

Dear Author,

This document is divided into a number of sections in which you can add your comments to the video, voiceover, online text, and .pdf. Please be aware that our policy is to do a single complementary revision, so it is critical that all participants in this project offer their comments collectively. In addition, please make sure that your comments are easily interpreted, transparent, and where appropriate, specifically reference a time-point in the video or step in the written section.

Have fun!

Project Name: An in vitro Enzymatic Assay to Measure Transcription Inhibition by Gallium(III) and H₃ 5,10,15-tris(pentafluorophenyl)corroles

Date: February 1, 2015

Authors and Affiliations

Please fill in any missing author information not included in the video.

Order	Author	Affiliation	Email

Video Comments:

Please fill in any comments you wish to make in the “comment” column and insert the time-code that references the video at the time of the mistake in the “time-code” column. Also, please make suggestions to correct the mistake. For example it is not enough to say that 0.25 mM Fluo-4 is incorrect. You must suggest the correct concentration. If you need more space to write, please do so below the table. DO NOT ADD CORRECTIONS TO THE VOICEOVER HERE. PLEASE DO THIS IN THE NEXT SECTION.

Time-code		Comment	Suggestion
1.	4:23	The pipetting shows bad technique (a drop of blue solution is on the outside of the pipette, and should not be there).	Delete that scene, or substitute with alternate footage that does not have that excess material on the outside of the pipette tip.
2.	6:43	Graphic appears and fades out immediately, giving little time to process the information.	Keep the image of the graph on screen for at least 2 more seconds (maybe longer,
3.	7:00-7:21 (specifically 7:05 and 7:20)	Sample lanes are highlighted when mentioned by the voice talent, but the Control lane is never highlighted.	Please highlight the Control lane when it is mentioned in the voiceover (when it is being compared to the other highlighted bands).
4.	8:52	Mary Tang was responsible for the creation of the art used in the schematic overview. Please include her in the credits/acknowledgements.	“Schematic Overview by Mary H. Tang” or “Illustration by Mary H. Tang”
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Audio Comments:

This section is used to specify the changes that need to be made to the voiceover. Please specify time code where each mistake in the voiceover occurs, your comment, the step in the script that needs to be changed, and the corrected text. Please include the entire step from the script in last column with the corrected text in bold. If there is a pronunciation change, please provide a phonetic pronunciation key.

Time code		Comment	Step in Script (ex 4.2)	Rewritten Text or Corrected Pronunciation
1.	0:20, 1:10, 1:40	The word “corrole” should be emphasized on “-role”, and not on “cor-”		Ko- <i>role</i> (or Kuh- <i>role</i> , but more similar to Ko- <i>role</i>)
2.	2:22- 2:25	Omit the sentence following “Thaw all frozen reagents on ice.” The sentence following is redundant and can lead to confusion.	2.4	Thaw all the frozen reagents on ice.
3.	2:57- 3:05	The sentence "Following the transcription...text protocol" may be unclear to the viewer <i>why</i> it is necessary to purify the RNA. We suggest a sentence change.	2.7	Next, remove 4 microliter aliquots from each reaction after each hour and store at minus 20 degrees Celsius, modifying as needed for desired time-points [CU-2.7.1]. Following the transcription reaction, purify the RNA from the other reaction components using the RNA spin columns as described in the text protocol [MED-2.7.2].
4.	5:21- 5:40	We suggest 3.8 and 3.9 voiceovers to be combined, since they contain redundant information. Also, since the	3.8, 3.9	3.8. Run the gel for however long is sufficient for significant separation between potential RNA fragments , using the migration of dyes to monitor the progress of the separation [CU-3.8.1]. 3.9. Turn off the power supply before the dye from the loading

		DNA/RNA cannot be imaged until using the UV imaging machine, it is clearer to not mention DNA/RNA and instead focus on the dyes, which can be seen.		buffer has migrated to the end of the gel [MED-over the shoulder-3.9.1].
5.	6:30	In step 4.3, a 1:4 dilution was relevant to our particular experiment, but may not be for the viewer. We advise changing the sentence to the true goal of obtaining “an optical density less than 1”	4.3	In the case that absorbance is above an optical density of 1, dilute the samples in nuclease-free water to obtain an optical density less than 1 [MED-4.3.1].
6.	7:48	While “0.07 times” is acceptable in written format, it’s usually not spoken in that way. 1/14 th or 14-fold is a more common way to say things.	5.4	Three of the four inhibitor-treated transcription reactions yielded less RNA than the control, with Gallium-tpfc-treated DNA transcribing only one-fourteenth as much RNA as the untreated DNA [LM-5.4.1].
7.	3:14-3:24	We’d like to emphasize that care should be taken when working with Ethidium Bromide, as it is extremely toxic.	3.1	To perform agarose gel electrophoresis, prepare Tris Acetate EDTA, or TAE running buffer, and a 1000x stock solution of ethidium bromide and as described in the text protocol [MED-3.1.1]. Always wear appropriate protective equipment when working with ethidium bromide, as it is toxic.
8.				

Text Protocol:

Please use this table to address changes that need to be made to the text. List the step in the text protocol where there is an issue, your comments pertaining to that issue, and how we should resolve it. For drastic changes to the protocol (major structural changes or more than 10 spelling or grammatical mistakes), we will require re-upload of the entire document.

1.	Step in Protocol	Comment	Suggestion
2.	3.7	The goal of the agarose gel electrophoresis is not to separate RNA and DNA, but that any fragments of RNA in a single lane would separate and give a high resolution image. Sorry about this.	Rewrite the following sentence (suggested changes are in bold): Attach the leads so that the DNA will migrate into the gel towards the positive lead. Set the voltage to the desired level, typically 1 to 10 V/cm of gel. Run the gel for however long is sufficient for significant separation between the RNA fragments , using the migration of dyes to monitor the progress of the separation.
3.	3.8	Same reasoning as previous comment (for 3.7).	Rewrite the following sentence (suggested changes are in bold): Turn off the power supply when the dye from the loading buffer has migrated a distance judged sufficient for separation between the RNA fragments .
4.	4.3	In step 4.3, a 1:4 dilution was relevant to our particular experiment, but may not be for the viewer. We advise changing the sentence to the true goal of obtaining "an optical density less than 1"	In the case that absorbance is above 1 O.D., dilute the samples in nuclease-free water to obtain an O.D. less than 1.
5.			

.PDF

Please use this table to address changes that need to be made to the pdf. List the step in the text protocol where there is an issue, your comments pertaining to that issue, and how we should resolve it. For drastic changes to the protocol, we will require re-upload of the entire document.

1.	Step in Protocol	Comment	Suggestion
2.	3.7	The goal of the agarose gel electrophoresis is not to separate RNA and DNA, but that any fragments of RNA in a single lane would separate and give a high resolution image. Sorry about this.	Attach the leads so that the DNA will migrate into the gel towards the positive lead. Set the voltage to the desired level, typically 1 to 10 V/cm of gel. Run the gel for however long is sufficient for significant separation between the RNA fragments , using the migration of dyes to monitor the progress of the separation.
3.	3.8	Same reasoning as previous comment (for 3.7).	Turn off the power supply when the dye from the loading buffer has migrated a distance judged sufficient for separation between the RNA fragments .
4.	4.3	In step 4.3, a 1:4 dilution was relevant to our particular experiment, but may not be for the viewer. We advise changing the sentence to the true goal of obtaining "an optical density less than 1"	In the case that absorbance is above 1 O.D., dilute the samples in nuclease-free water to obtain an O.D. less than 1.
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