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Salinity-dependent toxicity assay of silver nanocolloids using medaka eggs

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Abstract:	<p>Salinity is an important characteristic of the aquatic environment. For aquatic organisms it defines the habitats of freshwater, brackish water, and seawater. Tests of the toxicity of chemicals and assessments of their ecological risks to aquatic organisms are frequently performed in freshwater, but the toxicity of chemicals to aquatic organisms depends on pH, temperature, and salinity. There is no method, however, for testing the salinity dependence of toxicity to aquatic organisms. Here, we used medaka (<i>Oryzias latipes</i>) because they can adapt to freshwater, brackish water, and seawater. Different concentrations of embryo-rearing medium (ERM) (1×, 5×, 10×, 15×, 20×, and 30×) were employed to test the toxicity of silver nanocolloidal particles (SNCs) to medaka eggs (1× ERM and 30× ERM have osmotic pressures equivalent to freshwater and seawater, respectively). In six-well plastic plates, 15 medaka eggs in triplicate were exposed to SNCs at 10 mg L⁻¹ in different concentrations of ERM at pH 7 and 25 C in the dark.</p> <p>We used a dissecting microscope and a micrometer to measure heart rate per 15 s and eye diameter on day 6 and full body length of the larvae on hatching day (section 4). The embryos were observed until hatching or day 14; we then counted the hatching rate every day for 14 days (section 4). To see silver accumulation in embryos, we used inductively coupled plasma mass spectrometry to measure the silver concentration of test solutions (section 5) and dechorionated embryos (section 6). The toxicity of the SNCs to medaka embryos obviously increased with increasing salinity. This new method allows us to test the toxicity of chemicals in different salinities.</p>
Author Comments:	<p>August 25, 2015</p> <p>Ref: Salinity-dependent toxicity assay of silver nanocolloids using medaka eggs</p> <p>By : Chisato Kataoka, Kashiwada Shosaku</p>

	<p>Dear Editor-in-Chief:</p> <p>We would like to submit the following revised manuscript entitled "Salinity-dependent toxicity assay of silver nanocolloids using medaka eggs" for consideration for publication in JoVE. Thank you very much for your useful comments. According to your comment we sincerely revised the manuscript.</p> <p>Thank you for your consideration.</p> <p>Sincerely,</p> <p>Shosaku Kashiwada, Ph.D. Professor Department of Life Sciences, Toyo University, 1-1-1 Izumino, Itakura, Gunma 374-0193, Japan Phone: +1-(276) -82-9029/9337 Fax: +1-(276) -82-9029 Email: kashiwada@toyo.jp</p>
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August 25, 2015

Ref: Salinity-dependent toxicity assay of silver nanocolloids using medaka eggs

By : Chisato Kataoka, Kashiwada Shosaku

Dear Editor-in-Chief:

We would like to submit the following revised manuscript entitled “**Salinity-dependent toxicity assay of silver nanocolloids using medaka eggs**” for consideration for publication in JoVE. Thank you very much for your useful comments. According to your comment we sincerely revised the manuscript.

Thank you for your consideration.

Sincerely,

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Keywords:

aquatic toxicology; medaka; nanomaterials; nanotoxicology; salinity; seawater; silver nanocolloids

Short Abstract:

Embryonic stages are the most susceptible to xenobiotics. Although chemical toxicity depends on salinity, no method exists to test the salinity dependence of toxicity to aquatic organisms. Here, we describe a new and high-throughput method for determining the salinity dependence of toxicity to aquatic embryos.

Long Abstract:

Salinity is an important characteristic of the aquatic environment. For aquatic organisms it defines the habitats of freshwater, brackish water, and seawater. Tests of the toxicity of chemicals and assessments of their ecological risks to aquatic organisms are frequently performed in freshwater, but the toxicity of chemicals to aquatic organisms depends on pH, temperature, and salinity. There is no method, however, for testing the salinity dependence of toxicity to aquatic organisms. Here, we used medaka (*Oryzias latipes*) because they can adapt to freshwater, brackish water, and seawater. Different concentrations of embryo-rearing medium (ERM) (1×, 5×, 10×, 15×, 20×, and 30×) were employed to test the toxicity of silver nanocolloidal particles (SNCs) to medaka eggs (1× ERM and 30× ERM have osmotic pressures equivalent to freshwater and seawater, respectively). In six-well plastic plates, 15 medaka eggs in triplicate were exposed to SNCs at 10 mg L⁻¹ in different concentrations of ERM at pH 7 and 25 °C in the dark.

We used a dissecting microscope and a micrometer to measure heart rate per 15 s and eye

diameter on day 6 and full body length of the larvae on hatching day (section 4). The embryos were observed until hatching or day 14; we then counted the hatching rate every day for 14 days (section 4). To see silver accumulation in embryos, we used inductively coupled plasma mass spectrometry to measure the silver concentration of test solutions (section 5) and dechorionated embryos (section 6). The toxicity of the SNCs to medaka embryos obviously increased with increasing salinity. This new method allows us to test the toxicity of chemicals in different salinities.

Introduction:

Since the establishment of the Organisation for Economic Co-operation and Development (OECD) test guidelines for testing chemicals in 1979, 38 test guidelines have been published in Section 2 of the guidelines, Effects on Biotic Systems¹. All of the aquatic organisms tested have been from freshwater habitats, namely freshwater plants; algae; invertebrates such as daphnia and chironomids; and fishes such as medaka, zebrafish, and rainbow trout. Compared to saltwater environments, freshwater environments are more directly affected by human economic and industrial activities. Therefore, freshwater environments have been prioritized for testing because they are at higher risk from pollution.

In coastal areas, including estuaries, salinities vary between brackish water and seawater conditions, and these areas are often polluted by industrial activity². Coastal areas and their associated wetlands are characterized by high ecological biodiversity and productivity. Coastal ecosystems must therefore be protected from chemical pollution. However, there has been limited ecotoxicological research in brackish water and seawater habitats.

Sakaizumi³ studied the toxic interactions between methyl mercury and salinity in Japanese medaka eggs and found that increasing the osmotic pressure of the test solution enhanced the toxicity of the methyl mercury. Sumitani *et al.*⁴ used medaka eggs to investigate the toxicity of landfill leachate; they found that the osmotic equivalency of leachate to the eggs was the key to inducing abnormalities during embryogenesis. In addition, Kashiwada⁵ reported that plastic nanoparticles (39.4 nm in diameter) easily permeated through the medaka egg chorion under brackish conditions (15× embryo rearing medium (ERM)).

A typical small fish model, the Japanese medaka (*Oryzias latipes*) has been used in basic biology and ecotoxicology⁶. Japanese medaka can live in conditions ranging from freshwater to seawater because of their highly developed chloride cells⁷. They are therefore likely to be useful for testing in conditions with a wide range of salinities.

Protocol:

The Japanese medaka used in this study were treated humanely in accordance with the institutional guidelines of Toyo University, with due consideration for the alleviation of distress and discomfort.

1. Silver nanocolloids (SNCs)

1.1) Purchase purified SNCs (20 mg L⁻¹, 99.99% purity, particle mean diameter about 28.4 ± 8.5 nm suspended in distilled water).

1.2) Validate the purity and concentration of the silver by inductively coupled plasma mass spectrometry (ICP-MS) analyses according to operating manual⁸. The pretreatment method for ICP-MS analyses is described in section 7.

2. Preparation of SNC solutions (mixtures of silver colloids and Ag⁺) with different salinities

2.1) Prepare 60× ERM consisting of 60 g NaCl, 1.8 g KCl, 2.4 g CaCl₂·2H₂O, and 9.78 g MgSO₄·7H₂O in 1 L of ultrapure water; adjust the pH to 7.0 with 1.25% NaHCO₃ in ultrapure water.

2.2) Stir the ERM solution at 25 °C overnight.

2.3) Mix SNCs with diluted ERM. Prepare 40 mL of each SNC–ERM mixed solution. The final concentration is 10 mg L⁻¹ of SNCs in different concentrations of ERM (1×, 5×, 10×, 15×, 20×, or 30×).

2.4) Adjust pH of the SNC–ERM mixed solution to 7.0 with 0.625% NaHCO₃ in ultrapure water. pH adjustment is very important in preparing the SNC solution, because Ag⁺ release is facilitated by acidic conditions⁹.

2.5) Use AgNO₃ as a reference compound for SNCs.

2.5.1) Mix AgNO₃ with diluted ERM. Prepare 40 mL of AgNO₃–ERM mixed solution at an AgNO₃ concentration of 15.7 mg L⁻¹ (10 mg L⁻¹ silver) in different concentrations of ERM (1×, 5×, 10×, 15×, 20×, or 30×).

Note: To examine silver colloid toxicity, AgNO₃ solution, which is a source of soluble silver, is used as a reference compound for SNCs, which are a mixture of silver colloids and soluble silver.

3. Medaka culture and egg harvesting

3.1) Obtain the medaka (*O. latipes*) (orange-red strain) (60 males and 60 females).

3.2) Culture medaka as groups (20 males and 20 females as one group) in 1× ERM in 3-L tanks by using a medaka flow-through culturing system.

3.2.1) Culture at the following conditions:

pH range of the culture medium: 6.2 to 6.5

light:dark cycle: 16:8 h

temperature of the culture medium: 24 ± 0.5 °C

osmotic pressure of the culture medium: 257 mOsm

3.3) Feed medaka on *Artemia salina* nauplii at 10:00 h (once a day) and feed an artificial dry fish diet at 09:00, 11:00, 13:00, 15:00, and 17:00 h (five times a day).

3.3.1) Obtain *A. salina* nauplii.

3.3.2) Prepare 5 L of a 3.0% salt solution in a plastic beaker.

3.3.3) Add 30 g of brine shrimp eggs to the salt solution in the beaker.

3.3.4) Incubate the eggs at 25 °C for 48 h with bubbling (4 L min⁻¹) using an aeration pump.

3.3.5) After 48 h, stop the bubbling.

3.3.6) Allow the solution to stand for 5 to 10 min to separate the hatched *A. salina* nauplii (lower part of the solution) from the unhatched eggs and eggshells (upper part of the solution).

3.3.7) Remove the upper layer of the solution by decantation.

3.3.8) Filter the lower portion of the solution through a sieve with openings of 283 µm, and collect the nauplii that pass through on a net with openings of 198 µm.

3.3.9) Feed the nauplii to the medaka within 6 h.

3.4) After the female medaka have spawned, remove the external egg clusters gently from the females' bodies or collect the eggs from the bottom of the fish tank by using a small net (net size 5 cm × 5 cm, hole size 0.2 mm × 0.2 mm).

3.5) Rinse the egg cluster with flowing tap water for 5 s.

3.6) Add all of the rinsed egg clusters to 30× ERM solution.

3.7) Remove the clusters from the solution after 1 min and place the egg clusters between dry paper towels and roll gently.

3.8) Put the eggs back into the 30× ERM.

3.9) Select fertilized eggs under a dissecting microscope.

3.10) Place selected 810 eggs in 1× ERM in six-well plastic plates by using forceps.

3.11) Incubate the eggs at 25 ± 0.1 °C in an incubator until developmental stage 21. (Developmental stages of the medaka embryos were defined from the work of Iwamatsu¹⁰.)

3.12) Pick out incubated eggs at developmental stage 21 under a dissecting microscope.

3.13) Rinse selected eggs with 1× ERM.

3.14) Subject the rinsed eggs to exposure experiments (section 4).

4. Toxicity testing of SNCs or AgNO₃ at different ERM salinities

4.1) Rinse medaka eggs (stage 21) three times with test solution [SNCs (10 mg L⁻¹) or AgNO₃ (15.7 mg L⁻¹ as 10 mg L⁻¹ silver) at each concentration of ERM (1×, 5×, 10×, 15×, 20×, or 30×) at pH 7]. As controls, use eggs in 1× to 30× ERM at pH 7.

4.2) Add 15 rinsed eggs to 5 mL of each test solution in six-well plastic plates. (Perform the exposure experiments three times for SNC or AgNO₃ toxicity testing using each test solution.)

4.3) Wrap the plates in aluminum foil.

4.4) Incubate the wrapped plates at 25 °C in the dark until hatching or for 14 days.

4.5) Observe the exposed eggs every 24 h for biological changes and dead eggs (**Figures 1 and 2**).

4.6) Exchange the test solutions every 24 h.

4.7) Perform observations as follows.

4.7.1) On day 6 of exposure, count the heart rate (per 15 s) of medaka embryos under a dissecting microscope by using a stopwatch (**Figure 3a**).

4.7.2) On day 6 of exposure, measure the eye size (diameter) of medaka embryos under a dissecting microscope by using a micrometer (**Figure 3b**).

4.7.3) On hatching day, measure the full body lengths of larvae under a dissecting microscope by using a micrometer (**Figure 3c**).

4.7.4) Count the total number of exposed eggs that hatch over the 14 days (**Figure 3d**).

5. Isolation of soluble silver from SNC solution, and silver analysis

5.1) Isolate soluble silver from each SNC solution (a mixture of silver colloids and soluble silver) by filtering through a 3-kDa membrane filter at 14,000 xg and 4 °C for 10 min. Use a 3-kDa membrane filter to isolate soluble silver from the SNCs, because the reported mean diameter of aggregated SNCs in 1× ERM is 67.8 nm¹¹ and that of Ag⁺ is 0.162 nm¹²; the 3-kDa membrane

excludes particles with diameters of 2 nm or more¹³.

5.2) Measure the silver concentration in 50 µL of filtered solution (= the soluble silver concentration) by ICP-MS analysis (**Figure 3e**) according to the ICP-MS operating manual⁸. The pretreatment method for the ICP-MS analyses is described in section 7.

6. Measurement of silver bioaccumulation in medaka embryos

6.1) Expose medaka eggs (stage 21) to SNCs as described in section 4.

6.2) On day 6 of exposure, remove chorion from the egg (i.e., dechorion) by using medaka hatching enzyme according to the protocol described in the *Medaka Book*¹⁴.

6.3) Measure the silver concentration of the dechorionated eggs by ICP-MS analysis according to the ICP-MS operating manual⁸ (**Figure 3f**). The pretreatment method for the ICP-MS analyses is described in section 7.

7. Measurement of silver concentration by ICP-MS analysis

7.1) Add samples [50 µL of silver solution (for validation of the silver concentration; section 1); three dechorionated embryos (section 5); or 50 µL of filtered solution (section 5)] to a 50-mL Teflon beaker.

7.2) Add 2.0 mL of ultrapure nitric acid to the 50-mL beaker.

7.3) Heat the mixture on a hot plate at 110 °C until just before it dries out (about 3 h).

7.4) To dissolve the organic matter completely, add 2.0 mL of ultrapure nitric acid and 0.5 mL of hydrogen peroxide to the beaker.

7.5) Heat the mixture again on the hot plate until just before it dries out (about 3 h).

7.6) Dissolve the residue in 4 mL of 1.0% ultrapure nitric acid solution.

7.7) Transfer 4 mL of solution to a centrifuge tube.

7.8) Repeat 7.6 to 7.7 twice (a total of three times). The final volume is 12.0 mL.

7.9) Measure the silver concentration of the sample (dissolved in 1.0% ultrapure nitric acid) by using ICP-MS analysis according to the operating manual⁸.

7.9.1) Use an internal and an external standard solution (See Table of Materials) to quantify the silver concentration. The internal and external standard solution is accredited by American Association for Laboratory Accreditation (A2LA). Detection limits of silver were 0.0018 ng mL⁻¹

(solution) and $0.016 \text{ ng mg-weight}^{-1}$ (embryo body).

Representative Results:

The effect of salinity on SNC toxicity was very obvious: the induction of deformity or death was salinity dependent (**Figures 1** and **2**). We measured phenotypic biomarkers (heart rate, eye size, full body length, and hatching rate) in SNC (10 mg L^{-1})-exposed embryos. These phenotypic biomarkers revealed salinity-dependent SNC toxicity.

Heart rates ranged from 29.6 to 32.2 beats/15 s throughout $1\times$ to $30\times$ ERM in the controls. However, they decreased significantly ($P < 0.01$) with SNC or AgNO_3 exposure in $30\times$ ERM (**Figure 3a**). Decreasing heart rate indicates deteriorating health. There were no significant differences in full body length of the larvae under control or AgNO_3 exposure at salinities ranging from $5\times$ to $30\times$ ERM compared with the respective $1\times$ ERM solutions. Body length was consistently 4.55 to 4.69 mm. However, body length decreased significantly ($P < 0.01$) to 4.33, and 3.77, as a result of SNC exposure in $15\times$, and $20\times$ ERM compared with the respective $1\times$ ERM solutions; moreover, it decreased to 3.75 mm in $30\times$ ERM (statistical analysis was not available at $30\times$ ERM because only one hatched) (**Figure 3c**). Decreasing full body length indicates growth inhibition. There were no significant differences in eye diameter in the controls at salinities ranging from $1\times$ to $30\times$ ERM compared with $1\times$ ERM; eye diameter was consistently 0.357 to 0.366 mm. However, it decreased significantly upon SNC or AgNO_3 exposure in $20\times$ or $30\times$ ERM compared with in the respective $1\times$ ERM solutions (**Figure 3b**). Decreasing eye diameter indicates developmental inhibition of the nervous system. All control eggs hatched within 14 days. However, upon SNC exposure in $20\times$ and $30\times$ ERM the hatching rate decreased significantly to 71% and 2%, respectively, of the rate in $1\times$ ERM ($P < 0.01$) (**Figure 3d**). Also, upon AgNO_3 exposure it decreased significantly in $30\times$ ERM ($P < 0.01$). Decreasing hatching rate indicates the toxic effect of the presence of SNCs or AgNO_3 . These four phenotypic biomarkers therefore show salinity dependent SNC toxicity.

Salinity increases water-soluble metal complex formation, and these complexes might have toxic effects^{3,8}. In our study, ICP-MS analyses of silver revealed that the soluble silver concentrations in the test solutions increased as the salinity increased; the silver concentration in the embryos also increased (**Figures 3e** and **f**).

Figure 1: Increasing salinity increases SNC toxicity. Mortality and number of abnormally developed embryos increased with increasing salinity under SNC exposure.

(a) Image array of medaka eggs exposed to 10 mg L^{-1} SNC solution at different ERM concentrations. Images are typical of medaka eggs exposed to SNCs and observed under a dissecting microscope. Control medaka eggs were well developed, and all of them hatched in $1\times$ to $30\times$ ERM. At 10 mg L^{-1} SNC exposure, although all of the medaka eggs hatched in $1\times$ to $15\times$ ERM, developmental deformities (red outlined rectangles, unhatched) and embryos unhatched within 14 days (green outlined rectangles, unhatched) were observed at $20\times$ and $30\times$ ERM. **(b)** Magnified images of the lower right of Figure 1 (a).

Figure 2: Typical phenotypic biomarkers of medaka eggs exposed to SNCs. Medaka eggs at developmental stage 21 were exposed to SNCs (10 mg L^{-1}) in different concentrations of ERM for 6 days. **(a)** Control medaka embryo with normal development. **(b)** Developmental deformity (light degree of damage). This embryo displayed pericardiovascular edema; tubular heart; blood clots; inadequate development of the blood vessels (and thus ischemia), spinal cord, tail, eyes, and brain; and a short tail. **(c)** Developmental deformity (heavy degree of damage). This embryo showed destruction of the yolk sack; inadequate development of the blood vessels (and thus ischemia), spinal cord, tail, eyes, and brain; and a short tail. The signs in b) and c) were observed upon SNC exposure in 20× and 30× ERM.

Figure 3: Effects of exposure to SNCs or silver nitrate on toxicological biomarkers during medaka egg development. Developmental stage 21 medaka eggs exposed to SNCs (10 mg L^{-1}) or silver nitrate (10 mg L^{-1} as silver) in a series of ERMs were observed for 6 days. [blue] Control (ERM); [red] SNCs at 10 mg L^{-1} in ERM; [green] AgNO_3 at 10 mg L^{-1} as silver in ERM. **(a)** Heart rate per 15 s. Decreasing heart rate indicates deteriorating health. **(b)** Eye diameter. Decreasing eye diameter indicates developmental inhibition of the nervous system. **(c)** Full body length. Decreasing full body length indicates growth inhibition. **(d)** Hatching rate. Decreasing hatching rate indicates the toxic effect of the presence of SNCs. **(e)** Concentrations of soluble silver complexes from SNCs or silver nitrate in test solutions (mg L^{-1}). **(f)** Silver concentrations in embryos exposed to SNCs or silver nitrate in a series of ERMs. *Significant difference (analysis of variance, $P < 0.05$) compared with the respective 1× ERM solution. NA: not available because only one hatched. Error bars indicate standard deviation.

Discussion:

Medaka is a freshwater fish that is highly tolerant to seawater; it is not well known that the original natural habitat of this fish was saltwater off the Japanese coast⁶. Hence, medaka fish have well-developed chloride cells⁷. This unique property provides scientists with a new way to test the toxicity of chemicals in the environment as a function of salinity (freshwater to seawater) by using only a single species of fish.

To obtain medaka eggs at stage 21, eggs must be harvested every morning and selected at stage 20. Usually, medaka pairs start mating in the early morning (just before sunrise) and produce eggs by sunrise. Eggs harvested in the morning must be at about stage 10 or 11. If there is a need to control egg development before the start of the experiment, egg development can be slowed by using temperatures of 15 to 20 °C before stage 21 is reached. Measuring the silver concentration (soluble silver) in the test solutions and in dechorionated embryos was important to our investigation of the salinity dependence of SNC toxicity. Hatching enzyme is the best biologically suitable enzyme for removing the chorion, because its high specificity means that it has no harmful proteinase. Other proteinases are not recommended. So far, the only hatching enzyme available is that for medaka; this is one limitation of this method.

The obvious effect of salinity on the outcome of the chemical toxicity tests demonstrated that simulating such natural aquatic properties as realistically as possible, as in our experiments, was

useful for investigating the toxicity of chemicals in the environment. The discovery that SNC toxicity due to high silver concentrations was increased by salinity is highly applicable to the ecotoxicology of pollutant chemicals in all aquatic areas. In the case of general chemical toxicity testing in seawater, there is as yet no fish model nominated by authorized international organizations (e.g., the OECD and International Organization for Standardization). Among the freshwater fishes (e.g., medaka, zebrafish, carp, rainbow trout, and fathead minnow) that have been used for chemical toxicity testing, only the medaka has all of the advantages of salinity adaptation, hatching enzyme availability, high fecundity, and a size sufficiently small for easy use in laboratory experiments. Furthermore, medaka can be adapted to a wide temperature range (2 to 38 °C)⁶. In aquatic environments, salinity and temperature are the most important environmental influences on the fate of chemicals; our method should therefore be modifiable for a range of aquatic environmental research.

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Disclosures:

The authors declare that they have no competing financial interests.

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Figure 1a

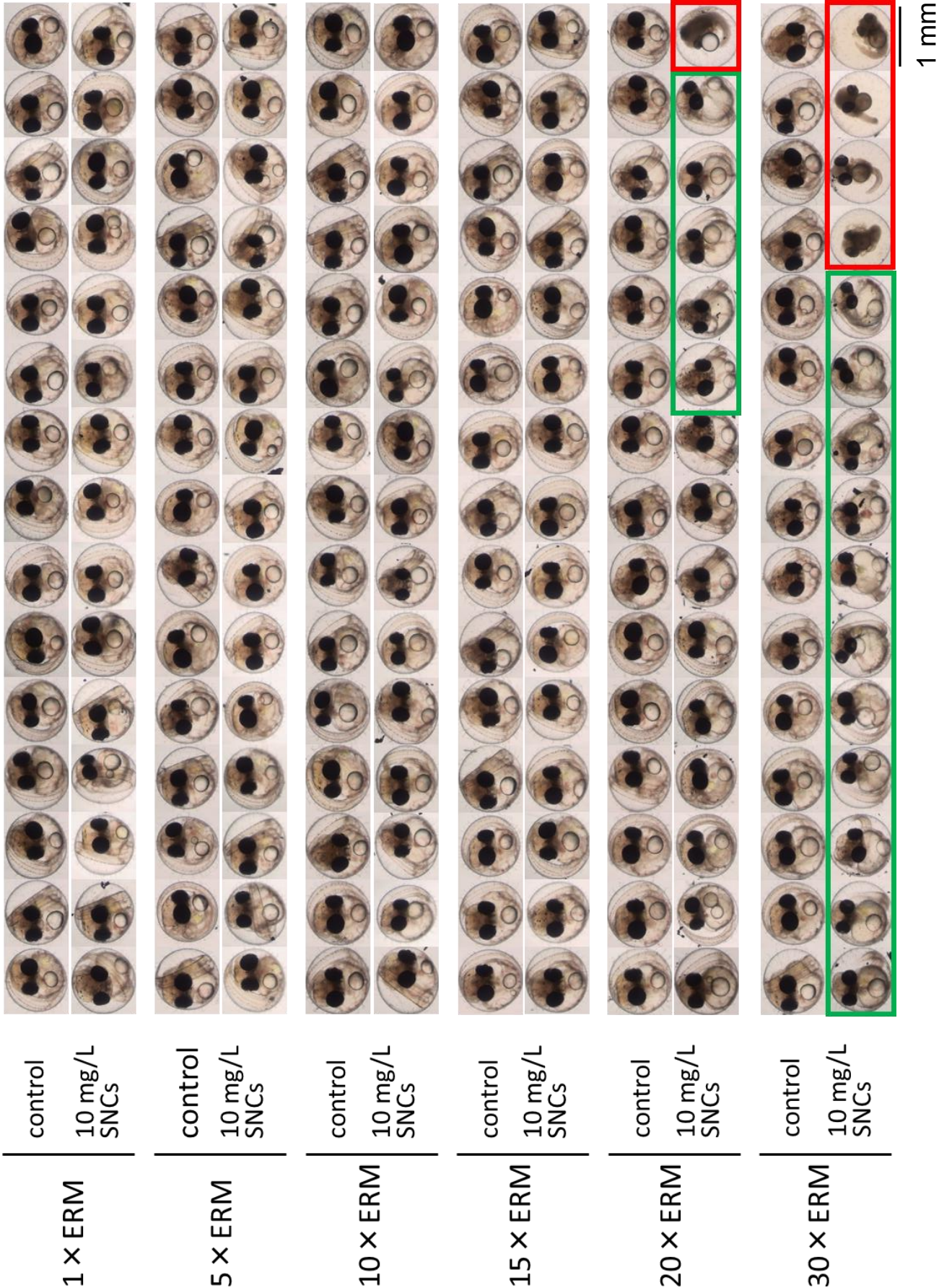


Figure 1b

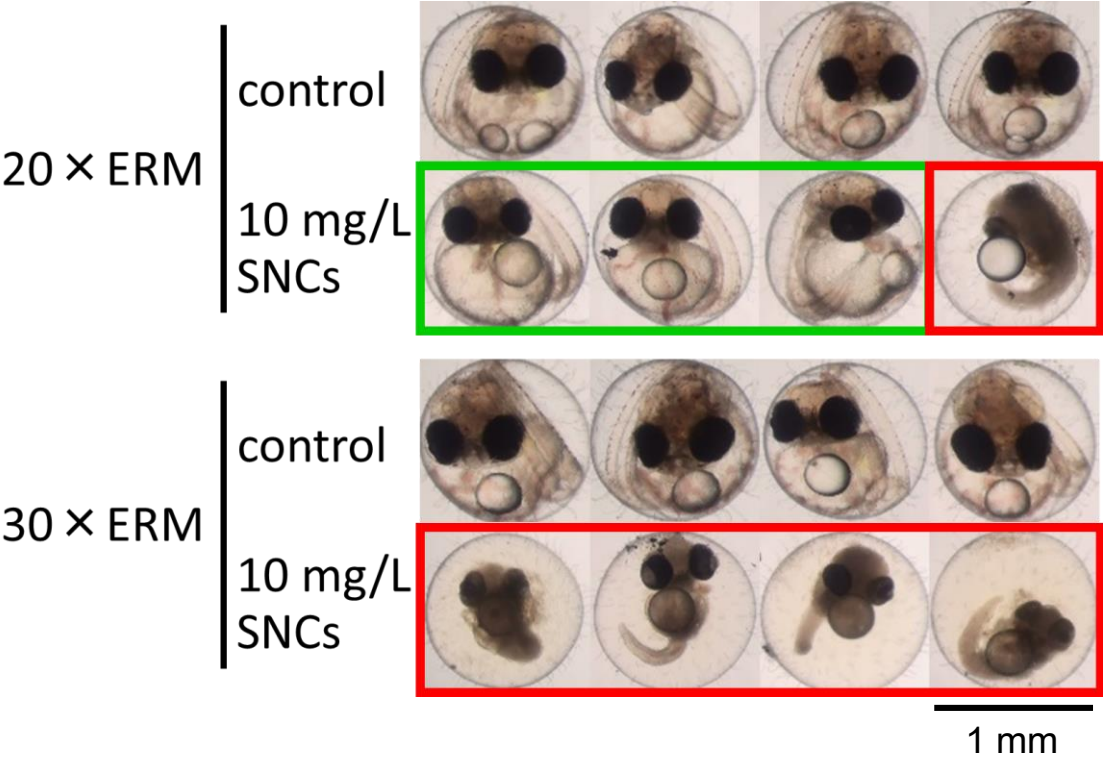


Figure 2

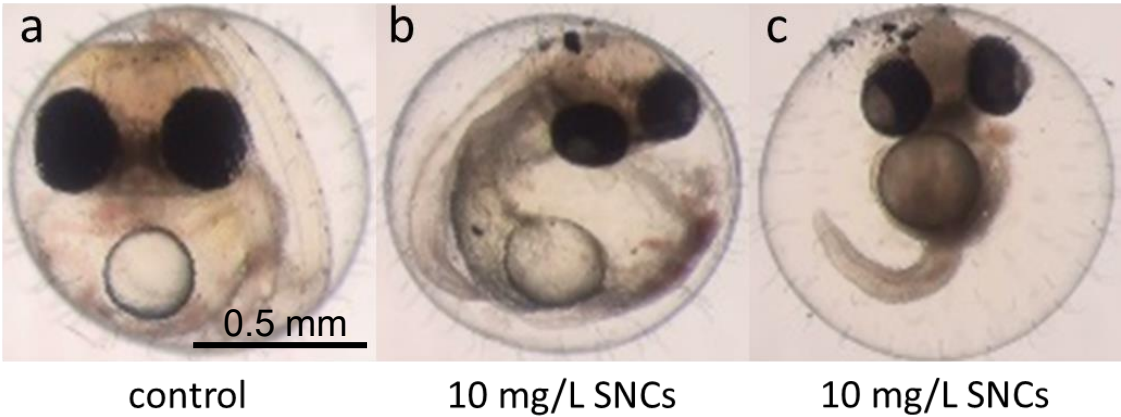
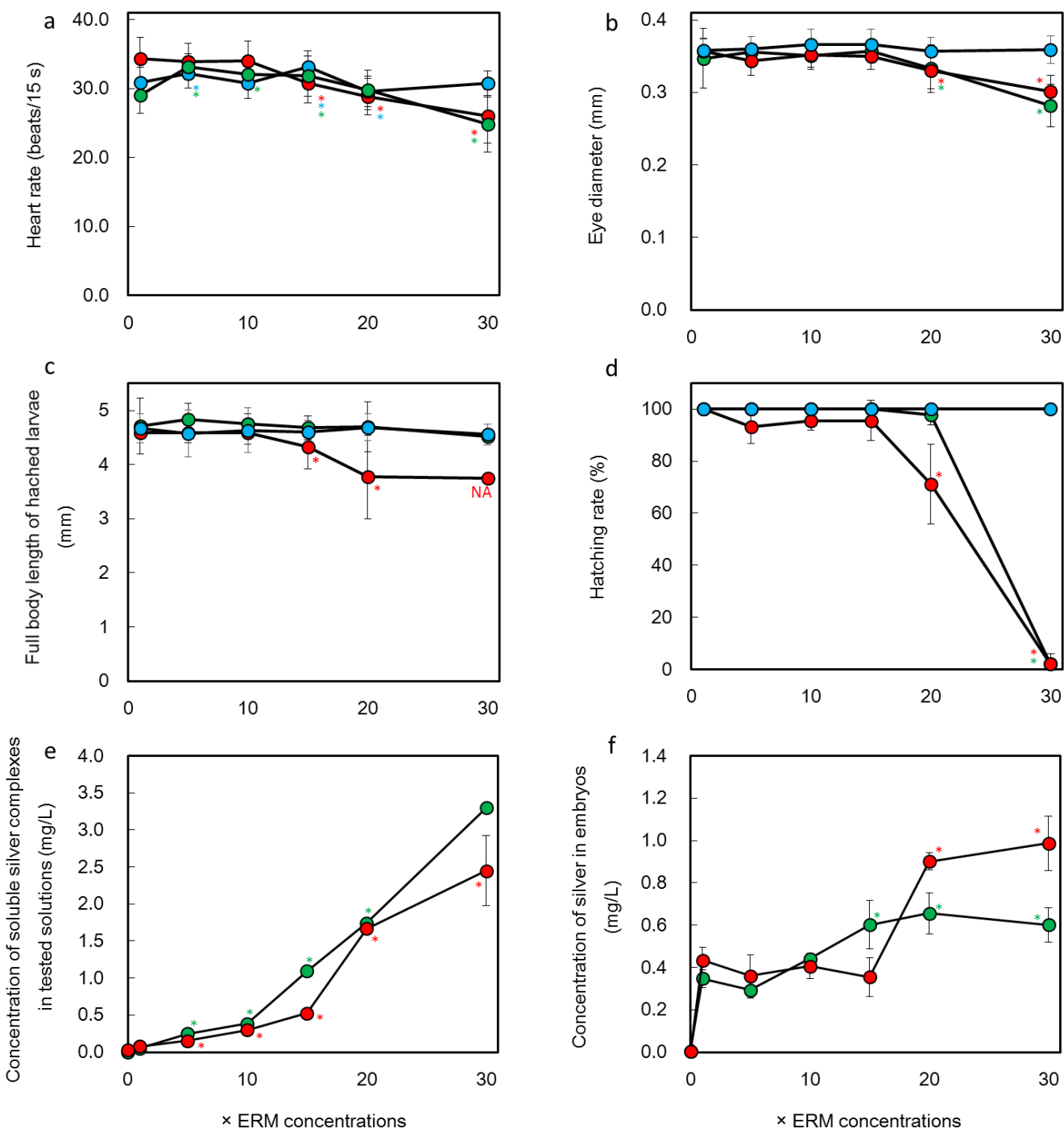


Figure 3



Name of Material/ Equipment	Company	Catalog Number	Comments/Description
Silver nanocolloids	Utopia Silver Supplements		
NaCl	Nacalai Tesque, Inc.	31319-45	For making ERM
KCl	Nacalai Tesque, Inc.	28513-85	For making ERM
CaCl ₂ ·2H ₂ O	Nacalai Tesque, Inc.	06730-15	For making ERM
MgSO ₄ ·7H ₂ O	Nacalai Tesque, Inc.	21002-85	For making ERM
NaHCO ₃	Nacalai Tesque, Inc.	31212-25	For making ERM
AgNO ₃	Nacalai Tesque, Inc.	31018-72	
pH meter	HORIBA, Ltd.	F-51S	
Balance	Mettler-Toledo International Inc.	MS204S	
medaka (Oryzias latipes) orange-red strain	National Institute for Environmental Studies		
medaka flow-through culturing system	Meito Suien Co.	MEITOsistem	
Artemia salina nauplii eggs	Japan pet design Co. Ltd	4975677033759	
aeration pump	Japan pet design Co. Ltd	non-noise w300	
Otohome larval β-1	Marubeni Nissin Feed Co. Ltd	Otohome larval β-1	Artificial dry fish diet
dissecting microscope	Leica microsystems	M165FC	
micrometer	Fujikogaku, Ltd.	10450023	
incubator	Nksystem	TG-180-5LB	
shaker	ELMI Ltd.	Aizkraukles 21-	
6-well plastic plates	Greiner CELLSTAR	M8562-100EA	
aluminum foil	AS ONE Co.	6-713-02	
stopwatch	DRETEC Co. Ltd.	SW-111YE	
3-kDa membrane filter	EMD Millipore Corporation	0.5-mL centrifugal-type filter	
50-mL Teflon beaker	AS ONE Co.	33431097	
Custom claritas standard	SPEXcertificate	ZSTC-538	For internal standard
Custom claritas standard	SPEXcertificate	ZSTC-622	For external standard
ultrapure nitric acid	Kanto Chemical Co.	28163-5B	
hydrogen peroxide	Kanto Chemical Co.	18084-1B	for atomic absorption spectrometry
ICP-MS	Thermo Scientific	Thermo Scientific X Series 2	

hot plate

Tiger Co.

CRC-A300



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Author(s): **Chisato Kataoka, Shosaku Kashiwada**

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Article Title: **Salinity-dependent toxicity assay of silver nanocolloids using medaka eggs**
Signature:  Date: **April - 8 - 2015**

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Editorial comments:

The manuscript has been modified by the Science Editor to comply with the JoVE formatting standard. Please maintain the current formatting throughout the manuscript. The updated manuscript (53550_R3_061815.docx) is located in your Editorial Manager account. In the revised PDF submission, there is a hyperlink for downloading the .docx file. Please download the .docx file and use this updated version for any future revisions.

Changes made by the Science Editor:

1. There have been edits made to the manuscript.

Changes to be made by the Author(s):

1. Please upload each Figure individually to your Editorial Manager account as a .png, .pdf, or a .tiff file. Please combine all panels of one figure into a single file.

2. Formatting/Grammar:

-lines 45-47 – Please correct the grammar in the sentence starting “After hatching, larvae were...” as well as in the sentence after it.

-Please use consistent units (ie 10 mg/L or 10 mg L⁻¹)

I corrected

-Please correct the grammar in the note under step 2.5.1.

I corrected

-5.2/6.3 – “is described in” instead of “was described at.”

OK

3. Additional detail is required:

-7.9.1 – What standard solutions were used?

I added information at 7.9.1.

-Figures 1 & 2 – Please provide a scale bar for at least one image.

I added scale bar

-Figure 3 – Please define the error.

I added information.

Reviewers' comments:

Editor's Note: Please note that the reviewers raised some significant concerns regarding your method and your manuscript. Please thoroughly address each concern by revising the manuscript or addressing the comment in your rebuttal letter.

Reviewer #1:*Manuscript Summary:*

Kataoka and colleagues improved procedures to test the salinity dependence of toxicity for aquatic organisms using fresh water medaka. The authors defined the relationship of the toxicity of the SNCs increasing the deformities of the medaka embryos, obviously increased in conjunction with increasing the concentration of the salinity in the rearing water surrounding the embryos using their new procedures.

Major Concerns:

The content of this manuscript is very interesting, however this reviewer highly recommends that the authors should have somebody who is a native English speaker check the English in their text and who can professionally assist the authors. Some descriptions in the text are a little vague, and the accuracy of their results are compromised by their poor description.

For example, the use of the term, "egg development", is a little vague. Are you talking about the "embryonic development within the egg"? The phrase "egg development" gives this reviewer the impression that your intention is to say, "oocyte development during the oogenesis", however, my comment does not pertain to the type of description the authors used concerning the phenomena after spawning. After spawning, embryos develop within the egg; however the eggs never develop. Please check the content.

Please check the content.

p. 1 lines 37, 38, & 39

In this research, different strengths of embryo rearing medium (ERM) (1x, 5x, 10x, 15x, 20x, and 30x) were employed to test the toxicity of silver nanocolloidal particles (SNCs) to medaka eggs.

What does it mean, "different strengths of embryo rearing medium"? What is "different strength" ?

I corrected strength to "concentration"

p. 2 lines 46 & 47

The embryos were observed until hatching or 14 days, and then counted hatching rate for 14 days

Does it mean "... and then we counted the hatching rate...? There is no subject in the second clause

"counted hatching rate for 14 days"

Correct the description to "... we counted the hatching rate every day for 14 days".

p. 2 lines 50 & 51

"The toxicity of the SNCs to medaka eggs obviously increased with increasing salinity."

Correct the description

The toxicity of the SNCs for medaka embryos obviously increased with the increasing salinity.

p. 2 lines 60 & 61

"Freshwater environments are more directly affected than saltwater ones by human economic and industrial activities."

Change the description:

"Freshwater environments are more directly affected by human economic and industrial activities than saltwater ones."

p. 2 lines 61 & 62

"Therefore freshwater environments have been prioritized for testing because they are at high risk from pollution."

Change the description:

Therefore freshwater environments have been prioritized for testing because they are at higher risk from pollution.

p. 7 lines 267, 268 & 269

"The effect of salinity on SNC toxicity is very obvious in Figures 1 and 2. Also, we measured phenotypic biomarkers (heart rate, eye size, full body length, and hatch rate) of SNC (10 mg L⁻¹) exposed embryos."

How obvious?

I added information.

Remove "Also"

"The effect of salinity on SNC toxicity is very obvious in Figures 1 and 2. We measured phenotypic biomarkers (heart rate, eye size, full body length, and hatch rate) of SNC (10 mg L⁻¹) exposed embryos."

line 270

Correct "In case of heart rate" to "in the case of"

Correct "In case of heart rate in the controls, it was ranged from 29.6....."to "In the case of the heart rates, they ranged from 29.6..... x ERM in the controls."

line 273

correct "post-hatch larvae" to "larvae" because larvae are not embryos; they have already hatched out.

line 273

correct "there were no significant difference under control and AgNO₃ exposure" to "there were no significant difference between control and AgNO₃ exposure"

What is control here?

I compared with the respective 1× ERM solution.

line 275

Correct "However, the body length was decreased significantly" to "However, the body length decreased significantly"

lines 275 & 276

Correct "the body length was decreased significantly ($P < 0.01$) to 4.33, 3.77, and 3.75 mm by

SNCs exposure" to "the body length decreased significantly ($P < 0.01$) to 4.33, 3.77, and 3.75 mm due to SNCs exposure"

lines 277 & 278

"there were no significant difference under control at salinities ranging from 1× to 30× ERM"

Rewrite this sentence.

line 284

correct "Decreasing hatch rate indicates the toxic ..." to "Decreasing hatching rate indicates the toxic ..."

This reviewer realized that the authors need to make a lot of correction in the section of "Discussion," So, this reviewer stopped making corrections here. However, this reviewer

highly recommends the authors find a native English speaker, to thoroughly read and correct this part of the article.

The manuscript has been edited carefully by two native-English-speaking professional editors from ELSS, Inc. (elss@elss.co.jp, <http://www.elss.co.jp>).

Minor Concerns:

N/A

Additional Comments to Authors:

N/A

Reviewer #2:

Manuscript Summary:

N/A

Major Concerns:

It is interesting article using salinity to investigate toxicity. However, this fish species have a very special property on salinity(they could live on fresh water & sea water).

If the author use other species fish to investigate relationship with salinity and toxicity, I think it maybe difference or couldn't investigate.

Silver nanoparticle could affect on difference ion concentration of medium(water), because silver nanoparticle is a type of ion(Ag⁺). Difference concentration of ion(Na⁺, Cl⁻) could be changed the particle size(make AgCl).

I couldn't find the result of SEM or TEM to observe the particle size change(before and after). It is very important process in this experiment.

I think image of SEM or TEM need not for this MN, because purpose of this MN is to describe new method of test for the toxicity of chemicals in different salinities by using only a single species of fish.

Minor Concerns:

N/A

Additional Comments to Authors:

N/A