

Journal of Visualized Experiments

Generation of Induced Pluripotent Stem Cells from Frozen Buffy Coats using non-integrating Episomal Plasmids --Manuscript Draft--

Manuscript Number:	JoVE52885R2
Full Title:	Generation of Induced Pluripotent Stem Cells from Frozen Buffy Coats using non-integrating Episomal Plasmids
Article Type:	Invited Methods Article - JoVE Produced Video
Keywords:	stem cell biology; cellular biology; molecular biology; induced pluripotent stem cells; peripheral blood mononuclear cells; reprogramming; episomal plasmids.
Manuscript Classifications:	1.11: Cells; 7.16: Biological Phenomena; 95.51.10: biology (general)
Corresponding Author:	Rossini Alessandra European Academy of Bozen-EURAC research Bolzano, Bolzano ITALY
Corresponding Author Secondary Information:	
Corresponding Author E-Mail:	Alessandra.Rossini@eurac.edu
Corresponding Author's Institution:	European Academy of Bozen-EURAC research
Corresponding Author's Secondary Institution:	
First Author:	Viviana Meraviglia
First Author Secondary Information:	
Other Authors:	Viviana Meraviglia
	Alessandra Zanon
	Lavdas A. Alexandros
	Christine Schwienbacher
	Rosamaria Silipigni
	Marina Di Segni
	Huei-Sheng Vincent Chen
	Peter P. Pramstaller
	Andrew A. Hicks
Order of Authors Secondary Information:	
Abstract:	Somatic cells can be reprogrammed into induced pluripotent stem cells (iPSCs) by forcing the expression of four transcription factors (Oct-4, Sox-2, Klf-4, and c-Myc), typically expressed by human embryonic stem cells (hESCs). Due to their similarity with hESCs, iPSCs have become an important tool for potential patient-specific regenerative medicine, avoiding ethical issues associated with hESCs. In order to obtain cells suitable for clinical application, transgene-free iPSCs need to be generated to avoid transgene reactivation, altered gene expression and misguided differentiation. Moreover, a highly efficient and inexpensive reprogramming method is necessary to derive sufficient iPSCs for therapeutic purposes. Given this need, an efficient non-integrating episomal plasmid approach is the preferable choice for iPSC derivation. Currently, the most common cell type used for reprogramming purposes are fibroblasts, the isolation of which requires tissue biopsy, an invasive surgical procedure

	<p>for the patient. Therefore, human peripheral blood represents the most accessible and least invasive tissue for iPSC generation.</p> <p>In this study, a cost-effective and viral-free protocol using non-integrating episomal plasmids is reported for the generation of iPSCs from human peripheral blood mononuclear cells (PBMNCs) obtained from frozen buffy coats after whole blood centrifugation and without Ficoll separation.</p>
Author Comments:	
Additional Information:	
Question	Response
If this article needs to be "in-press" by a certain date to satisfy grant requirements, please indicate the date below and explain in your cover letter.	
If this article needs to be filmed by a certain date to due to author/equipment/lab availability, please indicate the date below and explain in your cover letter.	



Alessandra Rossini, PhD

*EURAC, Center for Biomedicine
Via Galvani 31, 39100 Bolzano, Italy
Tel. +390471055504
Fax. +390471055599*

e-mail: alessandra.rossini@eurac.edu

Bolzano, December 18th, 2014

Dear Dr. Singh and Dr. Nguyen,

We revised the manuscript entitled **“Generation of induced pluripotent stem cells from frozen buffy coats using non-integrating episomal plasmids”** according to the Editorial and Reviewers’ comments.

The authors agreed with all the Editorial and Reviewers’ comments and uploaded the revised file with modifications in “track changes” mode, together with the authors’ reply to each Reviewer’s observation and suggestion.

We hope that our revised manuscript could be accepted for publication in *Journal of Visualized Experiments (JoVE)* as a Methods Article-JoVE Produced Video, under JoVE Biology section.

We thank you for your consideration to our manuscript.

Sincerely,

Alessandra Rossini, Ph.D.

TITLE:

Generation of Induced Pluripotent Stem Cells from Frozen Buffy Coats using non-integrating Episomal Plasmids

AUTHORS:

Meraviglia, Viviana *
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
viviana.meraviglia@eurac.edu

Zanon, Alessandra *
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
alessandra.zanon@eurac.edu

Lavdas, Alexandros A.
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
alexandros.lavdas@eurac.edu

Schwienbacher, Christine
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
christine.schwienbacher@eurac.edu

Silipigni, Rosamaria
Laboratory of Medical Genetics
Fondazione IRCCS Ca' Granda, Ospedale Maggiore Policlinico,
Milano, Italy
rosamariasilipigni@gmail.com

Di Segni, Marina
Laboratory of Medical Genetics
Fondazione IRCCS Ca' Granda, Ospedale Maggiore Policlinico,
Milano, Italy
marina.disegni@policlinico.mi.it

Chen, Huei-Sheng Vincent
Del E. Webb Center for Neuroscience, Aging & Stem Cell Research
Sanford-Burnham Medical Research Institute

La Jolla, California
hsv_chen@sanfordburnham.org

Pramstaller, Peter P.
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
peter.pramstaller@eurac.edu

Hicks, Andrew A.
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
andrew.hicks@eurac.edu

Rossini, Alessandra
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
alessandra.rossini@eurac.edu

* These authors equally contributed to this work.

CORRESPONDING AUTHOR:

Rossini, Alessandra
Center for Biomedicine
European Academy Bozen/Bolzano (EURAC) (affiliated institute of the University of Lübeck)
Bolzano, Italy
alessandra.rossini@eurac.edu

KEYWORDS:

Stem cell biology, cellular biology, molecular biology, induced pluripotent stem cells, peripheral blood mononuclear cells, reprogramming, episomal plasmids.

SHORT ABSTRACT:

Induced pluripotent stem cells (iPSCs) represent a source of patient-specific tissues for clinical applications and basic research. Here, we present a detailed protocol to reprogram human peripheral blood mononuclear cells (PBMNCs) obtained from frozen buffy coats into viral-free iPSCs using non-integrating episomal plasmids.

LONG ABSTRACT:

Somatic cells can be reprogrammed into induced pluripotent stem cells (iPSCs) by forcing the expression of four transcription factors (Oct-4, Sox-2, Klf-4, and c-Myc), typically expressed by human embryonic stem cells (hESCs). Due to their similarity with hESCs, iPSCs have become an

important tool for potential patient-specific regenerative medicine, avoiding ethical issues associated with hESCs. In order to obtain cells suitable for clinical application, transgene-free iPSCs need to be generated to avoid transgene reactivation, altered gene expression and misguided differentiation. Moreover, a highly efficient and inexpensive reprogramming method is necessary to derive sufficient iPSCs for therapeutic purposes. Given this need, an efficient non-integrating episomal plasmid approach is the preferable choice for iPSC derivation. Currently the most common cell type used for reprogramming purposes are fibroblasts, the isolation of which requires tissue biopsy, an invasive surgical procedure for the patient. Therefore, human peripheral blood represents the most accessible and least invasive tissue for iPSC generation.

In this study, a cost-effective and viral-free protocol using non-integrating episomal plasmids is reported for the generation of iPSCs from human peripheral blood mononuclear cells (PBMNCs) obtained from frozen buffy coats after whole blood centrifugation and without density gradient separation.

INTRODUCTION:

In 2006, the group of Shinya Yamanaka¹ demonstrated for the first time that somatic cells from adult mice and humans can be converted into a pluripotent state by ectopic expression of four reprogramming factors (Oct-4, Sox-2, Klf-4, and c-Myc), generating the so-called Induced Pluripotent Stem Cells (iPSCs)². Patient-specific iPSCs closely resemble human embryonic stem cells (hESCs) in terms of morphology, proliferation and ability to differentiate into the three-germ cell types (mesoderm, endoderm and ectoderm) while lacking the ethical concerns associated to the use of hESCs and bypassing possible immune rejection³. Thus, iPSCs appear as one of the most important sources of patient-specific cells for basic research, drug screening, disease modeling, evaluation of toxicity, and regenerative medicine purposes⁴.

Several approaches have been used for iPSC generation: viral integrating vectors (retrovirus⁵, lentivirus⁶), viral non-integrating vectors (adenovirus⁷), Sendai-virus⁸, BAC transposons⁹, episomal vectors¹⁰, proteins¹¹ or RNA delivery¹². Although the use of virus-mediated methods can lead to high efficiency reprogramming, viral vectors integrate into the genome of host cells and therefore potential random insertional mutagenesis, permanent alteration of gene expression, and reactivation of silenced transgenes during differentiation cannot be excluded¹³. To make iPSCs safer for regenerative medicine, efforts have been made to derive iPSCs without the integration of exogenous DNA into cellular genomes. Although excisable viral vectors and transposons have been developed, it is still unclear whether short vector sequences, which inevitably remain in the transduced cells after excision, and transposase expression, could induce alteration in cellular function¹³. Despite its high reprogramming efficiency, Sendai virus represents an expensive approach and reach-through licensing concerns with the company that developed this system have the potential to limit its application in translational studies. Furthermore, the need for direct introduction of proteins and RNA requires multiple delivery of reprogramming molecules with the inherent technical limitations this introduces, and overall reprogramming efficiency is very low¹⁴. Of note, cost-effective viral-free and non-integrating methods based on the use of episomal plasmids have been successfully reported for the

reprogramming of skin fibroblasts¹⁵. Specifically, in the present work we decided to use commercial available integration-free episomal plasmids, as previously reported^{10, 15}.

To date, skin fibroblasts represent the most popular donor cell type⁵. However, other cell sources have been successfully reprogrammed into iPSCs including keratinocytes¹⁶, bone marrow mesenchymal stem cells¹⁷, adipose stromal cells¹⁸, hair follicles¹⁹, and dental pulp cells²⁰. The isolation of these cells requires surgical procedures, and several weeks are needed for *in vitro* cell expansion in order to establish a primary cell culture.

In this light, the selection of starting cell type is critical and it is equally important to be able to produce iPSCs from easily accessible and less invasive tissues such as blood. Both cord blood mononuclear cells (CBMNCs)^{21, 22} and peripheral blood mononuclear cells (PBMNCs)^{14, 22-24} represent suitable sources of cells for the derivation of iPSCs.

Although the efficiency of adult PBMNC reprogramming is 20–50 times lower than that of CBMNCs²², they remain the most convenient cell type for sampling purpose. In fact, PBMNC sampling has the advantage of being minimally invasive, and in addition, these cells do not require extensive expansion *in vitro* before reprogramming experiments. To date, different protocols have reported that PBMNCs after density gradient separation can be frozen and thawed days to several months after freezing and expanded for few days before reprogramming into iPSCs^{22, 23}. Nevertheless, as far as we are aware no reports have described reprogramming of PBMNCs from frozen buffy coats. Importantly, frozen buffy coats collected without density gradient separation represent the most common blood samples stored in large scale biobanks from population studies, thus representing an easily accessible pool of material for iPSC production that avoids further sample collection.

Herein we report for the first time the generation of viral-free iPSCs from human frozen buffy coats, based on a previously described protocol²². In addition, iPSCs were generated from frozen PBMNCs obtained after density gradient separation, as a control protocol for the non-density gradient purified PBMNC results.

PROTOCOL:

Peripheral blood mononuclear cells (PBMNCs) were isolated from human peripheral blood samples of healthy donors after signed informed consent and approval of the Ethical Committee of the Province of South Tyrol. Experiments were conducted in accordance with the principles expressed in the Declaration of Helsinki. All data were collected and analyzed anonymously.

1. Isolation of Peripheral Blood Mononuclear Cells (PBMNCs)

1.1. PBMNCs obtained from buffy coats after whole blood centrifugation, without density gradient separation.

1.1.1. Collect 8 mL of venous peripheral blood into a sodium citrate buffered plastic tube, and store at room temperature (25 °C). Process blood within 12 h after collection.

1.1.2. Centrifuge the tube at 2000 x g for 15 min at 4 °C in a swinging bucket rotor, switching-off the centrifuge brakes.

1.1.3. Collect the cloudy buffy coat (the layer between the upper plasma phase and the lower phase containing most of the erythrocytes), containing the enriched PBMNC fraction (500 µL) into a cryovial tube.

1.1.4. Resuspend 500 µL of buffy coat with 500 µL of 2X freezing medium containing 90% FBS and 20% DMSO, in order to obtain a total volume of 1 mL with 10% DMSO concentration.

1.1.5. Freeze the vial in a controlled-rate freezing container at -80 °C. Store cells in liquid nitrogen for longer periods.

1.2. PBMNCs obtained after density gradient separation

1.2.1. Collect 12 mL of venous peripheral blood into a sodium citrate buffered plastic tube, and store at room temperature. Process blood within 12 h after collection.

1.2.2. Dilute the blood with sterile PBS to a final volume of 35 mL.

1.2.3. Carefully and slowly, layer 35 mL of diluted blood on 15 mL of polysucrose solution (used for density gradient separation) (P=1.077) in a 50 mL conical tube (ratio 3:1).

1.2.4. Centrifuge the tube at 800 x g for 30 min at room temperature in a swinging bucket rotor, switching-off the centrifuge brakes.

1.2.5. Aspirate the upper, yellow phase containing plasma and discard it. Carefully, transfer the opaque white interphase layer containing PBMNCs to a new 50 mL conical tube.

1.2.6. Bring total volume to 30 mL with sterile PBS and centrifuge at 300 x g for 10 min at room temperature.

1.2.7. Discard the supernatant and resuspend the cells with 30 mL of PBS. Count the number of viable cells, based on trypan blue exclusion²⁵.

1.2.8. Centrifuge PBMNCs at 300 x g for 10 min at room temperature, aspirate the supernatant and discard it.

1.2.9. Resuspend the cell pellet in an appropriate amount of freezing medium containing 90% FBS and 10% DMSO, in order to obtain a concentration of 5×10^6 cells/mL.

1.2.10. Aliquot 1 mL per vial (about 5×10^6 cells/mL) and freeze the vial in a controlled-rate freezing container at -80°C . Store cells in liquid nitrogen for longer periods.

2. Thawing and plating of PBMNCs (DAY 0)

2.1. PBMNCs obtained from buffy coats after whole blood centrifugation, without density gradient separation.

2.1.1. To start the protocol using PBMNCs from frozen buffy coats, thaw 1 vial of cells in a water bath at 37°C and dilute the buffy coat with sterile PBS to a final volume of 2 mL.

2.1.2. Transfer the diluted buffy coat into a 50 mL conical tube, add 10 mL of red cell lysis buffer and incubate for 10 min at room temperature.

2.1.3. To prepare 500 mL of red cell lysis buffer, add 0.5 g of potassium bicarbonate, 4.145 g of ammonium chloride, 100 μL of 0.5 M EDTA solution to 500 mL of ultrapure water, pH 7.2-7.4.

2.1.4. Bring total volume to 50 mL with sterile PBS and centrifuge at $300 \times g$ for 10 min at room temperature.

2.1.5. Discard the supernatant and wash the pellet with 50 mL of sterile PBS, centrifuge the buffy coat at $300 \times g$ for 10 min at room temperature.

2.1.6. Resuspend the cells in 1 mL of PBMNC medium, as previously described²², composed of: IMDM and Ham's F-12 (ratio 1:1), 1% of Insulin-Transferrin-Selenium-Ethanolamine (ITS-X), 1% of Chemically Defined Lipid Concentrate (CDL), 1% Penicillin/Streptomycin, 0.005% of L-ascorbic acid, 0.5% of Bovine Serum Albumin (BSA), 1-Thioglycerol (final concentration 200 μM), Stem Cell Factor SCF (100 ng/mL), Interleukin-3 IL-3 (10 ng/mL), Erythropoietin EPO (2 U/mL), Insulin-like Growth Factor IGF-1 (40 ng/mL), Dexamethasone (1 μM) and holo-transferrin (100 $\mu\text{g/mL}$).

2.1.7. Transfer them into one well of a 12-well plate with standard tissue culture treated surface, without coating, at a density of 2×10^6 cells/mL. Incubate the cells at 37°C , 5% CO_2 in a humidified incubator for 48 h, until the changing medium step (see Section 3).

2.2. PBMNCs obtained after density gradient separation

2.2.1. To start the protocol using frozen PBMNCs after density gradient separation, thaw 1 vial of cells in a water bath at 37°C , transfer the cells into 5 mL of PBMNC medium. Centrifuge at $300 \times g$ for 10 min at room temperature.

2.2.2. Resuspend the cells in 1 mL of PBMNC medium and transfer them into one well of a 12-well plate, at a density of 2×10^6 cells/mL. Incubate the cells at 37°C , 5% CO_2 in a humidified incubator for 48 h, until the changing medium step (see Section 3).

3. Culture and expansion of PBMNCs (DAY 2-13)

3.1. Collect PBMNCs in suspension culture, using a pipette with a 1 mL tip, in a 15 mL conical tube and centrifuge at 300 x g for 10 min at room temperature.

3.2. Discard the supernatant and resuspend the cells in 1 mL of fresh PBMNC medium.

3.3. Transfer the cells into 1 well of 12-well plate with standard tissue culture treated surface, without coating, and incubate them at 37 °C, 5% CO₂ in a humidified incubator.

3.4. Repeat these steps every two days, and split the cells at a ratio 1:2 when they reach the appropriate confluence (around 80%).

4. Transfection of PBMNCs with episomal plasmids (DAY 14)

Note: Perform the isolation of the four commercial intergation-free episomal plasmids, carrying the pluripotency genes using a commercial kit for plasmid purification and assess the quality of the purified plasmids using an agarose gel analysis, according to the manufacturer's instructions.

4.1. Collect PBMNCs in 15 mL conical tube and count the number of viable cells (on the basis of trypan blue exclusion)²⁵.

4.2. Centrifuge 2×10^6 cells at 300 x g for 10 min at room temperature.

4.3. Resuspend 2×10^6 cells in transfection mix containing 100 µL of Resuspension Buffer T and 1 µg of each plasmid DNA (one plasmid carrying OCT3/4 and shRNA against p53, one plasmid carrying SOX2 and KLF4, one plasmid carrying L-MYC and LIN28, and one plasmid carrying EGFP, to check the transfection efficiency).

4.4. Aspirate the transfection mix containing the cells into a 100 µL tip and insert the pipette with the sample vertically into the tube, filled with 3 mL of Electrolytic Buffer E2, placed in the pipette station of electroporation machine, according to manufacturer's instructions.

4.5. Efficiently electroporate cells using the following program: 1650 V, 10 ms, 3 pulses.

4.6. Resuspend the electroporated cells (2×10^6 cells) into 2 mL of pre-warmed PBMNC medium, adding 0.25 mM Sodium Butyrate (NaB) and count the number of viable cells after electroporation protocol (on the basis of trypan blue exclusion)²⁵.

4.7. Transfer the cells into one well of a 6-well plate without coating, gently rock the plate and incubate the cells at 37 °C, 5% CO₂ in a humidified incubator.

4.8. The day after electroporation, count the number of GFP-positive cells under

fluorescence microscopy from 8-10 random fields, in order to assess the transfection efficiency.

4.9. Maintain the transfected cells without splitting for 3 days, until plating them on MEFs (see Section 6).

4.10. Replace 2 mL of fresh PBMNCs medium, plus 0.25 mM NaB every two days.

5. Prepare dishes with feeder-cells for the co-culture (DAY 16)

5.1. Coat the wells of a 6-well plate with 1 mL of basement membrane matrix for 15 min at 37 °C and plate Mouse Embryonic Fibroblasts (MEFs) at a density of 2×10^5 cells/well with 2 mL of 10% FBS contained DMEM (MEF medium) one day before passaging the electroporated PBMNCs onto MEF feeder-cells.

6. Plating transfected PBMNCs onto MEFs (DAY 17-19)

6.1. Collect PBMNCs in suspension culture, using a pipette with 1 mL tip, in 15 mL conical tube and centrifuge transfected PBMNCs at 300 x g for 10 min at room temperature.

6.2. Discard the supernatant and resuspend the pellet in 2 mL of fresh PBMNC medium, plus 0.25 mM NaB.

6.3. Plate the cells onto MEF-coated 6 well-plate and incubate the cells at 37 °C, 5% CO₂ in a humidified incubator.

6.4. After two days, aspirate the medium and replace it with 2 mL of iPSC medium composed of knockout DMEM (KO-DMEM), 20% KO-Serum Replacement (KOSR), 1 mM NEAAs, 1% Penicillin/Streptomycin, 20 mM L-Glutamine, 0.1 mM β-mercaptoethanol, 10 ng/mL FGF (basic bFGF) and 0.25 mM NaB.

6.5. Replace 2 mL of fresh iPSC medium every day and check the cells daily.

Note: At about day 22-25, the first colonies of iPSCs should be visible.

7. Picking and expansion of Induced Pluripotent Stem Cells (iPSCs) clones (DAY 30-35)

Note: At about 30-35 days after transfection, iPSC colonies should be ready for the picking and replating. The reprogramming efficiency should be estimated as the percentage of number of iPSC colonies/total number of electroporated cells, as previously reported²⁶.

7.1. The day before passaging iPSC colonies, prepare the MEF feeder in a 12-well plate (1.25×10^5 cells/well).

7.2. Aspirate the MEF medium and change with 1 mL of iPSC medium with 10 ng/mL bFGF

and 10 μ M Y-27632. Manually dissect with a needle one colony at each time under the dissecting microscope in the picking-hood in order to obtain a grid, collect smaller clumps by pipetting and transfer them individually each into one well of 12-well plate coated with MEFs.

7.3. Passage and expand each 12-well plate to a 6-well plate in a ratio 1:2, then each 6-well plate in a ratio 1:4 for further propagation. Dissect and expand iPSCs manually for the first 2-3 passages (as thoroughly described in step 7.2), then use an enzyme dissociation procedure (start from step 7.4).

7.4. For an enzyme dissociation protocol, clean iPSC colonies by aspirating differentiated portions, under the dissecting microscope in the picking-hood.

7.5. Incubate iPSCs with 1 mL of collagenase IV (1 mg/mL) for 10 minutes at 37 °C.

7.6. Aspirate the enzyme solution, rinse the cells with 1 mL of KO-DMEM and collect the colonies in a 15-ml conical tube. Centrifuge at 100 x g for 2 min at room temperature.

7.7. Aspirate the supernatant, resuspend the colonies in 2 mL of iPSC medium with 10 ng/mL bFGF and 10 μ M Y-27632 and plate them on MEF-coated 6-well plate (ratio 1:4).

8. Characterization of iPSCs (between 5-15 passages)

8.1. Alkaline Phosphatase Staining

8.1.1. Culture the iPSC colonies for 3-5 days in iPSC medium with 10 ng/mL bFGF.

8.1.2. For starting an alkaline phosphatase staining protocol, rinse iPSCs once with PBS and then fix them with 1 mL of 4% PFA for 20 min at room temperature.

8.1.3. Wash the cells three times with PBS, to remove residual PFA.

8.1.4. Stain fixed cells using a commercial alkaline phosphatase staining kit according to the manufacturer's instructions.

8.2. Immunofluorescence

8.2.1. Culture the iPSC colonies for 3-5 days in iPSC medium with 10 ng/mL bFGF.

8.2.2. To start an immunofluorescence assay, wash iPSCs once with PBS and then fix them with 1 mL of 4% PFA for 20 min at room temperature.

8.2.3. Wash the cells three times with PBS, to remove residual PFA.

8.2.4. Treat fixed iPSCs with 1 mL of permeabilization/blocking solution containing 4% goat

serum, 0.1% BSA, 0.1% Triton X-100 and 0.05% sodium azide (NaN_3) for 1 h at room temperature.

8.2.5. Aspirate the blocking solution and incubate the fixed cells with primary antibodies in 3% BSA and 0.05% NaN_3 solution overnight at 4 °C (check the material table for primary antibody list and suggested antibody dilution factor).

8.2.6. The next day, wash iPSCs three times with PBS, then incubate the cells with Alexa Fluor 488 goat anti-mouse and Alexa Fluor 555 goat anti-rabbit antibodies, diluted 1:1000 in 3% BSA and 0.05% NaN_3 solution, for 1 h at 37 °C.

8.2.7. Stain the nuclei with DAPI (1 $\mu\text{g}/\text{mL}$) for 10 min at room temperature in dark, followed by three time washing with PBS.

8.3. Differentiation of iPSCs in embryoid bodies (EBs)

8.3.1. Culture the iPSC colonies for 5-6 days in iPSC medium with 10 ng/mL bFGF.

8.3.2. Remove differentiated portions from iPSC colonies by aspirating, under the dissecting microscope in the picking-hood.

8.3.3. Incubate iPSCs with 1 mL of collagenase IV (1 mg/mL) for 10 minutes at 37 °C.

8.3.4. Aspirate the enzyme solution, rinse the cells with 1 mL of KO-DMEM and collect the colonies in a 15 mL conical tube. Centrifuge at 100 x g for 2 min at room temperature.

8.3.5. Aspirate the supernatant and resuspend the colonies in 2 mL of EB medium composed of KO-DMEM, 20% defined fetal bovine serum (FBS), 1 mM NEAAs, 1% Penicillin/Streptomycin, 20 mM L-Glutamine, 0.1 mM β -mercaptoethanol.

8.3.6. Plate iPSC colonies in suspension for 6 days onto ultra-low attachment 6-well plates, in order to allow the formation of spheroidal three-dimensional embryoid bodies (EBs). Change 2 mL of fresh EB medium every two days.

8.3.7. On day 7, collect EBs and plate them onto 0.1% gelatin coated 6-well plates in 2 mL of EB medium and maintain EBs in differentiation for 20 days.

8.3.8. Change 2 mL of fresh EB medium every two days.

8.4. qRT-PCR for gene expression analysis

8.4.1. Culture the iPSC colonies for 5-6 days in iPSC medium with 10 ng/mL bFGF.

8.4.2. Extract total RNA from PBMNCs, iPSCs or EBs using 1 mL of commercial reagent in

accordance with manufacturer's instructions.

8.4.3. Evaluate RNA concentration and purity by suitable methods such as use of a spectrophotometer (high quality RNA with A_{260}/A_{280} and A_{260}/A_{230} ratios > 1.8)²⁷.

8.4.4. Check RNA integrity using a commercial kit according to manufacturer's protocol (high quality RNA RQI > 9.5)²⁸.

8.4.5. Perform a reverse transcription reaction on 1 μ g of total RNA using a commercial reverse transcriptase kit according to manufacturer's instructions.

8.4.6. Prepare the qPCR mixtures containing 5 ng of cDNA template, 7.5 μ L SYBR GREEN Supermix, 2 μ L of 5 μ M primers (forward: 1 μ L and reverse: 1 μ L) and 6 μ L of RNase/DNase-free water for each reaction²⁹.

8.4.7. Perform the qRT-PCR using the following program: an initial denaturation step at 95° C for 10 min, followed by 40 cycles divided into a denaturation step at 95°C for 15 s and primer annealing and extension step at 60°C for 45 s²⁹.

8.4.8. Normalize the expression of other genes using GAPDH as housekeeping gene²⁸ (see the primers listed in Table 1).

8.5. qPCR for transgene exclusion

8.5.1. Extract DNA from PBMNCs or iPSCs using 1 mL of lysis buffer containing 50 mM Tris, 100 mM EDTA, 100 mM NaCl, 1% Sodium Dodecyl Sulphate (SDS) and 20 μ g/mL Proteinase K.

8.5.2. Add an equal volume (1 mL) of 100% isopropanol, mix by inversion three times in order to allow DNA precipitation and centrifuge at 10000 x g for 5 min at room temperature.

8.5.3. Discard the supernatant, wash the DNA pellet with 1 mL of 70% ethanol and centrifuge at 10000 x g for 5 min at room temperature. Aspirate and discard the supernatant and resuspend it in RNase/DNase-free water.

8.5.4. Evaluate DNA concentration and purity by suitable methods such as use of a spectrophotometer (high quality RNA with A_{260}/A_{280} and A_{260}/A_{230} ratios > 1.8)²⁷.

8.5.5. Prepare the qPCR mixtures containing 5 ng of DNA template, 7.5 μ L SYBR GREEN Supermix, 2 μ L of 5 μ M primers (forward: 1 μ L and reverse: 1 μ L) and 6 μ L of RNase/DNase-free water for each reaction²⁹.

8.5.6. Normalize the expression of specific episomal plasmid EBNA-1 gene using FBX-15 as housekeeping gene²⁹(see the primers listed in Table 1).

8.6. Karyotype analysis

8.6.1. Culture the iPSC colonies for 5-6 days onto feeder-free basement membrane matrix coated plate with a commercial defined, feeder-free maintenance medium for iPSCs, following manufacturer's instructions.

8.6.2. To start the karyotype analysis protocol, detach iPSC colonies with 1 mL of cell detachment solution for 5 min at 37 °C, until obtaining single-cell dissociation.

8.6.3. Collect cells and centrifuge at 400 x g for 5 min at room temperature.

8.6.4. Resuspend iPSCs in 2 mL of a medium specifically developed for culturing cells for prenatal diagnostic techniques. Plate single-cell iPSCs onto basement membrane matrix coated glass dishes and incubate them in a 37 °C, 5% CO₂ incubator.

8.6.5. After 2-4 days, add 10 µL/mL colchicine directly to the cultures and incubate for 3 h at 37 °C, 5% CO₂ in a humidified incubator.

8.6.6. Aspirate the medium and incubate iPSCs in 1 mL of a hypotonic solution (0.6% sodium citrate and 0.13% potassium chloride) for 10 min at room temperature.

8.6.7. Rinse cells with 1 mL of 5% acetic acid solution and fix iPSCs with methanol/acetic acid solution (ratio 3:1) in a machine with controlled temperature and humidity (28 °C, 42% rH).

8.6.8. Immerse and stain fixed cells with Quinacrine solution for 15-20 min at room temperature in dark (to obtain Q-banding)³⁰.

8.6.9. Acquire cell metaphases under a fluorescence microscope at 100X magnification²⁹.

REPRESENTATIVE RESULTS:

In this study a simple and effective protocol for the generation of viral-free iPSCs by reprogramming of PBMNCs isolated from frozen buffy coats after whole blood centrifugation and comparison with reprogramming of PBMNCs obtained after density gradient separation is reported. **Figure 1A** shows a schematic representation of the detailed protocol. After thawing, the isolated PBMNCs, showing a typical rounded shape, are expanded in specific blood culture medium for 14 days (**Figure 1B**) and then transfected with episomal plasmids. After 10-15 days post-transfection, small rounded bright colonies with defined margins and embryonic stem cell-like morphology start to appear (**Figure 1C**). Approximately 20-30 days after transfection, iPSC colonies become highly compact, with sharp edges, increase their size and are ready for picking and expansion (**Figure 1D**). After the first passaging steps (2-3 passages), iPSC colonies maintain their undifferentiated state and show a typical human embryonic-stem cell morphology. During the first passages, manual picking is a more efficient expansion way compared to enzyme dissociation in order to select fully undifferentiated, compact and tightly packed colonies (**Figures 2A**). Larger magnification of iPSC colonies shows individual cells within the colonies and

the high nucleus to cytoplasm ratio is more easily observed (**Figure 2B**). After initial mechanical picking, selected iPSC clones are expanded with enzyme dissociation using Collagenase IV. After 10-15 passages of *in vitro* expansion, cytogenetic analysis is conducted on iPSC colonies. About 20 metaphases from at least two independent cultures, at approximately 300–400 band level, are analyzed and all samples show a normal karyogram (**Figure 2C**). This observation suggests that iPSCs are genetically stable.

After *in vitro* expansion (5-10 passages), iPSCs are characterized for the expression of stemness markers and their pluripotency potential. **Figures 3A, B** show that iPSCs are positive for alkaline phosphatase staining. Using qRT-PCR analysis, we have verified that iPSCs express endogenous pluripotent genes (SOX-2, OCT3/4, NANOG) at significantly higher levels than starting PBMNCs, while expression levels of endogenous C-MYC and KLF-4 are similar before and after iPSC reprogramming (**Figure 3C**). After only 5 passages in culture, expression of exogenous episomal transgenes cannot be detected in iPSCs, thus indicating the successful activation of endogenous pluripotent genes. PBMNCs 5 days after transfection, with strong transgene expression level, evaluated by episomal specific primers for EBNA-1, are used as positive control. PBMNCs that are never exposed to plasmids carrying the 4 exogenous pluripotency genes, are used as negative control(**Figure 3D**). Finally, immunofluorescence analysis reveals that iPSCs express pluripotency markers, such as OCT3/4, SOX-2, SSEA-4, TRA-1-60, and TRA-1-81 (**Figure 4**).

iPSCs are differentiated into embryoid bodies (EBs) using a spontaneous differentiation protocol, as shown in **Figure 5A**, in order to evaluate their *in vitro* pluripotency potential. qRT-PCR experiments reveal that iPSCs are able to differentiate into cells of the three germ layers; indeed, EBs obtained after 20 days of differentiation display significant up-regulation of lineage markers representative of the endoderm (SOX-7, AFP), mesoderm (CD31, ACTA-2, CDH-5, RUNX-1) and ectoderm (KTR-14, TH, GABRR-2) (**Figure 5B**). Confocal microscopy analysis shows that single-cell dissociated beating clusters express typical cardiomyocyte markers such as Troponin I (Tn-I) and α -Actinin, organized in a clear sarcomeric pattern (**Figure 6A**). In addition, immunofluorescence analysis of EBs shows a positive staining for the neuron-specific marker class III β -Tubulin (TUJ1) as well for the dopaminergic neuronal marker tyrosine hydroxylase (TH) (**Figure 6B**).

Figure 1: Protocol for the generation of iPSCs from human peripheral blood and cell morphological changes during the protocol. (A) Diagram of the protocol used to generate iPSCs obtained from frozen PBMNCs after DGS and PBMNCs isolated from frozen buffy coats, using a single transfection of non-integrating episomal plasmids. **(B)** Representative images of proliferating PBMNCs obtained from DGS and buffy coats after thawing and expansion in culture for 14 days, before being transfected. Scale bar 100 μ m. **(C)** Examples of iPSC colonies one week after the cells are plated on MEFs. Scale bar 500 μ m. **(D)** Representative images of iPSC colonies at 30-35 days after re-plating on a MEF feeder-layer. Scale bar 500 μ m. iPSCs = induced pluripotent stem cells; DGS= density gradient separation; PBMNCs = peripheral blood mononuclear cells; MEFs = mouse embryonic fibroblasts; GFs = growth factors; bFGF = basic fibroblast growth factor; NaB = sodium butyrate.

Figure 2: Morphology and karyotype analysis (A) Representative images of iPSC colonies after 2-3 manual passaging steps. Scale bar 500 μm . (B) Larger magnification of iPSC colonies. Scale bar 200 μm . (C) Representative normal karyograms from iPSCs after 10-15 passages of *in vitro* expansion.

Figure 3: iPSC characterization: alkaline phosphatase activity, re-expression of endogenous pluripotency genes of iPSCs and evaluation of transgene silencing. (A) and (B) Representative images showing the positive staining for alkaline phosphatase. Scale bar 500 μm . (C) The bar graph showing the re-expression of endogenous pluripotency genes (C-MYC, KLF-4, SOX-2, OCT3/4 and NANOG) in both iPSCs obtained from DGS and buffy coats, using qRT-PCR analysis (n=4, Student t-test : * p<0.05, § p<0.005 vs corresponding PBMNCs). (D) qRT-PCR with episomal-specific primers (EBNA-1) (relative to control FBOX15 primers) shows no genome-integration of episomal vectors. (n=4, Student t-test: * p<0.001 vs corresponding PBMNCs 5-day transfection).

Figure 4: Immunofluorescence analysis of embryonic stem cells pluripotency markers. Representative immunofluorescence staining showing significant expression of pluripotency proteins SSEA-4, TRA-1-60, TRA-1-81, OCT3/4 and SOX-2. Nuclei are counterstained with DAPI. Scale bar 200 μm for iPSC DGS Figure 4A and iPSC Buffy coat Figure 4B. Scale bar 100 μm for iPSC Buffy coat Figure 4A, iPSC DGS Figure 4B and Figure 4C.

Figure 5: Protocol for the differentiation of iPSCs and expression of three germ layer genes in EBs obtained from iPSCs. (A) Schematic representation of the protocol inducing spontaneous differentiation of iPSCs from PBMNCs after DGS and buffy coats in EBs. (B) qRT-PCR experiments showing the expression of all three germ layer genes: endoderm (SOX-7, AFP), mesoderm (CD31, ACTA-2, CDH-5, RUNX-1), and ectoderm (KRT-14, TH, GABRR-2) in EBs after 20 days of differentiation (n=4, Student t-test: * p<0.05 vs corresponding iPSCs counterpart). EBs = embryoid bodies.

Figure 6: Confocal microscopy analysis of cardiac and neuronal proteins in EBs at day 20 of differentiation obtained from PBMNCs after DGS and buffy coats-derived iPSCs. (A) Representative images (maximum projection of confocal image stack) for cardiac sarcomeric proteins α -Actinin and Troponin I (Tn-I) in single-cell dissociated beating areas (scale bar 10 μm) and (B) for neuron-specific marker class III β -Tubulin (TUJ1) and the dopaminergic neuronal marker tyrosine hydroxylase (TH) (scale bar 20 μm). Nuclei are counterstained with DAPI.

Table 1: List of qRT-PCR primers. Primer sequences used for evaluation of endogenous pluripotency gene expression, transgene silencing and expression of three germ layer markers.

DISCUSSION:

In the past, the only way to obtain human pluripotent stem cells carrying a particular genetic mutation was to recruit parents undergoing pre-implantation genetic diagnosis and generate embryonic stem cells from their discarded blastocysts^{31, 32}. Using a reprogramming approach, researchers can now generate iPSCs from patients carrying virtually any genotype. The

possibility to start from a patient-specific cell line and an easily accessible source is very important since it allows an investigation of the pathophysiology of disorders and subsequent identification of novel therapeutic approaches.

In the present study, we combined a plasmid approach¹⁵ with a method already applied to produce iPSCs from PBMNCs previously frozen after density gradient separation²² and successfully generated virus-free iPSCs from frozen buffy coat PBMNCs. The use of buffy coats instead of density gradient separated PBMNCs allows us to reduce costs, working time and manual steps for operators during sample processing. The set-up of a low-cost protocol using PBMNCs from frozen buffy coats is essential for studies with elevated number of participants and for sample collections in multi-center studies. Importantly, the ability to produce iPSCs from frozen buffy coats allows access to numerous frozen biosamples already stored in blood banks, using a simple, cost-effective and not so time consuming method for sample collection.

This induction method may be useful for the derivation of patient-specific integration-free iPSCs, where colonies are characterized by the absence of episomal transgene after only few passages (≥ 5 passages), in both starting materials. Remarkably, we observed that all the selected iPSC colonies obtained from both types of starting material display comparable pluripotency features and the preservation of cellular and molecular similarities to hESCs, indicating a successful iPSC derivation and a consistent reproducibility of the protocol.

Moreover, we have successfully generated iPSCs from more than 10 human skin fibroblast and 3 cardiac mesenchymal stromal cell lines (both from healthy donors and patients with cardiac and skeletal muscle diseases) using this method (data not shown), thus suggesting that the described protocol can be used for the successful reprogramming of different cell types. However, the choice of blood as starting material is advantageous compared to fibroblasts, especially when considering that extensive expansion of fibroblasts *in vitro* may affect the efficiency of reprogramming, based on fibroblast passage number and proliferation rate^{33, 34}. Our protocol also allows the generation of transgene-free iPSCs from frozen buffy coats after a minimal culture time, thus reducing by about 3-4 weeks the time needed for derivation of iPSCs compared to other sources, such as fibroblasts. Furthermore, recent studies revealed that blood cell-derived iPSCs display an epigenetic signature that more closely resembles hESCs than those obtained from mesenchymal stem cells (MSC)/fibroblasts^{14, 35}.

Despite of the successful generation of iPSCs from frozen buffy coats, a few limitations need to be considered in this protocol. Before reprogramming induction, the amplification of PBMNCs from frozen buffy coats could represent a critical step of the protocol. The quality of starting material can strongly influence the selection and isolation of PBMNCs, in order to achieve sufficient cell numbers for reprogramming procedure (at least 2×10^6 cells for electroporation). This is mostly due to intrinsic biological variability of the samples, but also to technical issues. Indeed, the amplification of PBMNCs from frozen buffy coats is less efficient compared to PBMNCs obtained from density gradient separation, due to a strong dependence on manual operator variability. In order to enhance reproducibility and efficiency of PBMNC amplification from frozen buffy coats without density gradient separation, the use of automated systems is

strongly recommended to collect buffy coat fractions. Although it is more difficult to expand a sufficient number of PBMNCs from buffy coats than PBMNCs after density gradient separation, the efficiency of reprogramming (about 0.001%-0.0005%) of the two starting materials is comparable, according to blood reprogramming efficiency previously reported^{14, 23}. Since the reprogramming efficiency is low for regenerative medicine purposes, the use of improved EBNA1/OriP (from Epstein-Barr virus, EBV) episomal vectors may increase the number of iPSC colonies for future cell therapy development^{22, 36}. Additionally, a MEF feeder layer for iPSC generation and maintenance is used in our protocol. In order to obtain clinical-grade iPSCs, a feeder-free culture could be an alternative method for further studies.

In conclusion, this protocol represents a valid method to generate iPSCs from an easily accessible patient cell source with the great advantage of using a non-integrative system that can potentially be used for the derivation of clinical-grade iPSCs, not only for disease modelling but also for a future personalized medicine.

DISCLOSURES:

The authors have nothing to disclose.

ACKNOWLEDGMENTS:

The study was supported by the Ministry of Health and Department of Educational Assistance, University and Research of the Autonomous Province of Bolzano and the South Tyrolean Sparkasse Foundation.

REFERENCES:

1. Takahashi, K. & Yamanaka, S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*. **126** (4), 663-676, doi: 10.1016/j.cell.2006.07.024(2006).
2. Takahashi, K., Okita, K., Nakagawa, M. & Yamanaka, S. Induction of pluripotent stem cells from fibroblast cultures. *Nat. Protoc.* **2** (12), 3081-3089, doi: 10.1038/nprot.2007.418 (2007).
3. Drews, K., Jozefczuk, J., Prigione, A. & Adjaye, J. Human induced pluripotent stem cells--from mechanisms to clinical applications. *J. Mol. Med. (Berl)* **90** (7), 735-745, doi: 10.1007/s00109-012-0913-0 (2012).
4. Bayart, E. & Cohen-Haguenauer, O. Technological overview of iPS induction from human adult somatic cells. *Curr. Gene Ther.* **13** (2), 73-92, doi: 10.2174/1566523211313020002 (2013).
5. Takahashi, K. *et al.* Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell*. **131** (5), 861-872, doi:10.1016/j.cell.2007.11.019 (2007).
6. Sommer, A.G. *et al.* Generation of human induced pluripotent stem cells from peripheral blood using the STEMCCA lentiviral vector. *J. Vis. Exp.* (68), e4327, doi: 10.3791/4327 (2012).
7. Zhou, W. & Freed, C. R. Adenoviral gene delivery can reprogram human fibroblasts to induced pluripotent stem cells. *Stem Cells*. **27** (11), 2667-2674, doi: 10.1002/stem.201 (2009).

8. Lieu, P.T., Fontes, A., Vemuri, M.C. & Macarthur, C.C. Generation of induced pluripotent stem cells with CytoTune, a non-integrating Sendai virus. *Methods Mol. Biol.* **997**, 45-56, doi: 10.1007/978-1-62703-348-0_5 (2013).
9. Woltjen, K. *et al.* piggyBac transposition reprograms fibroblasts to induced pluripotent stem cells. *Nature*. **458** (7239), 766-770, doi: 10.1038/nature07863 (2009).
10. Okita, K. *et al.* A more efficient method to generate integration-free human iPS cells. *Nat. Methods*. **8** (5), 409-412, doi: 10.1038/nmeth.1591 (2011).
11. Kim, D. *et al.* Generation of human induced pluripotent stem cells by direct delivery of reprogramming proteins. *Cell. Stem Cell*. **4** (6), 472-476, doi: 10.1016/j.stem.2009.05.005 (2009).
12. Warren, L., Ni, Y., Wang, J. & Guo, X. Feeder-free derivation of human induced pluripotent stem cells with messenger RNA. *Sci. Rep.* **2**, 657, doi: 10.1038/srep00657 (2012).
13. Stadtfeld, M. & Hochedlinger, K. Induced pluripotency: history, mechanisms, and applications. *Genes Dev.* **24** (20), 2239-2263, doi: 10.1101/gad.1963910 (2010).
14. Chou, B. K. *et al.* Efficient human iPS cell derivation by a non-integrating plasmid from blood cells with unique epigenetic and gene expression signatures. *Cell Res.* **21** (3), 518-529, doi: 10.1038/cr.2011.12 (2011).
15. Kim, C. *et al.* Studying arrhythmogenic right ventricular dysplasia with patient-specific iPSCs. *Nature*. **494** (7435), 105-110, doi: 10.1038/nature11799 (2013).
16. Aasen, T. *et al.* Efficient and rapid generation of induced pluripotent stem cells from human keratinocytes. *Nat. Biotechnol.* **26** (11), 1276-1284, doi: 10.1038/nbt.1503 (2008).
17. Streckfuss-Bomeke, K. *et al.* Comparative study of human-induced pluripotent stem cells derived from bone marrow cells, hair keratinocytes, and skin fibroblasts. *Eur. Heart J.* **34** (33), 2618-2629, doi: 10.1093/eurheartj/ehs203 (2013).
18. Miyoshi, N. *et al.* Reprogramming of mouse and human cells to pluripotency using mature microRNAs. *Cell. Stem Cell*. **8** (6), 633-8, doi: 10.1016/j.stem.2011.05.001 (2011).
19. Wang, Y. *et al.* Induced pluripotent stem cells from human hair follicle mesenchymal stem cells. *Stem Cell. Rev.* **9** (4), 451-460, doi: 10.1007/s12015-012-9420-5 (2013).
20. Beltrao-Braga, P. C. *et al.* Feeder-free derivation of induced pluripotent stem cells from human immature dental pulp stem cells. *Cell Transplant.* **20** (11-12), 1707-1719, doi: 10.3727/096368911X566235 (2011).
21. Haase, A. *et al.* Generation of induced pluripotent stem cells from human cord blood. *Cell. Stem Cell*. **5** (4), 434-441, doi: 10.1016/j.stem.2009.08.021 (2009).
22. Dowey, S. N., Huang, X., Chou, B. K., Ye, Z. & Cheng, L. Generation of integration-free human induced pluripotent stem cells from postnatal blood mononuclear cells by plasmid vector expression. *Nat. Protoc.* **7** (11), 2013-2021, doi: 10.1038/nprot.2012.121 (2012).
23. Staerk, J. *et al.* Reprogramming of human peripheral blood cells to induced pluripotent stem cells. *Cell. Stem Cell*. **7** (1), 20-24, doi: 10.1016/j.stem.2010.06.002 (2010).
24. Geti, I. *et al.* A practical and efficient cellular substrate for the generation of induced pluripotent stem cells from adults: blood-derived endothelial progenitor cells. *Stem Cells Transl. Med.* **1** (12), 855-865, doi: 10.5966/sctm.2012-0093 (2012).
25. Strober, W. Trypan blue exclusion test of cell viability. *Curr Protoc Immunol.* Appendix 3, Appendix 3B, doi: 10.1002/0471142735.ima03bs21 (2001).

26. Hasegawa, K. *et al.* Comparison of reprogramming efficiency between transduction of reprogramming factors, cell-cell fusion, and cytoplasm fusion. *Stem Cells*. **28** (8), 1338-1348, doi: 10.1002/stem.466 (2010).
27. Desjardins, P. & Conklin, D. NanoDrop microvolume quantitation of nucleic acids. *J Vis Exp.* (45), 2565, doi: 10.3791/2565 (2010).
28. Fleige, S. & Pfaffl, M. W. RNA integrity and the effect on the real-time qRT-PCR performance. *Mol Aspects Med.* **27** (2-3), 126-39, doi: 10.1016/j.mam.2005.12.003 (2006).
29. Meraviglia, V. *et al.* Human chorionic villus mesenchymal stromal cells reveal strong endothelial conversion properties. *Differentiation*. **83** (5), 260-270, doi: 10.1016/j.diff.2012.02.006 (2012).
30. Caspersson, T., Zech, L. & Johansson, C. Analysis of human metaphase chromosome set by aid of DNA-binding fluorescent agents. *Exp Cell Res.* **253** (2), 302-304, doi: 10.1006/excr.1999.4745 (1999).
31. Alikani, M. & Munne, S. Nonviable human pre-implantation embryos as a source of stem cells for research and potential therapy. *Stem Cell. Rev.* **1** (4), 337-343, doi: 10.1385/SCR:1:4:337 (2005).
32. Tropel, P. *et al.* High-efficiency derivation of human embryonic stem cell lines following pre-implantation genetic diagnosis. *In Vitro Cell. Dev. Biol. Anim.* **46** (3-4), 376-385, doi: 10.1007/s11626-010-9300-8 (2010).
33. Hanna, J. *et al.* Direct cell reprogramming is a stochastic process amenable to acceleration. *Nature*. **462** (72-73), 595-601, doi: 10.1038/nature08592 (2009).
34. Li, H. *et al.* The Ink4/Arf locus is a barrier for iPS cell reprogramming. *Nature*. **460** (7259), 1136-1139, doi: 10.1038/nature08290 (2009).
35. Kim, K. *et al.* Epigenetic memory in induced pluripotent stem cells. *Nature*. **467** (7313), 285-290, doi: 10.1038/nature09342 (2010).
36. Lindner, S. E. & Sugden, B. The plasmid replicon of Epstein-Barr virus: mechanistic insights into efficient, licensed, extrachromosomal replication in human cells. *Plasmid*. **58** (1), 1-12, doi: 10.1016/j.plasmid.2007.01.003 (2007).

Figure 1
[Click here to download Figure: Figure 1_revised.pdf](#)

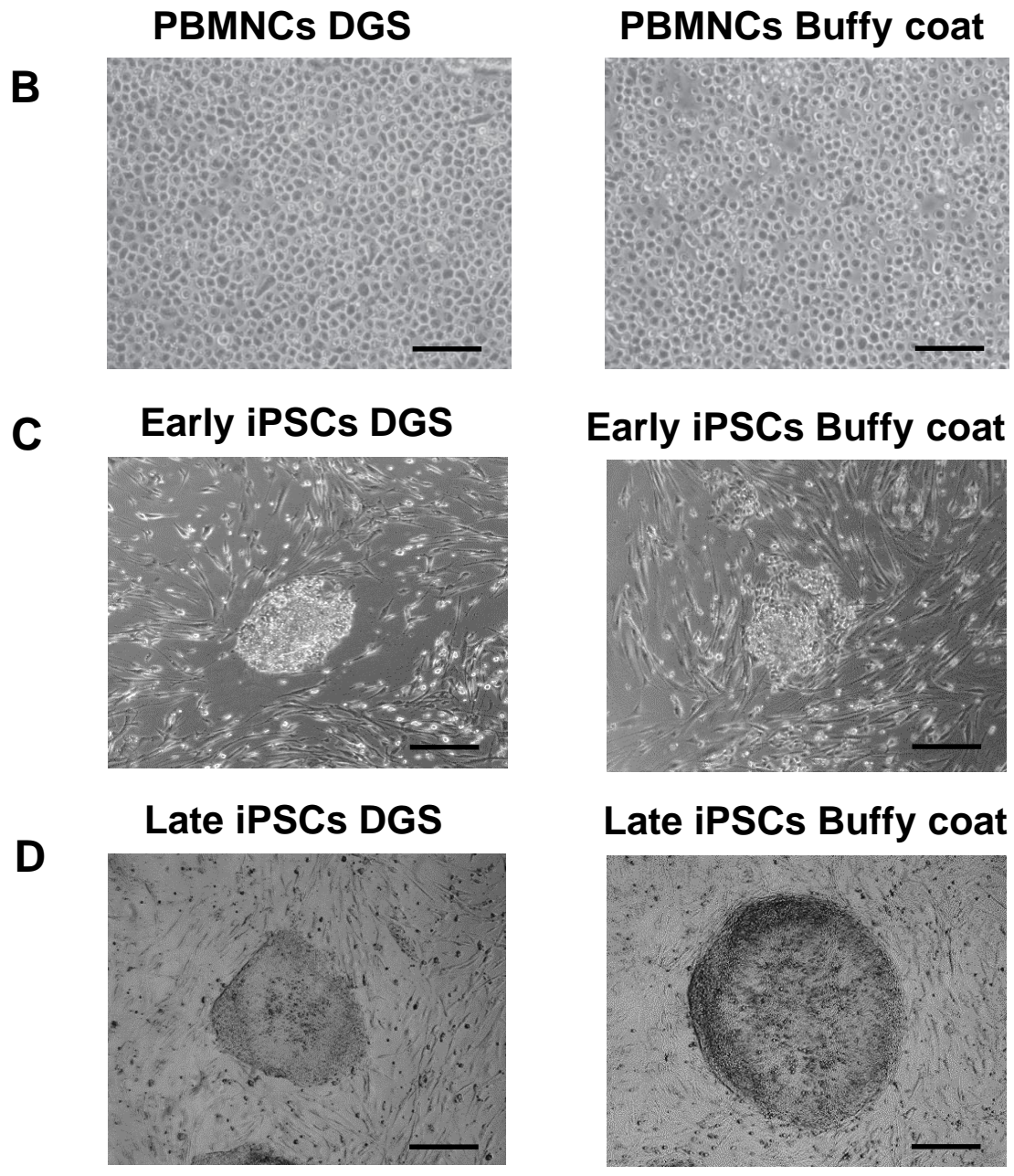
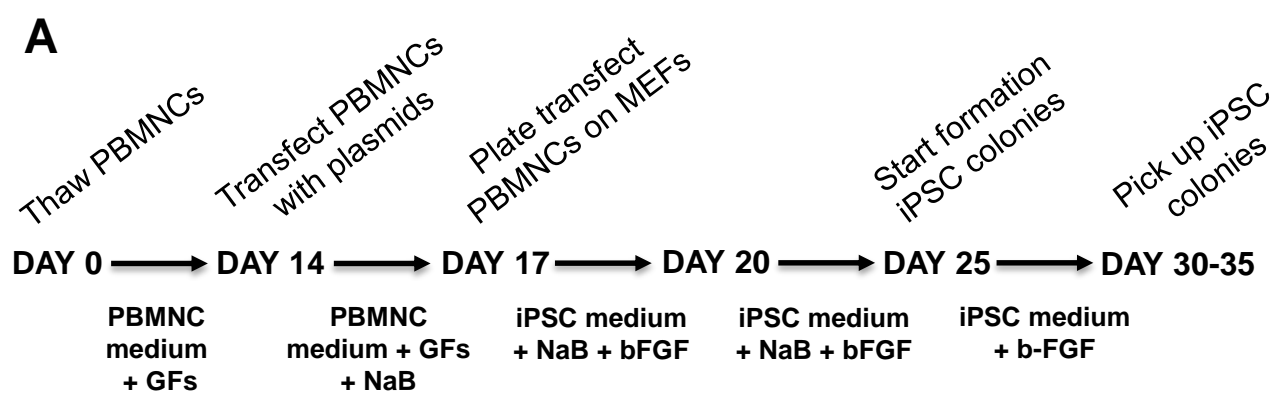
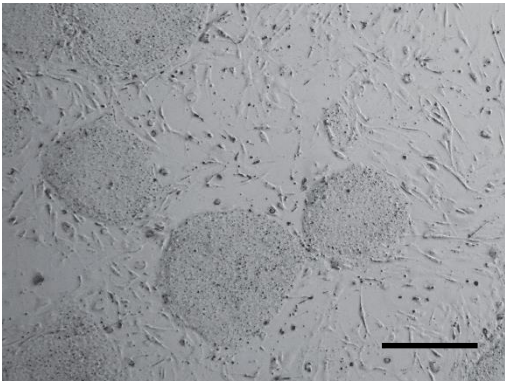
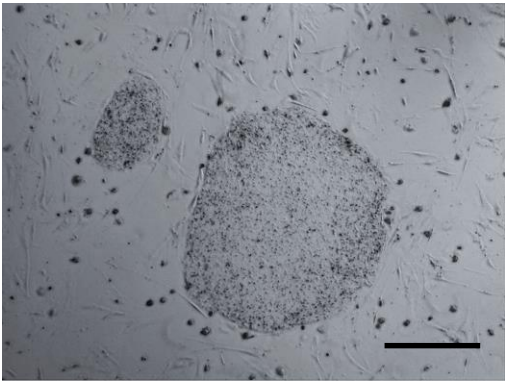


Figure 2
[Click here to download Figure: Figure 2_revised.pdf](#)

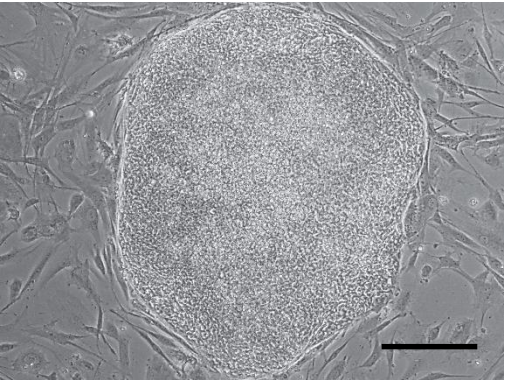
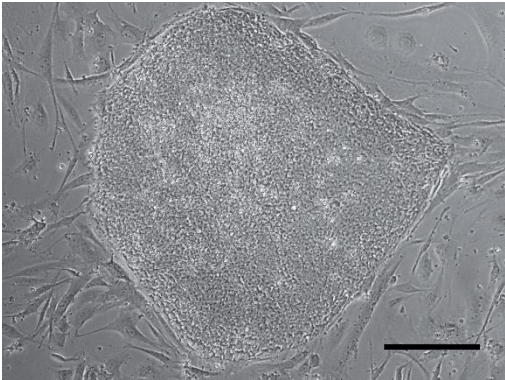
iPSCs DGS

iPSCs Buffy coat

A



B



C

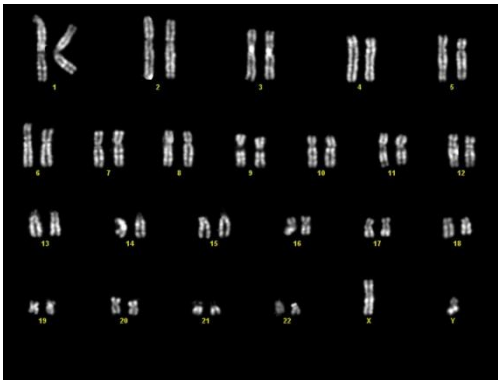
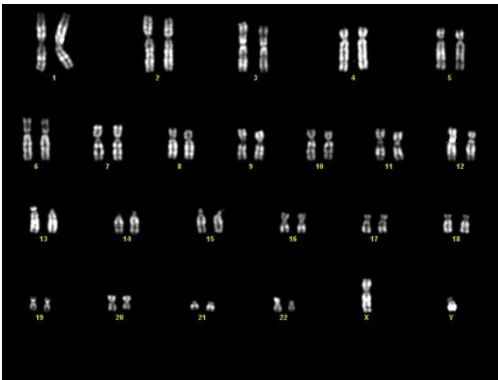


Figure 3
[Click here to download Figure: Figure 3_revised.pdf](#)

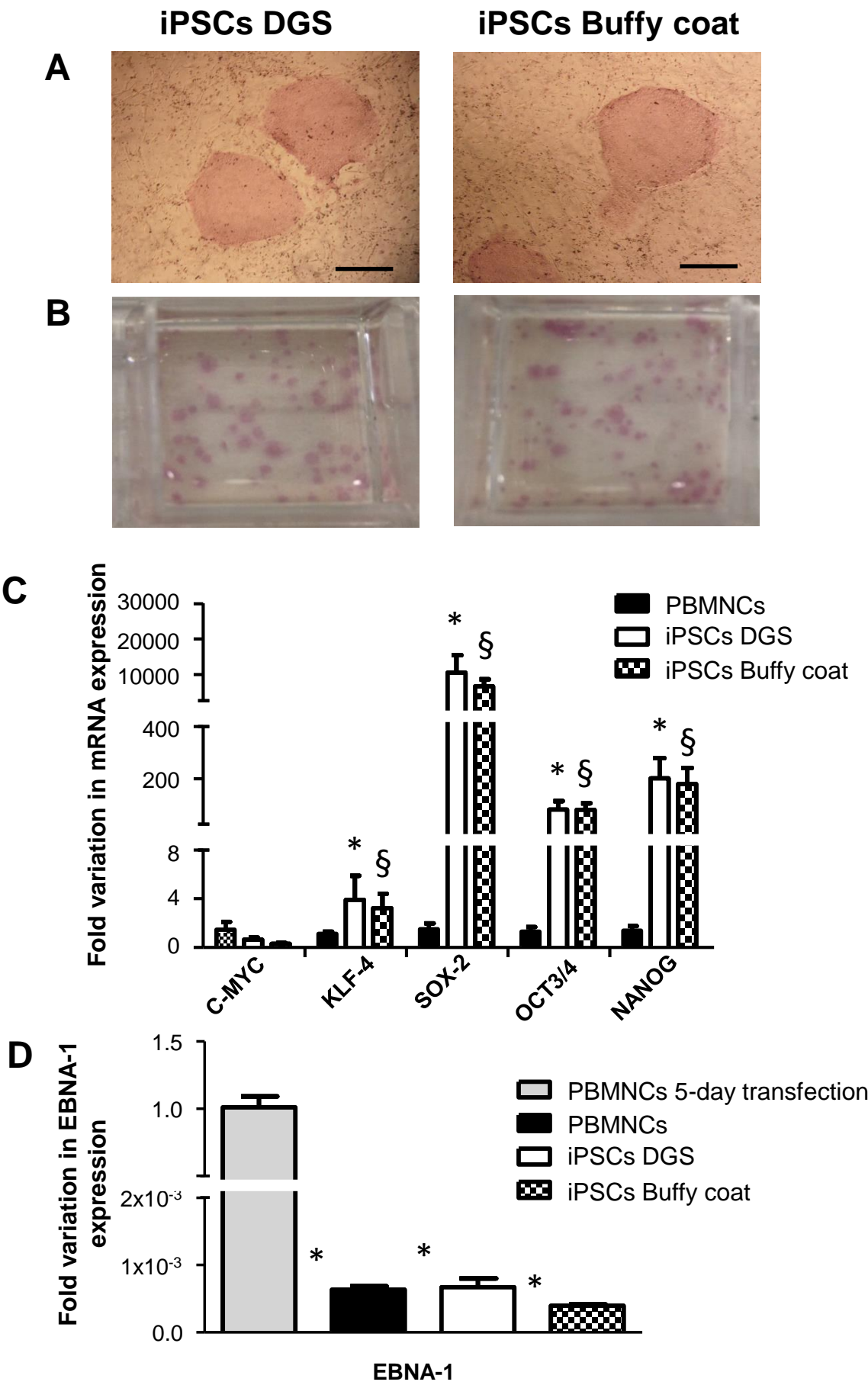


Figure 4
[Click here to download Figure: Figure 4_revised.pdf](#)

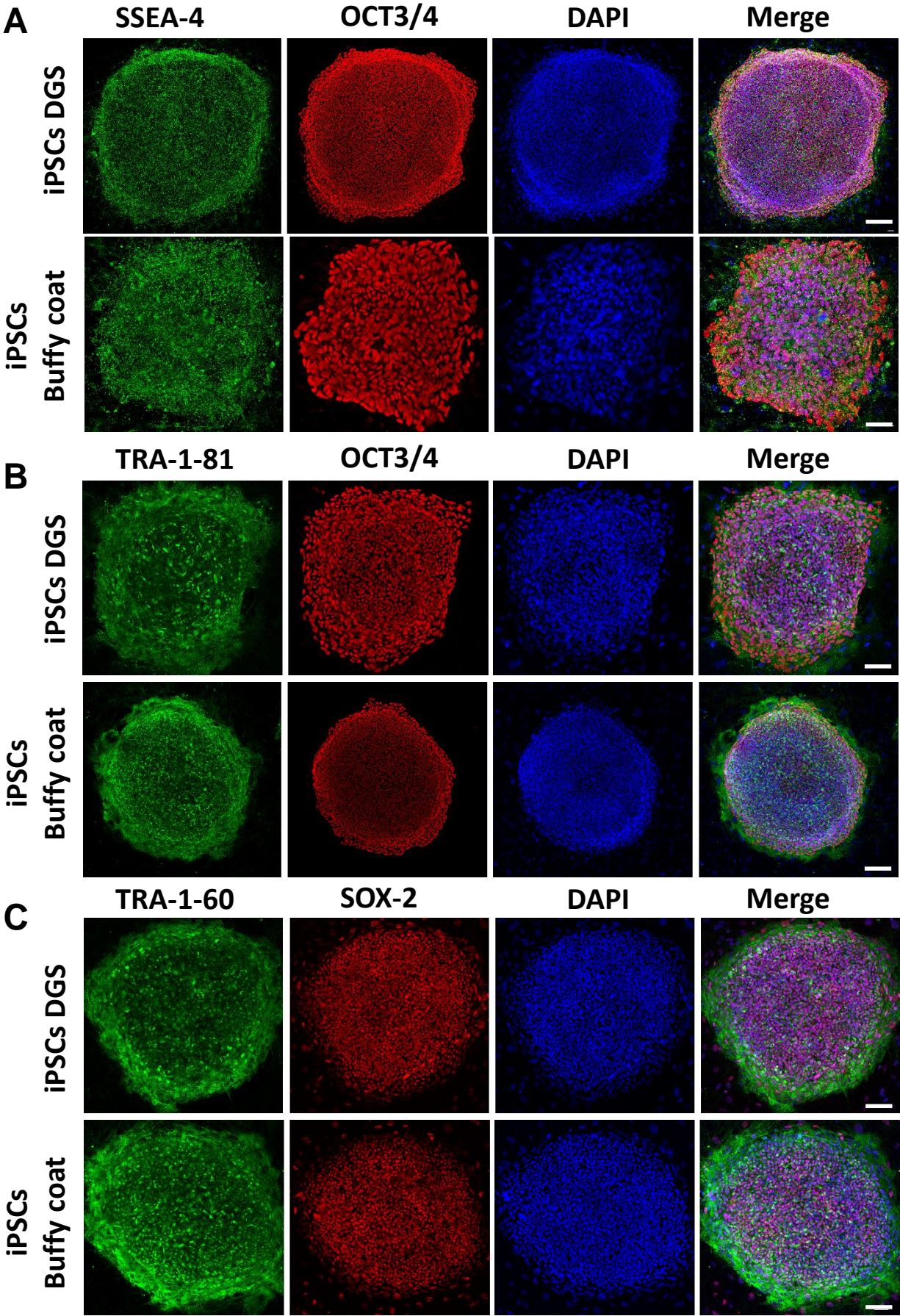
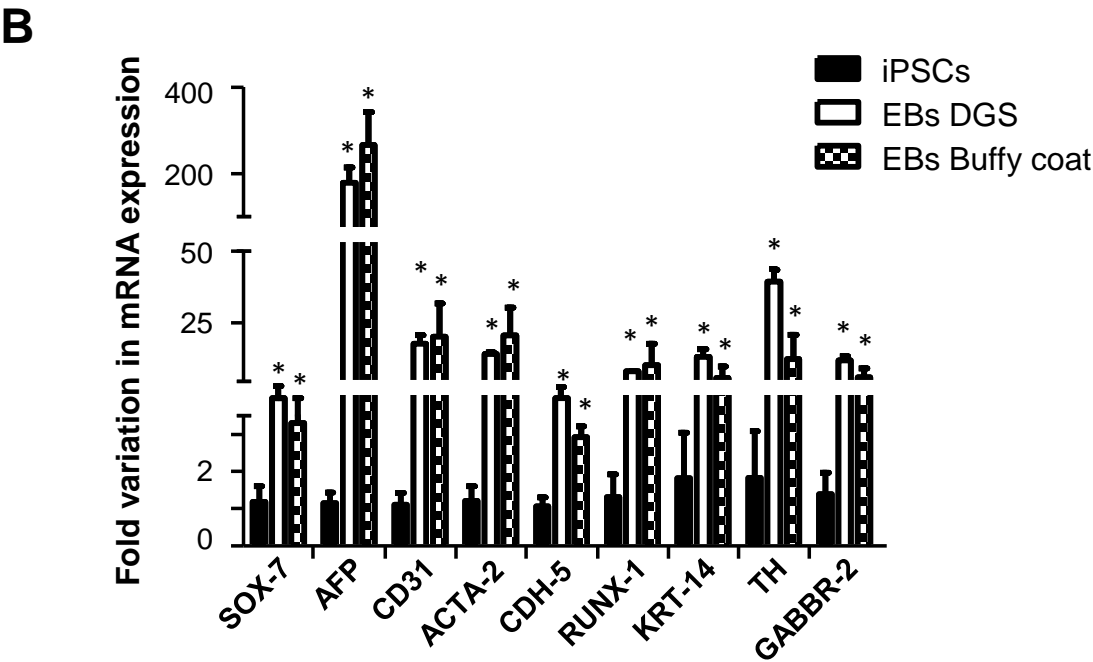
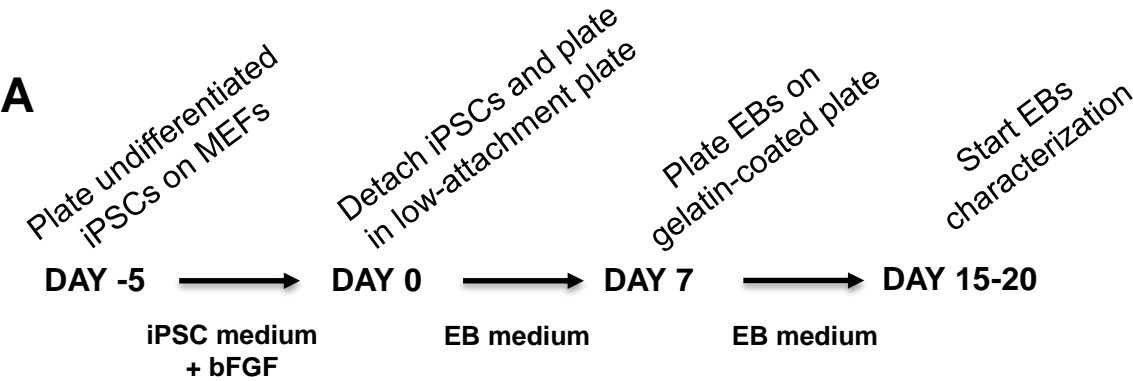


Figure 5
Click here to download Figure: Figure 5_revised.pdf



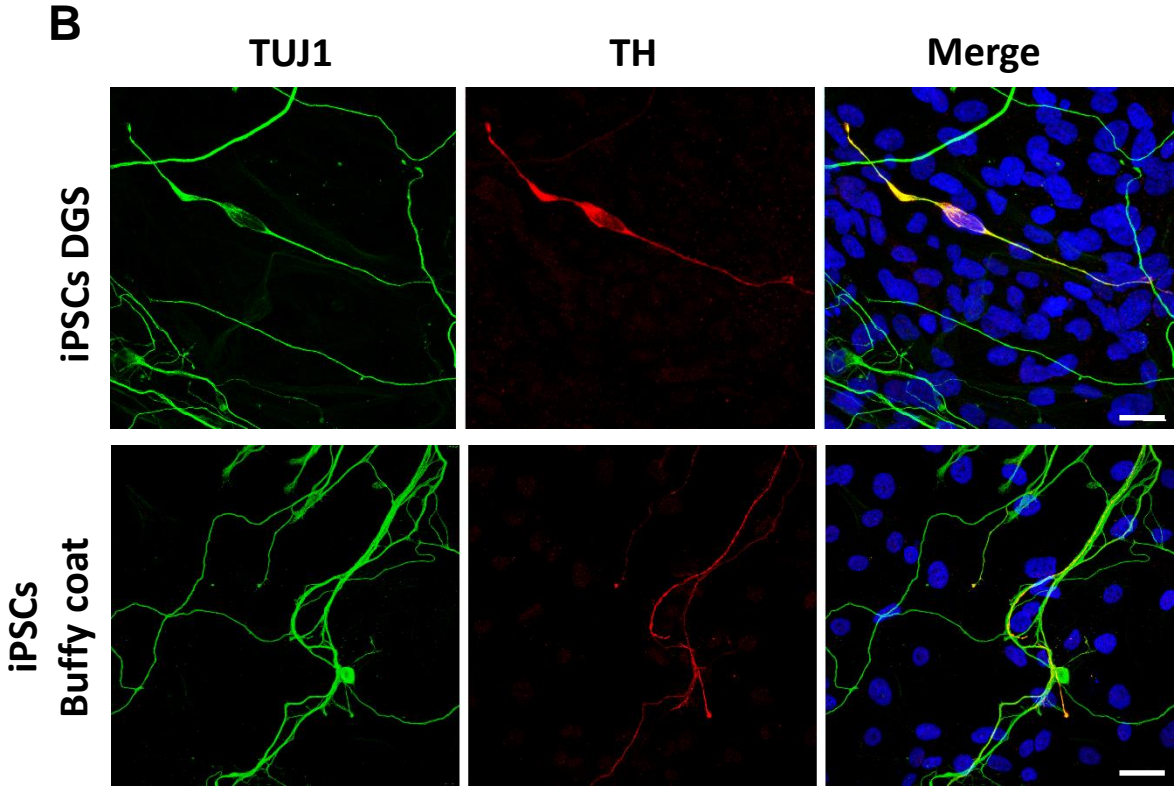
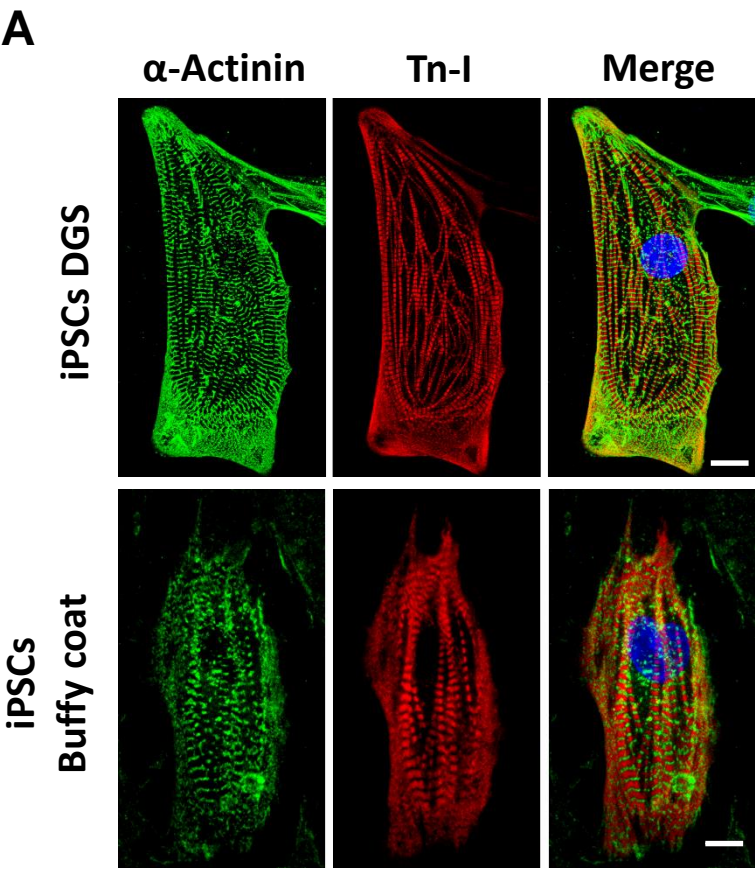


Table 1
[Click here to download Table: Primer list.xlsx](#)

Primer	Sequence
C-MYC fw	5'-AGAAATGTCCTGAGCAATCACC-3'
C-MYC rev	5'-AAGGTTGTGAGGTTGCATTTGA-3'
KLF-4 fw	5'-ATAGCCTAAATGATGGTGCTTGG-3'
KLF-4 rev	5'- AACTTTGGCTTCCTTGTGG-3'
SOX-2 fw	5'- GGGAAATGGGAGGGGTGCAAAAGAGG-3'
SOX-2 rev	5'- TTGCGTGAGTGTGGATGGGATTGGTG-3
OCT3/4 fw	5'- GACAGGGGGAGGGGAGGAGCTAGG-3'
OCT3/4 rev	5'- CTTCCCTCCAACCAAGTTGCCCAAAC-3'
NANOG fw	5'-TGCAAGAACTCTCCAACATCCT-3'
NANOG rev	5'-ATTGCTATTCTTCGCCAGTT-3'
SOX-7 fw	5'-TGAACGCCTTCATGGTTTG-3'
SOX-7 rev	5'-AGCGCCTTCCACGACTTT-3'
AFP fw	5'-GTGCCAAGCTCAGGGTGTAG -3'
AFP rev	5'-CAGCCTCAAGTTGTTCTCTG-3'
CD31 fw	5'-ATGCCGTGGAAGCAGATAC-3'
CD31 rev	5'-CTGTTCTTCTCGGAACATGGA-3'
ACTA-2 fw	5'-GTGATCACCATCGGAAATGAA-3'
ACTA-2 rev	5'-TCATGATGCTGTTGTAGGTGGT-3'
CDH-5 fw	5'-GAGCATCCAGGCAGTGGTAG-3'
CDH-5 rev	5'-CAGGAAGATGAGCAGGGTGA-3'
RUNX-1 fw	CCCTAGGGGATGTTCCAGAT-3'
RUNX-1 rev	TGAAGCTTTTCCCTCTTCCA-3'
KRT-14 fw	5'-CACCTCTCCTCCTCCCAGTT-3'
KRT-14 rev	5'-ATGACCTTGGTGCGGATTT-3'
TH fw	5'-TGTA CTGGTTCACGGTGGAGT-3'
TH rev	5'-TCTCAGGCTCCTCAGACAGG-3'
GABBR-2 fw	5'-CTGTGCCTGCCAGAGTTTCA-3'
GABBR-2 rev	5'-ACGGCCTTGACGTAGGAGA-3'
EBNA-1 fw	5'-ATCAGGGCCAAGACATAGAGATG-3'
EBNA-1 rev	5'-GCCAATGCAACTTGGACGTT-3'
FBX-15 fw	5'-GCCAGGAGGTCTTCGCTGTA-3'
FBX-15 fw	5'-AATGCACGGCTAGGGTCAAA-3'
GAPDH fw	5'-CCACCCATGGCAAATTCC-3'
GAPDH rev	5'-TCGCTCCTGGAAGATGGTG-3'

Name of Reagent/ Equipment	Company	Catalog Number
Sodium Citrate buffered Venosafe Plastic Tube	Terumo	VF-054SBCS07
Ammodium chloride	Sigma-Aldrich	A9434
Potassium bicarbonate	Sigma-Aldrich	60339
EDTA disodium powder	Sigma-Aldrich	E5134
Ficoll-Paque Premium	GE Healthcare Life Sciences	17-5442-02
Iscove's modified Dulbecco's medium (IMDM)	Gibco	21056-023
Ham's F-12	Mediatech	10-080-CV
Insulin-Transferrin-Selenium-Ethanolamine (ITS -X)	Gibco	51500-056
Chemically Defined Lipid Concentrate	Gibco	11905031
Bovine Serum Albumin (BSA)	Sigma-Aldrich	A9418
L-Ascorbic acid 2-phosphate sesquimagnesium salt hydrate	Sigma-Aldrich	A8960
1-Thioglycerol	Sigma-Aldrich	M6145
Recombinant Human Stem Cell Factor (SCF)	PeproTech	300-07
Recombinant Human Interleukin-3 (IL-3)	PeproTech	200-03
Recombinant Human Insulin-like Growth Factor (IGF-1)	PeproTech	100-11
Recombinant Human Erythropoietin (EPO)	R&D Systems	287-TC-500
Dexamethasone	Sigma-Aldrich	D2915
Human Holo-Transferrin	R&D Systems	2914-HT
Amniomax II	Gibco	11269016
mTeSR1	StemCell Technologies	5850
Knockout DMEM	Gibco	10829-018
Knockout Serum Replacement	Gibco	10828-028
Penicillin-Streptomycin (10,000 U/mL)	Gibco	15140-122
L-Glutamine (200 mM)	Gibco	25030-024

MEM Non-Essential Amino Acids Solution (100X)	Gibco	11140-050
2-Mercaptoethanol	Gibco	31350-010
Sodium Butyrate	Sigma-Aldrich	B5887
Recombinant Human FGF basic, 145 aa	R&D Systems	4114-TC
Y-27632 dihydrochloride	Sigma-Aldrich	Y0503
Fetal Defined Bovine Serum	Hyclone	SH 30070.03
EmbryoMax 0.1% Gelatin Solution	Merck-Millipore	ES-006-B
Matrigel Basement Membrane Matrix Growth	BD Biosciences	354230
Factor Reduced		
Collagenase, Type IV	Gibco	17104-019
Accutase	PAA Laboratories GmbH	L11-007
Mouse Embryonic Fibroblast (CF1)	Global Stem	GSC-6201G
Plasmid pCXLE-hOCT3/4-shp53-F	Addgene	27077
Plasmid pCXLE-hSK	Addgene	27078
Plasmid pCXLE-hUL	Addgene	27080
Plasmid pCXLE-EGFP	Addgene	27082
Alkaline Phosphatase Staining Kit	Stemgent	00-0009
Anti-Stage-Specific Embryonic Antigen-4 (SSEA-4)	Merck-Millipore	MAB4304
Antibody		
Anti-TRA-1-60 Antibody	Merck-Millipore	MAB4360
Anti-TRA-1-81 Antibody	Merck-Millipore	MAB4381
Anti-Oct-3/4 Antibody	Santa Cruz Biotechnology	sc-9081
Anti-Nanog Antibody	Santa Cruz Biotechnology	sc-33759
Anti-Troponin I Antibody	Santa Cruz Biotechnology	sc-15368
Anti- α -Actinin (Sarcomeric) Antibody	Sigma-Aldrich	A7732
Neuronal Class III β -Tubulin (TUJ1) Antibody	Covance Research Products Inc	MMS-435P-100
Anti-Tyrosine Hydroxylase (TH) Antibody	Calbiochem	657012
Alexa Fluor 488 Goat Anti-Mouse	Molecular Probes	A-11029
Alexa Fluor 555 Goat Anti-Rabbit	Molecular Probes	A-21429
Ultra-Low Attachment Cell Culture 6-well plate	Corning	3471

Trizol Reagent	Ambion	15596-018
SuperScript VILO cDNA Synthesis Kit	Invitrogen	11754050
iTaq Universal SYBR Green Supermix	Bio-Rad	172-5124
CFX96 Real-Time PCR Detection System	Bio-Rad	185-5195
Experion Automated Electrophoresis System	Bio-Rad	700-7000
Experion RNA Highsense Analysis kit	Bio-Rad	7007105
Dissecting microscope (SteREO Discovery V12)	Zeiss	495007
NeonTransfection System 100 µL Kit	Invitrogen	MPK10025
Neon Transfection System	Invitrogen	MPK5000
NanoDrop UV/Vis Spectrophotometer	Thermo Scientific	ND-2000
EndoFree Plasmid Maxi Kit	Qiagen	12362

Comments/Description

0.5 M solution

Polysucrose solution for density gradient centrifugation

No phenol red

1X (Stock: 100X)

1X (Stock: 100X)

Final Concentration at 200 μ M

100 ng/ml (Stock: 100 μ g/ml)

10 ng/ml (Stock: 10 μ g/ml)

40 ng/ml (Stock: 40 μ g/ml)

2 U/ml (Stock: 50 U/ml)

1 μ M (Stock: 1 mM)

100 μ g/ml (Stock: 20 mg/ml)

Medium for cytogenetic analysis

Medium for iPSC feeder-free culture

0.1 mM (Stock: 50 mM)
0.25 mM (Stock: 0.5 M)
10 ng/ml (Stock: 10 µg/ml)
10 µM (Stock: 10 mM)

1 mg/ml (Stock: 10 mg/ml)

Cell detachment solution

1×10^6 cells/6 well plate
1 µg (Stock: 1 µg/µl)
1 µg (Stock: 1 µg/µl)
1 µg (Stock: 1 µg/µl)
1 µg (Stock: 1 µg/µl)

1/250

1/250

1/250

1/500

1/500

1/500

1/250

1/500

1/200

1/1000

1/1000

reagent for RNA extraction
Reverse transcriptase kit

Instrument to check RNA integrity
Reagent kit to check RNA integrity

Reagent kit for electroporation
Instrument used for electroporation
Instrument for DNA/RNA quantification
Plasmid purification kit



1 Alewife Center #200
Cambridge, MA 02140
tel. 617.945.9051
www.jove.com

ARTICLE AND VIDEO LICENSE AGREEMENT

Title of Article:

GENERATION OF INDUCED PLURIPOTENT STEM CELLS FROM
FROZEN BUFFY COATS USING NON-INTEGRATING EPISOMAL PLASMIDS

Author(s):

ALESSANDRA ROSSINI

Item 1 (check one box): The Author elects to have the Materials be made available (as described at <http://www.jove.com/publish>) via: ☒ Standard Access ☐ Open Access

Item 2 (check one box):

- ☒ The Author is NOT a United States government employee.
- ☐ The Author is a United States government employee and the Materials were prepared in the course of his or her duties as a United States government employee.
- ☐ The Author is a United States government employee but the Materials were NOT prepared in the course of his or her duties as a United States government employee.

ARTICLE AND VIDEO LICENSE AGREEMENT

1. **Defined Terms.** As used in this Article and Video License Agreement, the following terms shall have the following meanings: **"Agreement"** means this Article and Video License Agreement; **"Article"** means the article specified on the last page of this Agreement, including any associated materials such as texts, figures, tables, artwork, abstracts, or summaries contained therein; **"Author"** means the author who is a signatory to this Agreement; **"Collective Work"** means a work, such as a periodical issue, anthology or encyclopedia, in which the Materials in their entirety in unmodified form, along with a number of other contributions, constituting separate and independent works in themselves, are assembled into a collective whole; **"CRC License"** means the Creative Commons Attribution-Non Commercial-No Derivs 3.0 Unported Agreement, the terms and conditions of which can be found at: <http://creativecommons.org/licenses/by-nc-nd/3.0/legalcode>; **"Derivative Work"** means a work based upon the Materials or upon the Materials and other pre-existing works, such as a translation, musical arrangement, dramatization, fictionalization, motion picture version, sound recording, art reproduction, abridgment, condensation, or any other form in which the Materials may be recast, transformed, or adapted; **"Institution"** means the institution, listed on the last page of this Agreement, by which the Author was employed at the time of the creation of the Materials; **"JoVE"** means MyJoVE Corporation, a Massachusetts corporation and the publisher of *The Journal of Visualized Experiments*; **"Materials"** means the Article and / or the Video; **"Parties"** means the Author and JoVE; **"Video"** means any video(s) made by the Author, alone or in conjunction with any other parties, or by JoVE or its affiliates or agents, individually or in collaboration with the Author or any other parties, incorporating all or any portion of the Article, and in which the Author may or may not appear.

2. **Background.** The Author, who is the author of the Article, in order to ensure the dissemination and protection of the Article, desires to have the JoVE publish the Article and create and transmit videos based on the Article. In furtherance of such goals, the Parties desire to memorialize in this Agreement the respective rights of each Party in and to the Article and the Video.

3. **Grant of Rights in Article.** In consideration of JoVE agreeing to publish the Article, the Author hereby grants to JoVE, subject to **Sections 4 and 7** below, the exclusive, royalty-free, perpetual (for the full term of copyright in the Article, including any extensions thereto) license (a) to publish, reproduce, distribute, display and store the Article in all forms, formats and media whether now known or hereafter developed (including without limitation in print, digital and electronic form) throughout the world, (b) to translate the Article into other languages, create adaptations, summaries or extracts of the Article or other Derivative Works (including, without limitation, the Video) or Collective Works based on all or any portion of the Article and exercise all of the rights set forth in (a) above in such translations, adaptations, summaries, extracts, Derivative Works or Collective Works and (c) to license others to do any or all of the above. The foregoing rights may be exercised in all media and formats, whether now known or hereafter devised, and include the right to make such modifications as are technically necessary to exercise the rights in other media and formats. If the "Open Access" box has been checked in **Item 1** above, JoVE and the Author hereby grant to the public all such rights in the Article as provided in, but subject to all limitations and requirements set forth in, the CRC License.

ARTICLE AND VIDEO LICENSE AGREEMENT

4. Retention of Rights in Article. Notwithstanding the exclusive license granted to JoVE in **Section 3** above, the Author shall, with respect to the Article, retain the non-exclusive right to use all or part of the Article for the non-commercial purpose of giving lectures, presentations or teaching classes, and to post a copy of the Article on the Institution's website or the Author's personal website, in each case provided that a link to the Article on the JoVE website is provided and notice of JoVE's copyright in the Article is included. All non-copyright intellectual property rights in and to the Article, such as patent rights, shall remain with the Author.

5. Grant of Rights in Video – Standard Access. This **Section 5** applies if the "Standard Access" box has been checked in **Item 1** above or if no box has been checked in **Item 1** above. In consideration of JoVE agreeing to produce, display or otherwise assist with the Video, the Author hereby acknowledges and agrees that, Subject to **Section 7** below, JoVE is and shall be the sole and exclusive owner of all rights of any nature, including, without limitation, all copyrights, in and to the Video. To the extent that, by law, the Author is deemed, now or at any time in the future, to have any rights of any nature in or to the Video, the Author hereby disclaims all such rights and transfers all such rights to JoVE.

6. Grant of Rights in Video – Open Access. This **Section 6** applies only if the "Open Access" box has been checked in **Item 1** above. In consideration of JoVE agreeing to produce, display or otherwise assist with the Video, the Author hereby grants to JoVE, subject to **Section 7** below, the exclusive, royalty-free, perpetual (for the full term of copyright in the Article, including any extensions thereto) license (a) to publish, reproduce, distribute, display and store the Video in all forms, formats and media whether now known or hereafter developed (including without limitation in print, digital and electronic form) throughout the world, (b) to translate the Video into other languages, create adaptations, summaries or extracts of the Video or other Derivative Works or Collective Works based on all or any portion of the Video and exercise all of the rights set forth in (a) above in such translations, adaptations, summaries, extracts, Derivative Works or Collective Works and (c) to license others to do any or all of the above. The foregoing rights may be exercised in all media and formats, whether now known or hereafter devised, and include the right to make such modifications as are technically necessary to exercise the rights in other media and formats. For any Video to which this Section 6 is applicable, JoVE and the Author hereby grant to the public all such rights in the Video as provided in, but subject to all limitations and requirements set forth in, the CRC License.

7. Government Employees. If the Author is a United States government employee and the Article was prepared in the course of his or her duties as a United States government employee, as indicated in **Item 2** above, and any of the licenses or grants granted by the Author hereunder exceed the scope of the 17 U.S.C. 403, then the rights granted hereunder shall be limited to the maximum rights permitted under such

statute. In such case, all provisions contained herein that are not in conflict with such statute shall remain in full force and effect, and all provisions contained herein that do so conflict shall be deemed to be amended so as to provide to JoVE the maximum rights permissible within such statute.

8. Likeness, Privacy, Personality. The Author hereby grants JoVE the right to use the Author's name, voice, likeness, picture, photograph, image, biography and performance in any way, commercial or otherwise, in connection with the Materials and the sale, promotion and distribution thereof. The Author hereby waives any and all rights he or she may have, relating to his or her appearance in the Video or otherwise relating to the Materials, under all applicable privacy, likeness, personality or similar laws.

9. Author Warranties. The Author represents and warrants that the Article is original, that it has not been published, that the copyright interest is owned by the Author (or, if more than one author is listed at the beginning of this Agreement, by such authors collectively) and has not been assigned, licensed, or otherwise transferred to any other party. The Author represents and warrants that the author(s) listed at the top of this Agreement are the only authors of the Materials. If more than one author is listed at the top of this Agreement and if any such author has not entered into a separate Article and Video License Agreement with JoVE relating to the Materials, the Author represents and warrants that the Author has been authorized by each of the other such authors to execute this Agreement on his or her behalf and to bind him or her with respect to the terms of this Agreement as if each of them had been a party hereto as an Author. The Author warrants that the use, reproduction, distribution, public or private performance or display, and/or modification of all or any portion of the Materials does not and will not violate, infringe and/or misappropriate the patent, trademark, intellectual property or other rights of any third party. The Author represents and warrants that it has and will continue to comply with all government, institutional and other regulations, including, without limitation all institutional, laboratory, hospital, ethical, human and animal treatment, privacy, and all other rules, regulations, laws, procedures or guidelines, applicable to the Materials, and that all research involving human and animal subjects has been approved by the Author's relevant institutional review board.

10. JoVE Discretion. If the Author requests the assistance of JoVE in producing the Video in the Author's facility, the Author shall ensure that the presence of JoVE employees, agents or independent contractors is in accordance with the relevant regulations of the Author's institution. If more than one author is listed at the beginning of this Agreement, JoVE may, in its sole discretion, elect not take any action with respect to the Article until such time as it has received complete, executed Article and Video License Agreements from each such author. JoVE reserves the right, in its absolute and sole discretion and without giving any reason therefore, to accept or decline any work submitted to JoVE. JoVE and its employees, agents and independent contractors shall have

ARTICLE AND VIDEO LICENSE AGREEMENT

full, unfettered access to the facilities of the Author or of the Author's institution as necessary to make the Video, whether actually published or not. JoVE has sole discretion as to the method of making and publishing the Materials, including, without limitation, to all decisions regarding editing, lighting, filming, timing of publication, if any, length, quality, content and the like.

11. **Indemnification.** The Author agrees to indemnify JoVE and/or its successors and assigns from and against any and all claims, costs, and expenses, including attorney's fees, arising out of any breach of any warranty or other representations contained herein. The Author further agrees to indemnify and hold harmless JoVE from and against any and all claims, costs, and expenses, including attorney's fees, resulting from the breach by the Author of any representation or warranty contained herein or from allegations or instances of violation of intellectual property rights, damage to the Author's or the Author's institution's facilities, fraud, libel, defamation, research, equipment, experiments, property damage, personal injury, violations of institutional, laboratory, hospital, ethical, human and animal treatment, privacy or other rules, regulations, laws, procedures or guidelines, liabilities and other losses or damages related in any way to the submission of work to JoVE, making of videos by JoVE, or publication in JoVE or elsewhere by JoVE. The Author shall be responsible for, and shall hold JoVE harmless from, damages caused by lack of sterilization, lack of cleanliness or by contamination due to the making of a video by JoVE its employees, agents or independent contractors. All sterilization, cleanliness or decontamination procedures shall be solely the responsibility of the Author and shall be undertaken at the Author's

expense. All indemnifications provided herein shall include JoVE's attorney's fees and costs related to said losses or damages. Such indemnification and holding harmless shall include such losses or damages incurred by, or in connection with, acts or omissions of JoVE, its employees, agents or independent contractors.

12. **Fees.** To cover the cost incurred for publication, JoVE must receive payment before production and publication the Materials. Payment is due in 21 days of invoice. Should the Materials not be published due to an editorial or production decision, these funds will be returned to the Author. Withdrawal by the Author of any submitted Materials after final peer review approval will result in a US\$1,200 fee to cover pre-production expenses incurred by JoVE. If payment is not received by the completion of filming, production and publication of the Materials will be suspended until payment is received.

13. **Transfer, Governing Law.** This Agreement may be assigned by JoVE and shall inure to the benefits of any of JoVE's successors and assignees. This Agreement shall be governed and construed by the internal laws of the Commonwealth of Massachusetts without giving effect to any conflict of law provision thereunder. This Agreement may be executed in counterparts, each of which shall be deemed an original, but all of which together shall be deemed to be one and the same agreement. A signed copy of this Agreement delivered by facsimile, e-mail or other means of electronic transmission shall be deemed to have the same legal effect as delivery of an original signed copy of this Agreement.

A signed copy of this document must be sent with all new submissions. Only one Agreement required per submission.

CORRESPONDING AUTHOR:

Name: ALESSANDRA ROSSINI
Department: CENTER FOR BIOMEDICINE
Institution: EUROPEAN ACADEMY OF BOZEN (EURAC)
Article Title: GENERATION OF INDUCED PLURIPOTENT STEM CELLS FROM FROZEN BUFFY COATS USING NON-INTEGRATING EPISOMAL PLASMIDS.
Signature: Alemaudio Biondi Date: 10/31/2014

Please submit a signed and dated copy of this license by one of the following three methods:

- 1) Upload a scanned copy of the document as a pdf on the JoVE submission site;
- 2) Fax the document to +1.866.381.2236;
- 3) Mail the document to JoVE / Attn: JoVE Editorial / 1 Alewife Center #200 / Cambridge, MA 02139

For questions, please email submissions@jove.com or call +1.617.945.9051

Response to Editorial and Reviewers' comments

The authors would like to thank the Reviewers for his/her comments to their work and they revised the manuscript JoVE52885R1 "Generation of Induced Pluripotent Stem Cells from Frozen Buffy Coats using non-integrating Episomal Plasmids" according to Editorial and Reviewers' comments. The authors uploaded the revised manuscript with modifications highlighted in red and in "track changes" function.

The text of the manuscript was updated with new comments as indicated below.

Editorial comments

Question 1. Please take this opportunity to thoroughly proofread the manuscript to ensure that there are no spelling or grammar issues. The JoVE editor will not copy-edit your manuscript and any errors in the submitted revision may be present in the published version.

Answer 1. The authors carefully proofread the manuscript and the errors/inaccuracies of the main text have been corrected.

Question 2.

2.2.2 - How long are cells incubated? Overnight?

3.1 - How are the cells collected?

3.3 - Incubate the cells in what? A new 12-well plate?

Answer 2. The authors added the additional details required in the following steps: 2.2.2, 3.1 and 3.3.

Question 3. JoVE reference format requires that DOIs are included, when available, for all references listed in the article. This is helpful for readers to locate the included references and obtain more information. Please note that often DOIs are not listed with PubMed abstracts and as such, may not be properly included when citing directly from PubMed. In these cases, please manually include DOIs in reference information.

Answer 3. The authors added DOIs in the reference list, when DOIs are available.

Reviewer 1#

Question 1.1. Freezing medium (final) composition in sections 1.1.4 and 1.2.9 is different. Please revise and adjust if needed.

Answer 1.1. The authors thank the reviewer for having noticed this mistake and corrected the error in step 1.1.4.

Question 1.2. Please specify cell count or dilution for freezing in steps 1.2.9-10.

Answer 1.2. The authors revised the text in step 1.2.9 in order to clarify the cell dilution in the freezing medium.

Question 1.3. Describe if plates are coated prior to cell plating (they might not be) in step 2.1.6

Answer 1.3. The authors clarified that plates are not coated, as suggested by the reviewer.

Question 1.4. Section 3: Before going into further detail, please describe detachment protocol applied prior to cell collection (also applies to 6.1). Also include plating density or splitting ratio.

Answer 1.4. The authors added more details in section 3 (and step 6.1) in order to address the comment of the reviewer. Specifically, the authors clarified that PBMNCs grow in suspension culture and no detachment protocol was applied, but PBMNCs were collected using a pipette with 1-ml tip.

Question 1.5. Section 4: It is unclear if you maintain/split cells for a number of days (fixed) or until you reach a critical number of cells. Explain in the text.

Answer 1.5. The authors provided details in the text in Section 4, in order to better explain those steps.

Question 1.6. Extended explanation on manual colony passaging might be required in step 7.3.

Answer 1.6. The authors explained more in detail the manual colony passaging in step 7.2-7.3.

Question 1.7. Splitting ratio is not specified in step 7.7. Please add

Answer 1.7. The authors added the splitting ration in step 7.7, as indicated by the reviewer.

Question 1.8. Consider adding details on feeder free culture of iPSCs in step 8.6.1.

Answer 1.8. The authors indicated the catalog number of the commercial medium for iPSC feeder-free culture in the Table of Materials and Reagents and the manufacturer reported in details all the procedure that we followed to culture feeder-free iPSC.

Reviewer 2#

Question 2.1. Peripheral blood mononuclear cells (PBMCs) consist of chiefly of lymphocytes and monocytes. These cells, usually considered primary and end cells, in absence of specific mitogens don't proliferate. The medium used for PBMCs amplification should be quoted or, if directly designed by authors, justified for its composition.

Answer 2.1. As stated by the reviewer, PBMCs are not able to proliferate without specific stimuli. For this reason, the authors used a defined medium previously described in literature to promote and sustain the proliferation of PBMCs, as indicated in the reference list.

Reviewer 3#

Question 3.1. Please cite Haase et al. Cell Stem Cell. 2009 Oct 2;5(4):434-41. First paper describing cord blood-derived hiPSC induction.

Answer 3.1. The authors thank the reviewer for having noticed this missing reference and added this work in the reference list.

Question 3.2. Provide some sentences on the characteristics of the episomal plasmids used in this study already in the introduction.

Answer 3.2. Episomal plasmids used in this work are commercial available (Addgene) and the catalog number for each plasmid is reported in the Table of Materials and Reagents. The manufacturer reported in details all the information and characteristics of the episomal plasmids. Specifically, these plasmids were described for the first time in the following paper by Yamanaka's group as cited in the reference list (Reference 10: Okita, K. et al. A more efficient method to generate integration-free human iPS cells. *Nat. Methods*. 8 (5), 409-412, doi: 10.1038/nmeth.1591 (2011)). Also, these plasmids were already used to reprogram skin fibroblasts in another work, quoted in the reference list (Reference 15: Kim, C. et al. Studying arrhythmogenic right ventricular dysplasia with patient-specific iPSCs. *Nature*. **494** (7435), 105-110, doi: 10.1038/nature11799 (2013)). Of note, the introduction (133-134) has been modified in order to better clarify that the plasmids used in this work have been already used in previous publications and additional episomal plasmid details are added in the method section (step 4.3).

Question 3.3. Provide a brief method sections on:

3.3.1 Plasmid preparation, isolation, quality assessment;

3.3.2 Method to test transfection efficiency via reporter gene constructs;

3.3.3. Method to assess cell viability after electroporation;

3.3.4. Assay to test and proof loss of the episomal plasmids in the generated hiPSC colonies / clones;

3.3.5. Method on how to measures / define reprogramming efficiency and provide expected efficiency data on fresh vs. frozen PMBMCS samples. This is important but currently only briefly mentioned in the discussion

Answer 3.3. Additional information are provided in order to address the following points highlighted by the reviewer:

3.3.1 Purification and isolation of the four plasmids was performed using the plasmid purification kit (Qiagen) listed in the Table of Materials and Reagents. The manufacturer reported in details all the procedure followed by the authors to isolate and check the quality of the purified plasmids used in this work. Specifically, the authors added this comments in the text at the beginning of Section 4 (lines 280-283).

3.3.2. To test the transfection efficiency, the authors added the step 4.8.

3.3.3. To assess cell viability after electroporation, cells were counted before and after electroporation protocol and cell viability was evaluated based on trypan blue exclusion (see in the text the step 4.6).

3.3.4. The authors would thank the reviewer for giving the chance to clarify this point. Indeed, the authors splitted in two different section qRT-PCR for gene expression analysis and qPCR for transgene exclusion, in order to better explain how to test the loss of episomal plasmids (see Sections 8.4-8.5).

3.3.5. The authors would thank the reviewer for giving the possibility to clarify this point and also to give the chance to revise and correct a mistake that the authors missed in the previous submission, regarding the reprogramming efficiency (see the correction in the text at line 674). Specifically, the authors measured the reprogramming efficiency as the percentage of average iPSC colony formation, calculated as number of iPSC colonies/total number of electroporated cells as stated in Section 7 (lines 346-347) (Hasegawa K, Zhang P, Wei Z, Pomeroy JE, Lu W, Pera MF. Comparison of reprogramming efficiency between transduction of reprogramming factors, cell-cell fusion, and cytoplasm fusion. Stem Cells. 2010 Aug;28(8):1338-48. doi: 10.1002/stem.466). Of note, the reprogramming efficiency obtained in the present work is comparable to those reported in previously published works (Reference 14: Chou, B. K. et al. Efficient human iPS cell derivation by a non-integrating plasmid from blood cells with unique epigenetic and gene expression signatures. Cell Res. 21 (3), 518-529, doi: 10.1038/cr.2011.12 (2011) and reference 23: Staerk, J. et al. Reprogramming of human peripheral blood cells to induced pluripotent stem cells. Cell. Stem Cell. 7 (1), 20-24, doi: 10.1016/j.stem.2010.06.002 (2010)). In the present work the reprogramming efficiency did not differ from the two frozen starting materials (see the text at line 674-675), but the authors cannot provide efficiency data on fresh blood PBMCs samples, because only frozen PBMCs were used in this study.

Also, the authors would thank the reviewer for his/her minor comments and changes in the text have been addressed, according reviewer's suggestion.