Dear Author,

This document is divided into a number of sections in which you can add your comments to the video, voiceover, online text, and .pdf. Please be aware that our policy is to do a single complementary revision, so it is critical that all participants in this project offer their comments collectively. In addition, please make sure that your comments are easily interpreted, transparent, and where appropriate, specifically reference a time-point in the video or step in the written section.

Have fun!

Project Name: An *in vitro* Enzymatic Assay to Measure Transcription Inhibition by Gallium(III) and H₃ 5,10,15-tris(pentafluorophenyl)corroles

Date: February 1, 2015

Authors and Affiliations

Please fill in any missing author information not included in the video.

Order	Author	Affiliation	Email

Video Comments:

Please fill in any comments you wish to make in the "comment" column and insert the time-code that references the video at the time of the mistake in the "time-code" column. Also, please make suggestions to correct the mistake. For example it is not enough to say that 0.25 mM Fluo-4 is incorrect. You must suggest the correct concentration. If you need more space to write, please do so below the table. DO NOT ADD CORRECTIONS TO THE VOICEOVER HERE. PLEASE DO THIS IN THE NEXT SECTION.

Time-code		Comment	Suggestion
1.	4:23	The pipetting shows bad	Delete that scene, or
		technique (a drop of blue	substitute with alternate
		solution is on the outside of	footage that does not have
		the pipette, and should not	that excess material on the
		be there).	outside of the pipette tip.
2.	6:43	Graphic appears and fades	Keep the image of the
		out immediately, giving	graph on screen for at least
		little time to process the	2 more seconds (maybe
		information.	longer,
3.	7:00-7:21 (specifically	Sample lanes are	Please highlight the Control
	7:05 and 7:20)	highlighted when	lane when it is mentioned
		mentioned by the voice	in the voiceover (when it is
		talent, but the Control lane	being compared to the
		is never highlighted.	other highlighted bands).
4.	8:52	Mary Tang was responsible	"Schematic Overview by
		for the creation of the art	Mary H. Tang" or
		used in the schematic	"Illustration by Mary H.
		overview. Please include	Tang"
		her in the	
		credits/acknowledgements.	
5.			
6.			
7.			
8.			
9.			
10.			

Audio Comments:

This section is used to specify the changes that need to be made to the voiceover. Please specify time code where each mistake in the voiceover occurs, your comment, the step in the script that needs to be changed, and the corrected text. Please include the entire step from the script in last column with the corrected text in bold. If there is a pronunciation change, please provide a phonetic pronunciation key.

EX:

Restinosis – (Reese-tin-oh-sis)

Time code Comment Cton in Coriet Descripts		Downitton Tout on Comments of		
Time code		Comment	Step in Script	Rewritten Text or Corrected
_			(ex 4.2)	Pronunciation
1.	0:20,	The word		Ko- <i>role</i> (or Kuh- <i>role</i> , but more
	1:10,	"corrole" should		similar to Ko-role)
	1:40	be emphasized on		
		"-role", and not		
		on "cor-"		
2.	2:22-	Omit the	2.4	Thaw all the frozen reagents on ice.
	2:25	sentence		
		following "Thaw		
		all frozen		
		reagents on ice."		
		The sentence		
		following is		
		redundant and		
		can lead to		
		confusion.		
3.	2:57-	The sentence	2.7	Next, remove 4 microliter aliquots
	3:05	"Following the		from each reaction after each hour
		transcriptiontext		and store at minus 20 degrees
		protocol" may be		Celsius, modifying as needed for
		unclear to the		desired time-points [CU-2.7.1].
		viewer why it is		Following the transcription reaction,
		necessary to		purify the RNA from the other
		purify the RNA.		reaction components using the
		We suggest a		RNA spin columns as described in
		sentence change.		the text protocol [MED-2.7.2].
4.	5:21-	We suggest 3.8	3.8, 3.9	3.8. Run the gel for however long is
	5:40	and 3.9		sufficient for significant separation
		voiceovers to be		between potential RNA fragments,
		combined, since		using the migration of dyes to
		they contain		monitor the progress of the

		redundant information. Also, since the DNA/RNA cannot be imaged until using the UV imaging machine, it is clearer to not mention DNA/RNA and instead focus on the dyes, which can be seen.		separation [CU-3.8.1]. 3.9. Turn off the power supply before the dye from the loading buffer has migrated to the end of the gel [MED-over the shoulder- 3.9.1].
5.	6:30	In step 4.3, a 1:4 dilution was relevant to our particular experiment, but may not be for the viewer. We advise changing the sentence to the true goal of obtaining "an optical density less than 1"	4.3	In the case that absorbance is above an optical density of 1, dilute the samples in nuclease-free water to obtain an optical density less than 1 [MED-4.3.1].
6.	7:48	While "0.07 times" is acceptable in written format, it's usually not spoken in that way. 1/14 th or 14-fold is a more common way to say things.	5.4	Three of the four inhibitor-treated transcription reactions yielded less RNA than the control, with Gallium-tpfc-treated DNA transcribing only one-fourtheenth as much RNA as the untreated DNA [LM-5.4.1].
7.		, 5-		
8.				
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12.				
13.				

Text Protocol:

Please use this table to address changes that need to be made to the text. List the step in the text protocol where there is an issue, your comments pertaining to that issue, and how we should resolve it. For drastic changes to the protocol (major structural changes or more than 10 spelling or grammatical mistakes), we will require re-upload of the entire document.

1.	Step in Protocol	Comment	Suggestion
2.	3.7	The goal of the agarose gel electrophoresis is not to separate RNA and DNA, but that any fragments of RNA in a single lane would separate and give a high resolution image. Sorry about this.	Rewrite the following sentence (suggested changes are in bold): Attach the leads so that the DNA will migrate into the gel towards the positive lead. Set the voltage to the desired level, typically 1 to 10 V/cm of gel. Run the gel for however long is sufficient for significant separation between the RNA fragments , using the migration of dyes to monitor the progress of the separation.
3.	3.8	Same reasoning as previous comment (for 3.7).	Rewrite the following sentence (suggested changes are in bold): Turn off the power supply when the dye from the loading buffer has migrated a distance judged sufficient for separation between the RNA fragments .
4.	4.3	In step 4.3, a 1:4 dilution was relevant to our particular experiment, but may not be for the viewer. We advise changing the sentence to the true goal of obtaining "an optical density less than 1"	In the case that absorbance is above 1 O.D., dilute the samples in nuclease-free water to obtain an O.D. less than 1.
5.			

.PDF

Please use this table to address changes that need to be made to the pdf. List the step in the text protocol where there is an issue, your comments pertaining to that issue, and how we should resolve it. For drastic changes to the protocol, we will require re-upload of the entire document.

1.	Step in Protocol	Comment	Suggestion
2.	3.7	The goal of the agarose gel electrophoresis is not to separate RNA and DNA, but that any fragments of RNA in a single lane would separate and give a high resolution image. Sorry about this.	Attach the leads so that the DNA will migrate into the gel towards the positive lead. Set the voltage to the desired level, typically 1 to 10 V/cm of gel. Run the gel for however long is sufficient for significant separation between the RNA fragments, using the migration of dyes to monitor the progress of the separation.
3.	3.8	Same reasoning as previous comment (for 3.7).	Turn off the power supply when the dye from the loading buffer has migrated a distance judged sufficient for separation between the RNA fragments .
4.	4.3	In step 4.3, a 1:4 dilution was relevant to our particular experiment, but may not be for the viewer. We advise changing the sentence to the true goal of obtaining "an optical density less than 1"	In the case that absorbance is above 1 O.D., dilute the samples in nuclease-free water to obtain an O.D. less than 1.
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