^{6,32-34} Additionally, the CoPhMoRe approach has the potential for developing antibody-free sensors for targets where there may not already exist a known binding moiety.²⁸ Specifically, (GT)₁₅-

DNA has been shown to selectively enhance the fluorescence emission of SWNTs in the presence of dopamine, enabling its use as a dopamine sensor. Bulk fluorescence measurements show an increase of as much as 80% in peak emission for certain SWNT chiralities. Immobilizing (GT)₁₅-DNA SWNTs on a glass slide enables fluorescence response measurements of individual (GT)₁₅-DNA functionalized SWNT, showing that fluorescence can increase by over 3-fold for single SWNT sensors in the presence of dopamine, without any appreciable photobleaching, under continuous laser illumination. The interaction between dopamine and the SWNT sensor is reversible, as evident by the recovery of the original fluorescent signal after washing dopamine out of the microfluidic chamber with buffer. Additionally, single-molecule measurements can be a powerful characterization technique for quantifying polymer adsorption (Figure 4) or the kinetics of binding events. Also, ratiometric sensing can be achieved by chemically isolating a singular SWNT chirality with a unique excitation-emission peak, functionalizing it with (GT)₁₅-DNA, and isolating a second SWNT chirality to be insensitive to dopamine. Monitoring both SWNT chiralities provides a steady control fluorescence channel that can be compared to the modulating fluorescence of the (GT)₁₅-DNA SWNTs. Combining different polymers (e.g. polyethylene glycol and (GT)₁₅-DNA) can add additional functionality such as modifying diffusivity or cellular uptake characteristics, properties that are critical when performing *in vivo* experiments.

Currently, a limitation of the CoPhMoRe approach to sensor development include polymer library development. Because binding moieties are not known *a priori*, developing a sensor for a particular target can be time intensive and require a large number of chemically varied polymers for constructing the sensor library for screening. Additionally, stability and compatibility of sensors in *in vivo* environments can vary from sensor to sensor. However, once a candidate sensor has been identified, further modification strategies can be employed to optimize properties as necessary for *in vivo* applications.

Herein, we have demonstrated a methodology for dispersing SWNTs in aqueous solutions applicable to a wide variety of dispersing agents. This approach can be used to create libraries of dispersed SWNTs for the discovery of novel new nIR sensors for small molecules and biological markers. Of particular interest are sensors for the detection of neurotransmitters, which could enable real time, spatially precise detection of these molecules in complex biological environments.

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DISCLOSURES:

The authors have nothing to disclose.

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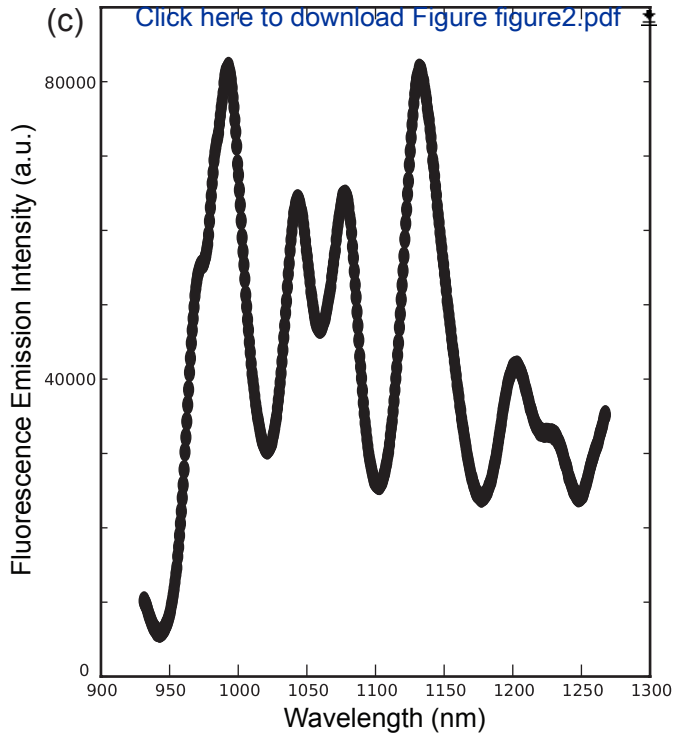
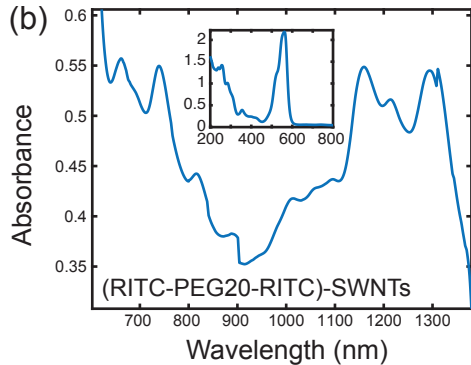
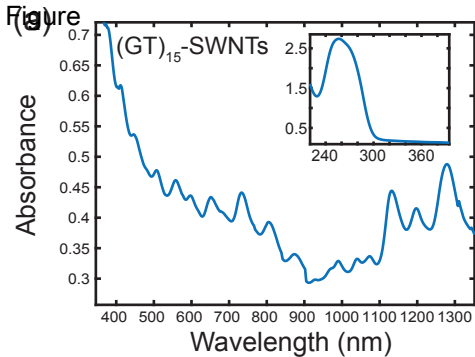
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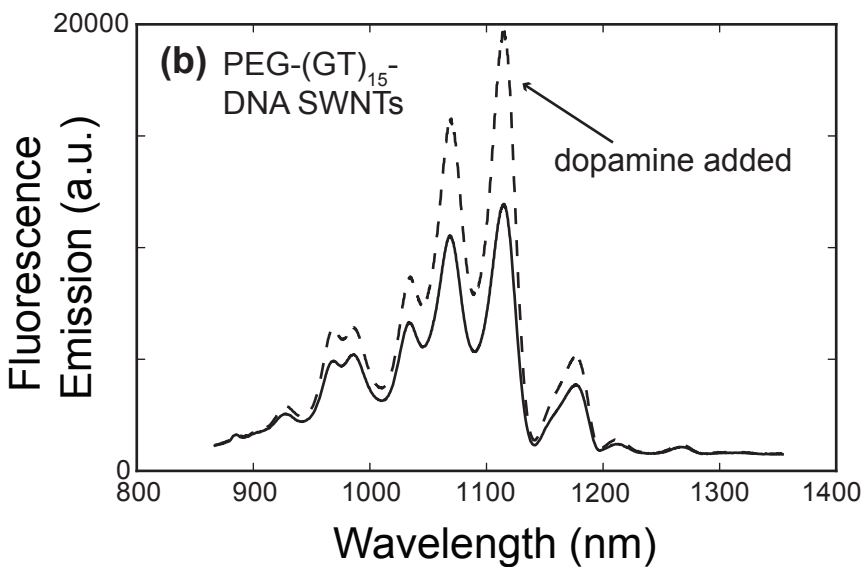
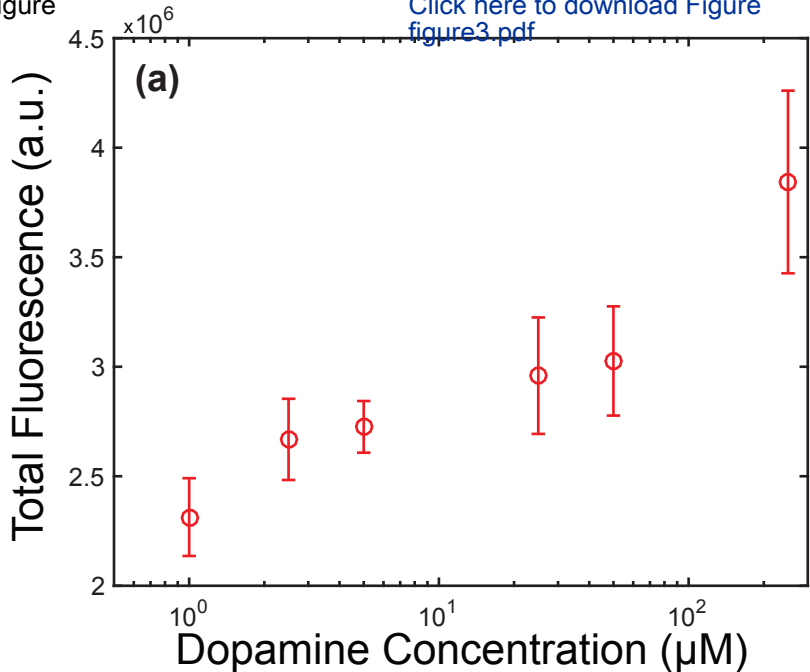
Figure



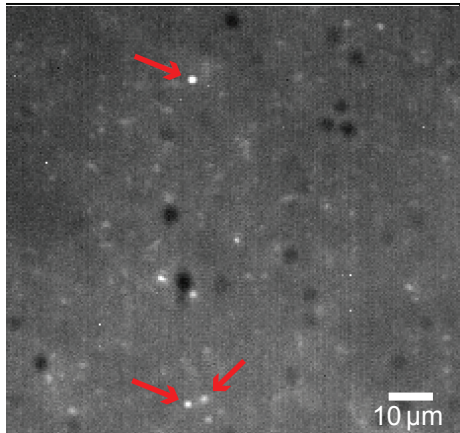
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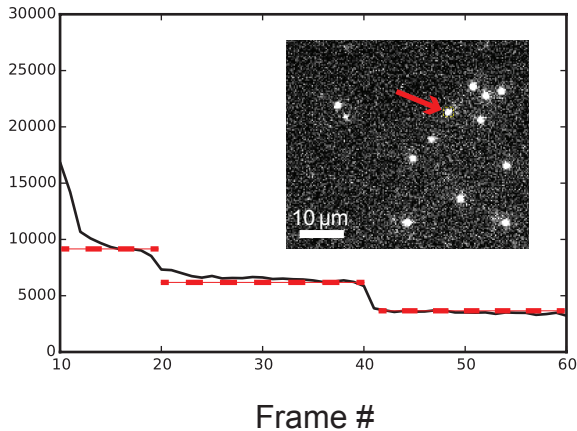


(a)



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Fluorescence Intensity (a.u.)



Name of Material/ Equipment	Company	Catalog Number
sodium chloride	Fisher Scientific	S271-1
sodium dodecyl sulfate	Sigma Aldrich	L6026
sodium cholate hydrate	Sigma Aldrich	C6445
tris base (Trizma base)	Sigma Aldrich	93362
hydrochloric acid	Fisher Scientific	A144-212
Amine-PEG-amine,NH2-PEG-NH2	Nanocs Inc	PG2-AM-5k
rhodamine B isothiocyanate	Sigma Aldrich	283924
fluorescein isothiocyanate	Sigma Aldrich	F7250
dichloromethane	Sigma Aldrich	676853
dimethylformamide	Sigma Aldrich	D4551
N,N-diisopropylethylamine	Sigma Aldrich	D125806
diethyl ether	Sigma Aldrich	673811
Tris(2-carboxyethyl)phosphine hydrochloride	Sigma Aldrich	C4706
5'-thiol-modified DNA	Integrated DNA Technologies	
methoxypolyethylene glycol maleimide	Sigma Aldrich	63187
100k Da spin filters	Millipore	
HiPCO Super purified single walled carbon nanotubes	Integris	HiPco SuperPurified
phosphate buffered saline	Sigma Aldrich	P5493
anti static gun	Milty	Milty Zerostat 3
centrifuge	Eppendorf	5415 D
ultra sonicator	Cole Parmer	CV18
	Thermo	Slide-A-Lyzer G2
dialysis cassettes	scientific	87722

BSA-biotin	Thermo scientific	29130
Neutravidin protein	Thermo scientific	31000
(3-Aminopropyl)triethoxysilane (APTES)	Sigma Aldrich	440140
inverted microscope	Zeiss	Axio Observer.Z1
kinematic mirrors	ThorLabs	KM200-E03
periscope	ThorLabs	RS99
immersion oil	Zeiss	Immersol 518f
100X objective	Zeiss	Plan-apochromat 100X oil, 1.4NA, PH3, 420791-9911-000
		N-Achroplan 0.45
		NA, 420953-9901-
20X objective	Zeiss	000
	Healthrow	
cover glass	Scientific	HS159879H
dopamine hydrochloride	Sigma Aldrich	H8502
	Princeton	
infrared 2d array camera	Instruments	NIRvana
	Princeton	
infrared 1d sensor array	Instruments	PyLoN IR
	Princeton	
nIR spectrograph	Instruments	SCT-320
planoconvex lens	ThorLabs	LA1384
wellplates (glass bottom)	Corning	4580



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Please use the attached manuscript as it represents the most current version of the manuscript and is the one that was sent out for peer review.

1. Formatting: -Please define all abbreviations at first occurrence (ie nIR, CoPhMoRe, TIRF etc.).
-Please use “x g” rather than cfg.

We have modified the manuscript to properly define all acronyms.

2. Please fix the following for grammar: -2.1 – “growth are” -2.2.6, 2.3.6 – Should be “spectrophotometer” -Line 434 – “FITC add binding functionality”

These grammatical errors are now corrected in the manuscript.

3. Additional detail is required: -1.5.3 – Is the precipitate filtered first before dissolving in DMF? -2.1.2 – What are the sonication settings? -2.2.5 – What happens to the pellet? Is this discarded?

In the manuscript, we clarify that the precipitate should be filtered before redissolving. Additionally, we have included more information regarding sonication settings as well as whether the pellet should be discarded.

4. Please remove commercial branding:

-CoMoCAT – 2.1 -HiPCO – 2.1, 2.2.2, 2.3.1, Figure 1 and 2 legends, Discussion -Neutravidin - 3.1, 3.1.2, 3.2.5, 3.2.6 – Please use a generic term or abbreviate as NAV after first occurrence.

Although the terms CoMoCAT and HiPCO are trademarked, they are the colloquial terms used exclusively in academic literature to refer to the types of SWNTs produced by particular manufacturing processes. In the manuscript, we clarify by clearly defining the meaning of each acronym on page 9 (Results section) and on page 12 (Discussion section) of the preprint. Additionally, we added a definition of Neutravidin (page 6, section 3.1.2) and abbreviated subsequent mention using the acronym as suggested.

5. Results: -Please discuss what the results mean in the results section. What is the interpretation of the data shown in Figures 3 & 4? -Figure 3 - Please define error bars in the figure legend (SD, SEM, etc.).

We have included more discussion regarding the interpretation of the results presented in Figures 3 and 4. We have also included a label for the error bars in Figure 3.

6. Discussion: Please discuss the significance with respect to alternative methods and provide independent citations. Please also discuss the critical steps and limitations of the method.

We more clearly outline the advantages of this method compared to covalent modification and included several independent citations on page 12 (Discussion section). Additionally, we more clearly highlight sonication as the critical step in the discussions section on page 11. A brief discussion of current limitations was also added.

Reviewers' comments: Reviewer #1: Manuscript Summary: In this method manuscript Jackson T et al. described a protocol to engineer the corona phase of nIR fluorescent single walled carbon nanotubes using amphiphilic polymers and DNA to develop sensors for molecular targets without known recognition elements.

This is manuscript would have wide range of readership as researchers are keen to investigate on CNT for biological application, especially for imaging. Therefore CNT still is a growing platform. Although some other researcher already have established protocol for CNT suspensions, however amphiphilic polymers suspension would be interesting. Therefore I would highly recommend this method manuscript for publication.

Major Concerns:

Authors used absorbance at 632 nm quantified the concentration of suspended SWNT according Ref. 14. However, CNT (6,5) has strong abs at 990nm rater than 632nm. Zheng M., Nano Res (2008) 1: 185-194 and in SI of Mangalum A., J. Am. Chem. Soc., 2013, 135 (7), pp 2451-2454 Therefore I would highly recommend having wide range (200-1200nm) abs spectra in fig. 2a

We appreciate the detailed feedback and have updated the absorbance spectra plots to the range 200-1200nm. Regarding the absorbance wavelength used herein, absorbance at 990 nm is specific to that particular chirality of SWNT. In the methods presented here, we use mixed chirality samples, which partially obscures the absorbance peaks of individual chiralities. Additionally, we are interested in a measurement of total SWNT concentration, not just a particular chirality. For this reason, we use absorbance at 632 nm (a global absorbance minimum) and the empirically derived extinction coefficient (Zhang *et al.* 2013) to estimate total SWNT concentration for our multi-chirality samples.

Minor Concerns:

Add references for DNA-CNT 1. Zheng M., Nano Res (2008) 1: 185-194 2. Mangalum A., J. Am. Chem. Soc., 2013, 135 (7), pp 2451-2454

We have added these references to carbon nanotubes in the introduction as they relate to SWNT chirality separations that would be of interest to readers wishing to use ratiometric single-chirality sensing.

Additional Comments to Authors:

N/A

Reviewer #2:

Manuscript Summary:

The authors present a protocol for engineering the corona phase of NIR fluorescent single-walled carbon nanotubes (SWNTs) using amphiphilic polymers and DNA to develop sensors for molecular targets without known recognition elements. The corona phase molecular recognition using SWNTs is timely and very interesting. Along with the unique optical properties of SWNTs, this technique would find wide applications in the future. But, there are several major concerns that should be addressed in the article.

Major Concerns:

-The Introduction should be rewritten. The Introduction doesn't well present the current development of SWNT sensing technologies. The corona phase molecular recognition is not clearly or adequately described. For example, the advantages described for the corona phase molecular recognition is the general advantages of SWNTs rather than those of the corona phase molecular recognition. Recent key references are not cited; most references cited are published before 2005.

The introduction has been rewritten to better review SWNT sensing technologies as well as to more clearly introduce and explain corona phase molecular recognition.

-The Introduction should better describe what techniques are described in the protocol. The authors prepared several molecules to disperse SWNTs. But, it is not clear why they choose them and how these molecules make SWNTs sensors. -The advantages of the corona phase molecular recognition over the conventional molecular recognition are not well described. The limitations of the corona phase molecular recognition are not described.

We have improved the introduction by describing the procedures and techniques used in the protocol. Additionally, we discuss the specific molecules used and how one in particular is used for dopamine detection. The introduction also now includes a detailed discussion of the advantages as well as limitations of this method.

-Higher quality images are required for Figure 1 (e.g., Figure 1 right).

We have updated the images in Figure 1 to better represent methods outlined in the protocol as well as improved the image quality.

-Full range UV-Vis-NIR absorbance spectra (e.g., 400 to 1400 nm range) would be desired to show not only the concentration of SWNTs but also how well SWNTs are dispersed.

We have acquired new absorbance spectra and now include a full UV-Vis-NIR absorbance for both GT15-DNA SWNTs and RITC-PEG-RITC tubes (Figure 2).

-Sensing results shown in Figure 3 are confusing.

We have remade Figure 3 to more clearly show how dopamine affects fluorescence intensity of the (GT)15-PEG-SWNT sensor.

-As the focus of the protocol is sensing, it would be better to show the results of sensing experiments than the results of bleaching experiments in Figure 4.

We believe that our new Figure 3 clearly emphasizes the sensing of dopamine using GT15-DNA SWNT sensors. The objective of Figure 4 is to provide examples of single molecule measurements and demonstrate how this approach can also be used for characterization of polymer wrapped SWNTs, which are commonly used to determine why certain sensors work while others don't. We have edited the introduction and discussion sections to outline these motivating factors more clearly.

-One of the key molecules prepared in the protocol is RITC-PEG-RITC polymer. The results with RITC-PEG-RITC polymer are not shown.

Results for RITC-PEG-RITC SWNTs are now included in Figure 1 and Figure 2.

-It is not clear why the authors selected and prepared C26 and (GT)15-DNA SWNTs. How do they function differently?

We present (GT)15-DNA as a sensor for dopamine. Figure 4 shows representative data for single molecule measurements of SWNTs coated in a different, unrelated DNA sequence (C26) using the same preparation methods as the (GT)15-DNA. This particular set of data is unrelated to the GT15-DNA sensors and is instead intended to demonstrate the ubiquity of this method to encapsulating SWNTs with any arbitrary DNA sequence. We have modified the results and discussion sections to make this more apparent to the reader.

Minor Concerns:

-Line 58: In "fluorescence emission that is stable, biocompatible and sensitive," it is not clear what "fluorescence emission that is biocompatible" means.

We have rewritten this sentence and resolved this ambiguity.

-Line 59: It would be better to be more specific for what "coating" means.

We have replaced the word "coating" with "non-covalently functionalized" and in the next sentence clarify that these molecules are adsorbed onto the surface of the SWNT.

-Line 68: The definition of SWNTs, cylinders of graphene, is not clear.

We have rewritten the first several sentences of the introduction to more clearly define SWNTs.

-Line 69: It may be better to change "This includes" to "These include."

This has been rewritten to improve the grammar.

-Line 72: "owing to their structure" is not clear. This statement may not represent the current state of the field. For examples, the references cited (references 5 and 6) are published in 2000 and 2002.

We now cover, in more detail, the general properties of SWNTs as well as include numerous additional references to provide a better background to the field. This now includes more references to recent literature.

The use of "tubes" is not clear; it would be better to use a consistent terminology for SWNTs. It seems like that the authors use "tubes" for microcentrifuge tubes in the protocol.

We have replaced mention of "tubes" with "SWNTs" for clarity. Additionally, we now specifically use the term "microcentrifuge tubes" in the protocol to eliminate potential confusion.

-Line 73: The sentence is not clear, including the use of "potential."

This sentence has been rewritten to clearly outline how the SWNT sensors function and their applications.

-Line 85: The sentence starting with "perturbations" may not adequately describe the concept of corona phase molecular recognition.

We have rewritten and expanded the introduction to more clearly describe the concept of corona phase molecular recognition.

-Line 91: The sentence starting with "additionally" needs references.

An appropriate reference has been added to this sentence that reviews the applications of SWNTs in biological sensing.

-Lines 95 to 98: These statements are not clear or adequate.

These sections pertaining to the above comments have been completely rewritten to improve content as well as clarity. This includes a better explanation of corona phase molecular recognition and addition of current references.

-Page 3: What is the difference between ultrapure water and deionized water?

We have replaced instances of "ultrapure" with "deionized" for consistency.

-Line 184: The use of "tubes" can be confusing.

We have replaced mention of "tubes" with "SWNTs" for clarity. Additionally, we now specifically use the term "microcentrifuge tubes" in the protocol to eliminate potential confusion.

-The use of terms is not consistent (e.g., x g and cfg).

To correct this inconsistency, all instances were replaced with "x g".

-Line 204: The use of "static" is not clear.

We are referring to static electricity. The manuscript has been modified to make this more clear.

-Line 224: What is CoPhMoRe?

CoPhMoRe is the acronym for Corona Phase Molecular Recognition. The manuscript now more clearly defines this acronym in the introduction.

Additional Comments to Authors:

N/A